Package 'BCBCSF'

May 6, 2026

Version 1.0-2

Date 2026-03-27

Title Bias-Corrected Bayesian Classification with Selected Features

Depends R (>= 2.13.1), abind

Description Fully Bayesian Classification with a subset of high-dimensional features, such as expression levels of genes. The data are modeled with a hierarchical Bayesian models using heavy-tailed t distributions as priors. When a large number of features are available, one may like to select only a subset of features to use, typically those features strongly correlated with the response in training cases. Such a feature selection procedure is however invalid since the relationship between the response and the features has be exaggerated by feature selection. This package provides a way to avoid this bias and yield better-calibrated predictions for future cases when one uses F-statistic to select features.

License GPL (>= 2)

URL https://www.r-project.org, https://longhaisk.github.io/

RoxygenNote 7.3.2

NeedsCompilation yes

Author Longhai Li [aut, cre]

Maintainer Longhai Li <longhai.li@usask.ca>

Repository CRAN

Date/Publication 2026-03-30 14:20:10 UTC

Contents

d1:bcbcsfexamples *Examples of fitting models, predicting class labels, evaluating prediction, and analyzing fitting results*

Description

These examples demonstrate how to use BCBCSF package. They use all prior and Markov chain sampling settings by default (except no_rmc as noted below). The methods for setting others can be found from documents for specific functions. However, the default settings may work well for a wide range of gene expression data.

References

Li, L. (2012), Bias-corrected Hierarchical Bayesian Classification with a Selected Subset of High-dimensional Features, *Journal of American Statistical Association*,107:497,120-134

See Also

bcbcsf_fitpred, bcbcsf_pred, cross_vld, eval_pred, reload_fit_bcbcsf, bcbcsf_sumfit, bcbcsf_plotsumfit

Examples

```
##\dontrun{
## load lymphoma microarray data
data (lymphoma)

## select some cases as testing data set
ts <- c (sort(sample (1:42,5)), 43:44, 61:62)

## training data
X_tr <- lymph.X[-ts,]
y_tr <- lymph.y[-ts]
## test data
X_ts <- lymph.X[ts,]
y_ts <- lymph.y[ts]

#############################################################################
######################### training and prediction ###########################
#############################################################################

## To comply with CRAN policies, we use R's temporary directory for saving files
my_prefix <- file.path(tempdir(), "bcbcsf_")

## fitting training data with top features selected by F-statistic
out_fit <- bcbcsf_fitpred (X_tr = X_tr, y_tr = y_tr, nos_fsel = c(20, 50),
                           no_rmc = 10, fit_bcbcsf_filepre = my_prefix)
## note 1: if 'X_ts' is given above, prediction is made after fitting
## note 2: no_rmc = 100 is too small, omit it and use the default
```

```
## predicting class labels of test cases
out_pred <- bcbcsf_pred (X_ts = X_ts, out_fit = out_fit)

## evaluate prediction given true labels
eval_pred (out_pred = out_pred, y_ts = y_ts)

#############################################################################
###################### visualizing prediction results ######################
#############################################################################
## reload one bcbcsf fit result from hardrive
fit_bcbcsf <- reload_fit_bcbcsf (out_fit$fitfiles[1])
## the fitting result for no_fsel = 50 can be retrieved directly from
## out_fit:
fit_bcbcsf_fsel50 <- out_fit$fit_bcbcsf
## summarize the fitting result
sum_fit <- bcbcsf_sumfit (fit_bcbcsf)

## visualize fitting result
bcbcsf_plotsumfit (sum_fit)

#############################################################################
############################ cross validation ###############################
#############################################################################

cv_prefix <- file.path(tempdir(), "cv_bcbcsf_")

## doing cross validation with bcbcsf_fitpred on lymphoma data
cv_pred <- cross_vld (
       ##################### classifier, data, and fold ###################
       fitpred_func = bcbcsf_fitpred, X = lymph.X, y = lymph.y, nfold = 2,
       ################ all other arguments passed classifier ############
       nos_fsel = c(10,12), no_rmc = 20, fit_bcbcsf_filepre = cv_prefix)
## note: no_rmc = 100 is too small, omit it and use the default in practice

## evaluate prediction given true labels
eval_pred (out_pred = cv_pred, y_ts = lymph.y)

## warning: this function is slow if nfold is large; if you have a
## computer cluster, you better parallel the cross validation folds.
##}
```

`d2:fitpred` *Functions for fitting models with MCMC, predicting class labels of test cases, and finding predictive probabilities with cross-validation*

Description

`bcbcsf_fitpred` trains models with Gibbs sampling for each number of retained features. The results are saved in files. This function also makes predictions for test cases if they are provided.

bcbcsf_pred uses the posterior samples saved by bcbcsf_fitpred to predict the class labels of test cases. Prediction results are an array of predictive probabilities array_probs_pred, whose rows for test cases, columns for classes, and the 3rd dimension for different numbers of retained features.

cross_vld uses cross-validation to obtain predictive probabilities for all cases of a data set. This generic function can be used with bcbcsf_fitpred and other classifiers.

Usage

```
bcbcsf_fitpred (
  ## arguments specifying info of data sets
  X_tr, y_tr, nos_fsel = ncol (X_tr),
  X_ts = NULL,  standardize = FALSE, rankf = FALSE,
  ## arguments for prediction
  burn = NULL, thin = 1, offset_sdxj = 0.5,
  ## arguments for Markov chain sampling
  no_rmc = 1000, no_imc = 5, no_mhwmux = 10,
  fit_bcbcsf_filepre = NULL,
  ## arguments specifying priors for parameters and hyerparameters
  w0_mu = 0.05, alpha0_mu = 0.5, alpha1_mu = 3,
  w0_x  = 1.00, alpha0_x  = 0.5, alpha1_x  = 10,
  w0_nu = 0.05, alpha0_nu = 0.5, prior_psi = NULL,
  ## arguments for metropolis sampling for wmu, wx
  stepadj_mhwmux = 1, diag_mhwmux = FALSE,
  ## arguments for computing adjustment factor
  bcor = 1, cut_qf = exp (-10), cut_dpoi = exp (-10), nos_sim = 1000,
  ## whether look at progress
  monitor = TRUE)

bcbcsf_pred (X_ts, out_fit, burn = NULL, thin = 1, offset_sdxj = 0.5)

cross_vld (X, y, nfold = 10, folds = NULL,
           fitpred_func = bcbcsf_fitpred,  ...)
```

Arguments

X_tr, X_ts, X	matrices containing gene expression data; rows should be for the cases, and columns for different genes; X_tr are training data, X_ts are test data or future data for which prediction are needed, X are a data set used for cross-validation.
y_tr, y	class labels in training or test data set, or just a data set.
nos_fsel	a vector of numbers of features to be retained.
burn, thin	burn of Markov chain (super)iterations will be discarded for prediction, and only every thinth are used; by default, 20% of (super)iterations are burned, and thin=1.
offset_sdxj	a value between 0 and 1; 100*offset_sdxj% quantile of the samples of all standard deviations $\sqrt{w_j^x}$ is added to the all standard deviations; this is to remedy the non-normality in real gene expression data sets, and especially offset some very small standard deviations; by default, median is used.

`no_rmc`, `no_imc` `no_rmc` of super Markov chain transitions are run, with `no_imc` Markov chain iterations for each; only the last state of each super transition is saved.

`fit_bcbcsf_filepre`
: a string added to the names of files saving Markov chain fitting results; the actual file names contain also the data dimension and number of retained features. By default, this is set to `NULL`, meaning no fitting files will be created, and `bcbcsf_fitpred` returns only the fitting result corresponding to the last number of retained features in `nos_fsel`. If you wish to save the files, provide a string prefix (e.g., "my_fit_").

`w0_mu`, `alpha0_mu`, `alpha1_mu`, `w0_x`, `alpha0_x`, `alpha1_x`, `w0_nu`, `alpha0_nu`
: settings of priors for means and variances of genes; they are denoted by w_0^μ, α_1^μ, α_1^μ,w_0^x,α_0^x,α_1^x,w_0^ν,α_0^ν in the reference.

`prior_psi`
: a vector of length the number of classes, specifying the Dirichlet prior distribution for probabilities of classes; it is denoted by $c_{1:G}$ in the reference; by default, they are all equal to 1.

`no_mhwmux`, `stepadj_mhwmux`, `diag_mhwmux`
: arguments specifying Metropolis sampling for $\log(w^\mu)$ and $\log(w^x)$; respectively the number of iterations, stepsize adjustment, and an indicator representing whether one wants to pause and look into this sampling.

`bcor`
: taking value 0 or 1, indicating whether bias-correction is to be applied.

`cut_qf`, `cut_dpoi`, `nos_sim`
: arguments specifying approximation of adjustment factor; `cut_qf` is f_ℓ in the reference, `cut_dpoi` is the threshold below which Poisson probabilities are omitted, `nos_sim` is the number of random Λ.

`nfold`, `folds`
: `folds` should be a list of test cases for different folds; if `folds` is NULL (by default), `folds` will be generated by the software, with `nfold` is set to the smaller value of the given value and the smallest number of cases in all classes.

`out_fit`
: a list returned by `bcbcsf_fitpred`, which are used to make prediction for test cases.

`standardize`
: if it is set to TRUE, the original gene expression values are centralized and divided by the pooled standard deviation; by default, it is FALSE.

`rankf`
: if it is set to TRUE, the original features will be re-ordered by F-statistic; by default, it is FALSE.

`monitor`
: if it is set to TRUE, progress of fitting is shown on screen

`fitpred_func`
: an R function that can fit with training data, and predict for test data; the arguments of `fitpred_func` must include `X_tr`, `y_tr`, `X_ts`, and the outputs of `fitpred_func` must include `array_probs_pred`

`...`
: arguments passed to classifier `fitpred_func`

Value

`nos_fsel`
: a vector of numbers of features retained.

`fitfiles`
: a string vector of length `nos_fsel`, each saving file name of Markov chain fitting result for a number of retained features in `nos_fsel`; the `fitfiles` returned by `cross_vld` is for the training in the last fold.

`cv_fitfiles` a list of strings returned by `cross_vld`, containing the names of all files saved across all folds. This is only populated if `fit_bcbcsf_filepre` is not `NULL`.

`array_probs_pred` an array of predictive probabilities, whose rows for test cases, columns for classes, and the 3rd dimension for different numbers of retained features.

`fit_bcbcsf` a list of Markov chain sampling results from the fitting with number of retained features equal to the last number in `nos_fsel`. Note that, the fitting results for other numbers (including the last one) of retained feature are saved in harddrive files if `fit_bcbcsf_filepre` isn't empty, and can be retrieved using function `reload_fit_bcbcsf`. Particularly, the list component of `fit_bcbcsf` has `fsel` saving the indice of features selected by F-statistic.

Examples

```
## Generate a small simulated dataset
set.seed(1)
X <- matrix(rnorm(50 * 20), nrow = 50, ncol = 20)
y <- c(rep(1, 25), rep(2, 25))

## Fit and predict with bcbcsf_fitpred (no files saved to disk by default)

out_fit <- bcbcsf_fitpred(X_tr = X[1:40, ], y_tr = y[1:40], no_rmc=20,
                          X_ts = X[41:50, ], nos_fsel = c(5, 10))

## Cross-validation (no files saved to disk by default)
cv_res <- cross_vld(X = X, y = y, nfold = 3, no_rmc = 20, nos_fsel = c(5, 10))
```

`d3:evalpred` *A function for evaluating arrays of predictive probabilities with the true class labels of test cases*

Description

This function is used to find error rate, amlp, loss and predictive probabilities at true labels.

Usage

```
eval_pred (out_pred, y_ts, Mloss = NULL)
```

Arguments

`out_pred` a list returned by function `bcbcsf_fitpred` with `X_ts` given, or `bcbcsf_pred`, or by `cross_vld`.

`y_ts` a vector of true class labels.

`Mloss` a matrix indicting loss function, with element m_{ij} saving the losss from predicting class i with class label j

; by default, it is NULL.

Value

`probs_at_truelabels`
: a matrix of predictive probabilities at true labels, with rows for cases, and columns for different numbers of retained features

`summary`
: a data frame, with rows for different numbers of retained features, and columns: `Error.Rate`: fraction of cases misclassified with fair threshold, and `AMLP`: minus average log probabilities at true labels, often called "deviation", and `Loss` (if `Mloss` is given): average loss.

---

d4:analyzefit *Functions for analyzing and visualizing a BCBCSF fitting result*

Description

These functions are used to look at the fitting results, especially plot the gene signals.

Usage

```
reload_fit_bcbcsf (fit_bcbcsf_afile)

bcbcsf_sumfit (fit_bcbcsf = NULL, fit_bcbcsf_afile = NULL,
               burn = NULL,  thin = 1)

bcbcsf_plotsumfit (sum_fit)
```

Arguments

`fit_bcbcsf_afile`
: a string of name of a file saving a Markov chain fitting result; it can be found from the value `fitfiles` of function `bcbcsf_fitpred`.

`fit_bcbcsf`
: a list of Markov chain fitting result, returned by function `reload_fit_bcbcsf` and `bcbcsf_fitpred`; if it is NULL, it will be retrieved by running `reload_fit_bcbcsf` with value in `fit_bcbcsf_afile`.

`burn, thin`
: `burn` of Markov chain (super)iterations will be discarded (burned) for evaluation, and only every `thin`th are used; by default, 20% of (super)iterations are burned, and `thin`=1.

`sum_fit`
: a list returned by function `bcbcsf_sumfit`

Value

`reload_fit_bcbcsf` returns a list of Markov chain fitting results, including how to do feature selection and data preprocessing.

`bcbcsf_sumfit` returns a list of point estimates of means and variances.

`bcbcsf_plotsumfit` returns nothing; it plots the normalized means (for each gene, original expression means substracted by their means and divided by the common standard deviation), and overall signals (Euclid distance of normalized means) for the selected features.

d5:lymphoma *Lymphoma Microarray Data*

Description

This is one of the microarray data sets used to demonstrate BCBCSF in the reference article. Information about this data set can be found from the reference.

Usage

```
data(lymphoma)
```

Value

lymph.X	a matrix of gene expression data for p = 4026 genes on n = 62 cases in G=3 classes
lymph.y	a vector of class labels coded by 1,2,3.

Index