

# Package ‘DTAT’

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**Title** Dose Titration Algorithm Tuning

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**Imports** km.ci, pomp, Hmisc, data.table, dplyr, r2d3, shiny, jsonlite, methods

**Suggests** knitr, rmarkdown, lattice, latticeExtra, widgetframe, tidyr, RColorBrewer, invgamma, zipfR, rms

**Description** Dose Titration Algorithm Tuning (DTAT) is a methodologic framework allowing dose individualization to be conceived as a continuous learning process that begins in early-phase clinical trials and continues throughout drug development, on into clinical practice. This package includes code that researchers may use to reproduce or extend key results of the DTAT research programme, plus tools for trialists to design and simulate a '3+3/PC' dose-finding study. Please see Norris (2017a) <[doi:10.12688/f1000research.10624.3](https://doi.org/10.12688/f1000research.10624.3)> and Norris (2017c) <[doi:10.1101/240846](https://doi.org/10.1101/240846)>.

**URL** <https://precisionmethods.guru/>

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## Contents

as_d3_data,DE-method . . . . .	2
DE-class . . . . .	3
de.bioRxiv.240846 . . . . .	3
dose.survfit . . . . .	4
dose.survival . . . . .	5
ds.curve . . . . .	6
dtat1000 . . . . .	7
newton.raphson . . . . .	9
Onoue.Friberg . . . . .	10
plot,DE,missing-method . . . . .	12
runDTATapp . . . . .	13
scaled . . . . .	14
seq.function . . . . .	14
sim . . . . .	15
titrate . . . . .	16
titration . . . . .	17
<b>Index</b>	<b>19</b>

---

as\_d3\_data,DE-method    *Convert a DE object to JSON*

---

### Description

Convert a DE object to JSON

### Usage

```
## S4 method for signature 'DE'
as_d3_data(x, ...)
```

### Arguments

x	An object of class DE
...	Unused.

DE-class

*An S4 class for simulating dose-titration study designs***Description**

An S4 class for simulating dose-titration study designs

**Slots**

`doses` A numeric vector of prospectively-determined discrete doses to trial.

`units` A string indicating dose units, e.g. "mg/kg".

`MTDi` A numeric vector of optimal doses for simulated study participants. Optionally a call to an `r<distribution>(...)` function which may be parsed to calculate the `mtd_quantiles` slot.

`mtd_quantiles` A numeric vector of quantiles of the distribution from which the `MTDi` slot was simulated. Intended mainly to support visualization of this distribution, e.g. as an transparent overlay on the dose-survival plot. NULL in case `MTDi` is provided verbatim.

`fractol` A numeric vector of probabilities for the simulated `MTDi` slot. Intended mainly to support visualization, e.g. plotting of 'MTD pointers' on the interactive dose-survival plot.

`data` A data.frame with columns:

- `id` Participant identifier
- `period` DLT assessment period, numbered consecutively from 1
- `dose` Dose level, numbered consecutively starting from 1
- `dlt` A logical indicator: did this this participant experience a DLT during this period?

`stop_esc` integer Period in which 'stop rule' was triggered

`ds_conf_level` numeric Confidence level for confidence band around Kaplan-Meier estimate of the dose-survival curve.

`dose_drop_threshold` numeric Threshold for triggering the 'bypass rule'.

`stop_esc_under` numeric Threshold for triggering the 'stop rule'.

`undo_esc_under` numeric Threshold for triggering the 'rollback rule'.

de.bioRxiv.240846

*Simulated '3+3/PC' dose-titration study from bioRxiv paper no. 240846***Description**

This is a length-10 list of data frames, summarizing the simulated trial from this paper, at the end of periods 1, 2, ..., 10. This structure reflects an awkward S3 implementation that package DTAT v0.3 reimplemented using S4. This data set is retained to support regression tests.

**Format**

A length-10 list of data frames, each with the following columns:

**id** Participant identifier

**period** DLT assessment period, numbered consecutively from 1

**dose** Dose level, numbered consecutively starting from 1

**dlr** A logical indicator: did this this participant experience a DLT during this period?

**Details**

A `stop_esc` attribute is attached to data frames in this list, indicating when escalation stopped during the simulated trial.

**References**

Norris DC. Precautionary Coherence Unravels Dose Escalation Designs. *bioRxiv*. December 2017:240846. doi:10.1101/240846. <https://www.biorxiv.org/content/10.1101/240846v1>

**Examples**

```
data(de.bioRxiv.240846)
# Demonstrate that the new S4 3+3/PC implementation reproduces the
# simulated trial from the paper:
set.seed(2017)
CV <- 0.7; mean_mtd <- 1.0
shape <- CV^-2; scale <- mean_mtd/shape
trial <- new("DE", doses=0.25 * 1.4^(0:6),
            MTDi=rgamma(24, shape=shape, scale=scale),
            units="mg")
trial <- titration(trial, periods=10)
stopifnot(all(trial@data == de.bioRxiv.240846[[10]]))
stopifnot(trial@stop_esc == attr(de.bioRxiv.240846[[10]], 'stop_esc'))
```

---

dose.survfit

*Calculate a dose-survival curve from a dose titration study, adding a confidence band*

---

**Description**

The 'dose-survival curve' is nothing other than an empirical cumulative distribution for MTDi in the sampled population. The term 'survival' is suggested in part by our application of the Kaplan-Meier estimator to interval-censored toxicity information.

**Usage**

```
dose.survfit(de, method = "rothman", avoid.degeneracy = TRUE, conf.level = 0.8)
```

**Arguments**

de	A dose titration experiment like the data slot of class <a href="#">DE</a>
method	The method to be used by <a href="#">km.ci</a> when calculating CI
avoid.degeneracy	When TRUE, this parameter directs the function to introduce artificial events into the dose titration experiment, to avoid degeneracies at the lower and upper ends of the dose-survival curve.
conf.level	Confidence level for KM confidence band.

**Details**

TODO: Describe details of degeneracy avoidance, once these have stabilized.

**Value**

An object of class `survfit`.

**Author(s)**

David C. Norris

**See Also**

[dose.survival](#), [km.ci](#)

**Examples**

```
CV <- 0.7; mean_mtd <- 1.0
shape <- CV^-2; scale <- mean_mtd/shape
trial <- new("DE", doses=0.25 * 1.4^(0:6),
            MTDi=rgamma(24, shape=shape, scale=scale),
            units="mg")
trial <- titration(trial, periods=10)
sf <- dose.survfit(trial@data)
summary(sf)
```

---

dose.survival

*Extract interval-censored dose tolerance data from a dose titration study*

---

**Description**

Constructs a [Surv](#) object from a dose-escalation experiment, using interval-censoring constructs of type='interval2'.

**Usage**

```
dose.survival(de)
```

**Arguments**

de                    A data frame describing a dose-titration study

**Value**

A Surv object of type='interval2'

**Author(s)**

David C. Norris

**See Also**

[dose.survfit](#)

**Examples**

```
CV <- 0.7; mean_mtd <- 1.0
shape <- CV^-2; scale <- mean_mtd/shape
trial <- new("DE", doses=0.25 * 1.4^(0:6),
            MTDi=rgamma(24, shape=shape, scale=scale),
            units="mg")
trial <- titration(trial, periods=10)
dose.survival(trial@data)
```

---

ds.curve

*Extract the dose-survival curve, with its upper and lower confidence band limits*

---

**Description**

This utility function simply makes the results of `dose.survfit` available in the convenient form of a list.

**Usage**

```
ds.curve(de, ...)
```

**Arguments**

de                    A data frame describing a dose-titration study.  
 ...                   Passed through to function `dose.survfit`

**Value**

A list with components `surv`, `upper` and `lower`, each containing a vector that can be indexed by dose level.

**Author(s)**

David C. Norris

**See Also**

[dose.survfit](#)

**Examples**

```
CV <- 0.7; mean_mtd <- 1.0
shape <- CV^-2; scale <- mean_mtd/shape
trial <- new("DE", doses=0.25 * 1.4^(0:6),
            MTDi=rgamma(24, shape=shape, scale=scale),
            units="mg")
trial <- titration(trial, periods=10)
ds.curve(trial@data)
```

---

dtat1000

*Precomputed neutrophil-guided chemotherapy dose titration for 1000 simulated subjects.*

---

**Description**

This dataset is provided to support fast reproduction of a forthcoming pharmacoeconomic paper that includes examination of the empirical distribution of MTDi in N=1000 simulated subjects.

**Format**

A data frame showing end-of-cycle state of neutrophil-guided dose titration for 1000 simulated subjects, across 10 cycles of chemotherapy.

**cycle** Cycle number 1..10

**id** Subject identifiers; an ordered factor with levels `id1 < ... < id1000`

**Cc** Central-compartment drug concentration

**Cp** Peripheral-compartment drug concentration

**ProI** Progenitor cells in proliferating compartment of Friberg et al. (2002) model

**Tx.1** Transit compartment 1

**Tx.2** Transit compartment 1

**Tx.3** Transit compartment 1

**Circ** Concentration (cells/mm<sup>3</sup>) of circulating neutrophils

**dose** Dose of 1-hour infusion administered this cycle

**CircMin** Neutrophil nadir (cells/mm<sup>3</sup>)

**tNadir** Time (days) of neutrophil nadir

**scaled.dose** Fourth root of dose

**time** Time (weeks) of dose administration

## Details

Running the examples interactively, you can verify the reproducibility of this dataset. (That demo is included in a `donttest` block to spare the CRAN servers.)

## References

1. Norris DC. Dose Titration Algorithm Tuning (DTAT) should supersede ‘the’ Maximum Tolerated Dose (MTD) in oncology dose-finding trials. *F1000Research*. 2017;6:112. doi:10.12688/f1000research.10624.3. <https://f1000research.com/articles/6-112/v3>
2. Norris DC. Costing ‘the’ MTD. *bioRxiv*. August 2017:150821. doi:10.1101/150821. <https://www.biorxiv.org/content/10.1101/150821v3>

## Examples

```
data(dtat1000)
# 1. Extract the N final doses, assuming convergence by the tenth course
MTD_i <- with(dtat1000, dose[time==27])
MTD_i <- MTD_i[MTD_i < 5000] # Exclude few outliers
# 2. Do a kernel density plot
library(Hmisc)
library(latticeExtra)
hist <- histogram(~MTD_i, breaks=c(0,100,200,300,400,600,900,1500,2500,4000,5000)
, xlab=expression(MTD[i]))
approx <- data.frame(mtd_i=seq(0, 5000, 10))
approx <- upData(approx,
gamma = dgamma(mtd_i, shape=1.75, scale=200))
dist <- xyplot(gamma ~ mtd_i, data=approx, type='l', col='black', lwd=2)
library(grid)
hist + dist
grid.text(expression(MTD[i] %~%
paste("Gamma(", alpha==1.75, ", ", ", beta==1/200,")"))
, x=unit(0.5,"npc")
, y=unit(0.75,"npc")
)
## A very long repro, which a user of this package may well wish to verify
## by running the examples interactively, although it takes many minutes
## to compute. (Enclosed in a dontest block to avoid overburdening CRAN.)

# Demonstrate close reproduction of original titration (the titration takes many minutes!)
set.seed(2016)
library(pomp)
Onoue.Friberg(N=1000)
# This titration may take an hour to run ...
```

```

chemo <- titrate(doserange = c(50, 3000),
                dta=newton.Raphson(dose1 = 100,
                                   omega = 0.75,
                                   slope1 = -2.0,
                                   slopeU = -0.2)
                )

dtat1k <- upData(chemo$course
                , time = 3*(cycle-1)
                , labels = c(time="Time")
                , units = c(time="weeks")
                , print = FALSE)

c10dose1k <- subset(dtat1k, cycle==10)$scaled.dose
c10dose1000 <- subset(dtat1000, cycle==10)$scaled.dose
stopifnot(0.999 < cor(c10dose1k, c10dose1000))

```

---

newton.Raphson	<i>A dose titration algorithm (DTA) 'factory' based on the Newton-Raphson heuristic</i>
----------------	-----------------------------------------------------------------------------------------

---

## Description

This higher-order ('factory') function produces a simple dose titration algorithm for neutrophil-guided chemotherapy dosing.

## Usage

```
newton.Raphson(dose1, omega, slope1, slopeU)
```

## Arguments

dose1	The starting dose for titration
omega	A relaxation parameter used to moderate dose increments
slope1	Dose-response slope assumed prior to 2nd measured neutrophil nadir
slopeU	Upper bound imposed on slope estimates

## Details

This function manifests the core concept of Dose Titration Algorithm Tuning by delivering an objectively realized 'DTA'. It therefore enables a variety of DTAs to be implemented and compared.

## Value

A dose titration function that advises dose for next cycle of chemotherapy.

**Author(s)**

David C. Norris

**See Also**[titrate](#)


---

Onoue.Friberg	<i>POMP PK/PD model for docetaxel, combining Onoue et al (2016) with Friberg et al (2002)</i>
---------------	-----------------------------------------------------------------------------------------------

---

**Description**

This function produces a POMP model combining docetaxel pharmacokinetics (PK) drawn from Table 2 of Onoue et al (2016) with myelosuppression dynamics drawn from Friberg et al (2002). This model enables simulation of neutrophil-guided dose titration of docetaxel, as done in Norris (2017).

**Usage**

```
Onoue.Friberg(
  N,
  cycle.length.days = 21,
  data = data.frame(time = c(seq(0, 1.95, 0.05), seq(2, cycle.length.days * 24, 1)), y =
    NA),
  delta.t = 0.1
)
```

**Arguments**

N	Size of simulated population.
cycle.length.days	Duration (in days) of chemotherapy cycle to be simulated.
data	Passed through as the data argument of the pomp constructor.
delta.t	Time-step (in hours) of pomp's euler plug-in.

**Value**

No value is returned; rather, the function sets global variables in package environment `DTAT::sim`.

**Author(s)**

David C. Norris

## References

1. Onoue H, Yano I, Tanaka A, Itohara K, Hanai A, Ishiguro H, et al. Significant effect of age on docetaxel pharmacokinetics in Japanese female breast cancer patients by using the population modeling approach. *Eur J Clin Pharmacol*. 2016 Jun;72(6):703-10. doi:10.1007/s00228-016-2031-3.
2. Friberg LE, Henningsson A, Maas H, Nguyen L, Karlsson MO. Model of chemotherapy-induced myelosuppression with parameter consistency across drugs. *J Clin Oncol*. 2002 Dec 15;20(24):4713-21. doi:10.1200/JCO.2002.02.140.
3. Norris DC. Dose Titration Algorithm Tuning (DTAT) should supersede ‘the’ Maximum Tolerated Dose (MTD) in oncology dose-finding trials. *F1000Research*. 2017;6:112. doi:10.12688/f1000research.10624.3. <https://f1000research.com/articles/6-112/v3>

## See Also

[pomp](#), [sim](#)

## Examples

```
# Reproduce the sim$pkpd model and sim$pop population from reference #3:
library(pomp)
Onoue.Friberg(N=25)
sim$pop # NB: this differs from pop of original paper...

# Whereas the present version of Onoue.Friberg() draws simulated populations
# using pomp::rprior(), to reproduce the original F1000Research paper [3] we
# re-draw sim$pop as originally & prosaically done (see https://osf.io/vwnqz/):
set.seed(2016)
N <- 25
dtx.mm <- 0.808 # molar mass (mg/μM) of docetaxel
sim$pop$Circ0 <- rlnorm(N, meanlog=log(5050), sdlog=0.42) # units=cells/mm^3
sim$pop$MTT <- rlnorm(N, meanlog=log(89.3), sdlog=0.16) # mean transit time
sim$pop$gamma <- rlnorm(N, meanlog=log(0.163), sdlog=0.039) # feedback factor
sim$pop$Emax <- rlnorm(N, meanlog=log(83.9), sdlog=0.33)
sim$pop$EC50 <- rlnorm(N, meanlog=log(7.17*dtx.mm), sdlog=0.50)
# PK params from 2-compartment docetaxel model of Onoue et al (2016)
sim$pop$CL <- rlnorm(N, meanlog=log(32.6), sdlog=0.295)
sim$pop$Q <- rlnorm(N, meanlog=log(5.34), sdlog=0.551)
sim$pop$Vc <- rlnorm(N, meanlog=log(5.77), sdlog=0.1) # Onoue gives no CV% for V1
sim$pop$Vp <- rlnorm(N, meanlog=log(11.0), sdlog=0.598) # Called 'V2' in Onoue
sim$pop$kTR=4/sim$pop$MTT

# Now we run the sim$pkpd model, separately for each of N simulated individuals:
allout <- data.frame() # accumulator for N individual ODE solutions
for (id in 1:sim$N) {
  out <- trajectory(sim$pkpd,
    params=c(sim$pop[sim$pop$id==id, -which(names(sim$pop) %in% c('id', 'MTT'))]
      , sigma=0.05, dose=100, duration=1),
    format="data.frame")
  # drop 'traj' and shift 'time' to first column
  out <- out[,c('time', setdiff(colnames(out), c('time', 'traj')))]
  out$id <- paste("id", id, sep="")
}
```

```

  allout <- rbind(allout, out)
}

library(Hmisc)
allout <- upData(allout
  , id = ordered(id, levels=paste("id",1:sim$N,sep=""))
  , units=c(Prol="cells/mm^3", Tx.1="cells/mm^3",
    Tx.2="cells/mm^3", Tx.3="cells/mm^3",
    Circ="cells/mm^3",
    Cc="ng/mL", Cp="ng/mL",
    time="hours"), print=FALSE)

library(tidyr)
cout <- gather(allout, key="Series", value="Concentration"
  , Cc, Cp
  , factor_key = TRUE)

label(cout$Concentration) <- "Drug Concentration"

# Figure 1 from reference [3]:
library(RColorBrewer)
xYplot(Concentration ~ time | id, group=Series
  , data=cout, subset=time<6
  , layout=c(5,NA)
  , type='l', as.table=TRUE
  , label.curves=FALSE
  , par.settings=list(superpose.line=list(lwd=2,col=brewer.pal(4,"PRGn"))[c(1,4)]))
  , scales=list(y=list(log=TRUE, lim=c(10^-3,10^1)))
  , auto.key=list(points=FALSE, lines=TRUE, columns=2))

mout <- gather(allout, key="Series", value="ANC"
  , Prol, Tx.1, Tx.2, Tx.3, Circ
  , factor_key = TRUE)

mout <- upData(mout
  , time = time/24
  , units = c(time="days")
  , print = FALSE)

# Figure 3 from citation [3]:
xYplot(ANC ~ time | id, group=Series
  , data=mout
  , layout=c(5,5)
  , type='l', as.table=TRUE
  , label.curves=FALSE
  , par.settings=list(superpose.line=list(lwd=2,col=brewer.pal(11,"RdYlBu"))[c(1,3,4,8,10)]))
  , scales=list(y=list(log=TRUE, lim=c(100,15000), at=c(200, 500, 1000, 2000, 5000, 10000)))
  , auto.key=list(points=FALSE, lines=TRUE, columns=5))

```

---

```
plot,DE,missing-method
```

*Plot a DE object as an interactive htmlwidget*

---

### Description

Plot a DE object as an interactive htmlwidget

### Usage

```
## S4 method for signature 'DE,missing'
plot(x, y, ..., devtree = FALSE)
```

### Arguments

x	An object of class DE
y	Unused; included for S4 generic consistency
...	Passed to <a href="#">r2d3</a> , enabling caller to (e.g.) the override the default viewer = "internal".
devtree	Logical indicator used to select local package dir

---

```
runDTATapp
```

*Run Shiny apps included in package DTAT*

---

### Description

Run Shiny apps included in package DTAT

### Usage

```
runDTATapp(app)
```

### Arguments

app	Character vector of length 1. Name of app to run.
-----	---------------------------------------------------

### Value

Invoked for side effect. Runs the named Shiny app.

### Examples

```
if(interactive()){
  runDTATapp("Sim33PC")
  runDTATapp("TheCost")
}
```

---

scaled	<i>Power-law scaling for doses</i>
--------	------------------------------------

---

**Description**

Implement an inverse power-law scaling for drug dose.

**Usage**

```
scaled(dose, a = 4)
```

**Arguments**

dose	A numeric vector of doses
a	A numeric exponent for power-law rescaling

**Value**

A rescaled vector of doses

**Author(s)**

David C. Norris

---

seq.function	<i>A seq method supporting custom-scaled plot axes.</i>
--------------	---------------------------------------------------------

---

**Description**

This provides a seq method for class function, supporting a natural axis scaling idiom.

**Usage**

```
## S3 method for class '`function`'  
seq(scalefun, from, to, length.out, digits = NULL, ...)
```

**Arguments**

scalefun	A numeric function that will be invoked componentwise, and so need not be vectorized)
from, to	The starting and ending values of the sequence returned
length.out	Desired length of the sequence
digits	If non-NULL, returned value is rounded accordingly
...	Unused; included for S3 generic/method consistency.

**Value**

A numeric vector that (not considering the effect of any rounding applied), becomes an arithmetic sequence after application of `scalefun` to it. The initial and final elements of that vector are `from` and `to`.

**Author(s)**

David C. Norris

**Examples**

```
# Provide evenly-spaced length-6 sequence from 100 to 1000,  
# evenly spaced on a fourth-root scale:  
seq(function(dose, a=4.0) dose^(1/a), from=100, to=1000, length.out=6, digits=0)
```

---

sim

*Environment for simulation global variables.*

---

**Description**

To simplify the code of package DTAT, as well as client tasks, this exported environment contains a handful of global variables useful for the simulations.

**Details**

Global variables maintained within environment `sim` are:

1. `pkpd`: The population PK/PD model to be simulated.
2. `pop`: A sample drawn from the population model.
3. `N`: Restricts simulation to first `N` subjects in `pop`.
4. `params.default`: Default parameters.

**Examples**

```
# Even when nrow(pop) is large, one may easily restrict  
# time-consuming simulations to pop[1:N,], as follows:  
sim$N <- 25  
# Now perform simulation work  
## Not run:  
titrate(...)  
  
## End(Not run)
```

---

titrate	<i>Perform neutrophil-guided dose titration of a chemotherapy drug.</i>
---------	-------------------------------------------------------------------------

---

### Description

This is included in package DTAT mainly for archival purposes, with the aim to document a reproduction of Figure 5 from the 2017 *F1000Research* paper (referenced below), using a clearer and more general software design than is found in the online code supplement available at <https://osf.io/vwnqz/>.

### Usage

```
titrate(draw.days = NULL, Ncycles = 10, doserange = c(50, 500), dta = NULL)
```

### Arguments

draw.days	Integer days on which ANC is to be measured
Ncycles	Number of chemo cycles through which to simulate titration
doserange	Range of doses to consider
dta	A Dose Titration Algorithm (DTA) to drive the titration

### Value

A list with 2 components:

course	A data frame containing cycle-wise measures of each id's titration course
anc.ts	A data frame detailing high-frequency ANC measures for each id

### Author(s)

David C. Norris

### References

Norris DC. Dose Titration Algorithm Tuning (DTAT) should supersede 'the' Maximum Tolerated Dose (MTD) in oncology dose-finding trials. *F1000Research*. 2017;6:112. doi:10.12688/f1000research.10624.3. <https://f1000research.com/articles/6-112/v3>

### Examples

```
if(interactive()){
# Reproduce Figure 5 from the F1000Research paper (run time > 10 s).
# 1. Set up sim$pop & sim$pkpd by running the repro for Figures 1 & 3:
example(topic="Onoue.Friberg", package="DTAT", ask=FALSE)
# 2. Do the neutrophil-nadir-guided dose titration:
chemo <- titrate(doserange = c(50, 3000),
                 dta=newton.raphson(dose1 = 50,
                                   omega = 0.75,
                                   slope1 = -2.0,
```

```

                                slopeU = -0.2)
)
library(latticeExtra)
newton <- chemo$course
new.ts <- chemo$anc.ts
anc.tics <- c(200,500,1500,4000,10000)
right <- xyplot(ANC ~ time | id, data=new.ts
               , as.table=TRUE, type="l"
               , layout=c(5,5)
               , scales=list(y=list(log=TRUE, lim=c(100,1.5e4)
                                , at=anc.tics
                                , lab=as.character(anc.tics)),
                             x=list(at=seq(0,30,3)))
)
newton <- upData(newton
               , time = 3*(cycle-1)
               , labels = c(time="Time")
               , units = c(time="weeks")
               , print = FALSE)
dose.tics <- c(50, 200, 600, 1500, 3000)
left <- xyplot(scaled.dose ~ time | id, data=newton
              , as.table=TRUE, type='p', pch='+', cex=1.5
              , layout=c(5,5)
              , scales=list(y=list(lim=DTAT::scaled(c(30,3200))
                                    , at=DTAT::scaled(dose.tics)
                                    , lab=as.character(dose.tics)),
                             x=list(lim=c(-1,31)
                                    , at=seq(0,30,3)
                                    , lab=c('0',' ','6',' ','12',' ','18',' ','24',' ','30'))))
)
update(doubleYScale(left, right, add.ylab2=TRUE)
       , par.settings = simpleTheme(col=brewer.pal(4,"PRGn"))[c(4,1)])
)
}

```

---

titration

*Simulate a '3+3/PC' dose-titration trial*


---

### Description

Simulate a '3+3/PC' dose-titration trial

### Usage

```
titration(x, periods, ...)
```

```
## S4 method for signature 'DE,numeric'
```

```
titration(x, periods, ...)
```

**Arguments**

x	An object of S4 class <a href="#">DE</a>
periods	The number of DLT assessment periods to titrate over. Should be a positive integer.
...	May be used to pass <code>verbatim = 'TRUE'</code> to internal <code>step_time</code> method.

**References**

Norris DC. Precautionary Coherence Unravels Dose Escalation Designs. *bioRxiv*. December 2017:240846. doi:10.1101/240846. <https://www.biorxiv.org/content/10.1101/240846v1>

# Index

- \* **datasets**
  - de.bioRxiv.240846, 3
  - dtat1000, 7
  - sim, 15
- \* **survival**
  - dose.survfit, 4
  - dose.survival, 5
  - ds.curve, 6
  
- as\_d3\_data, DE-method, 2
  
- DE, 5, 18
- DE-class, 3
- de.bioRxiv.240846, 3
- dose.survfit, 4, 6, 7
- dose.survival, 5, 5
- ds.curve, 6
- dtat1000, 7
  
- km.ci, 5
  
- newton.raphson, 9
  
- Onoue.Friberg, 10
  
- plot, DE, missing-method, 12
- pomp, 11
  
- r2d3, 13
- runDTATapp, 13
  
- scaled, 14
- seq.function, 14
- sim, 11, 15
- Surv, 5
  
- titrate, 10, 16
- titration, 17
- titration, DE, numeric-method  
(titration), 17