

Package ‘FastRet’

May 7, 2026

Title Retention Time Prediction in Liquid Chromatography

Version 1.3.0

Description A framework for predicting retention times in liquid chromatography. Users can train custom models for specific chromatography columns, predict retention times using existing models, or adjust existing models to account for altered experimental conditions. The provided functionalities can be accessed either via the R console or via a graphical user interface. Related work: Bonini et al. (2020) <[doi:10.1021/acs.analchem.9b05765](https://doi.org/10.1021/acs.analchem.9b05765)>.

License GPL-3

Language en-US

URL <https://github.com/spang-lab/FastRet/>,
<https://spang-lab.github.io/FastRet/>

BugReports <https://github.com/spang-lab/FastRet/issues>

biocViews Retention, Time, Chromotography, LC-MS

Encoding UTF-8

RoxygenNote 7.2.2

Depends R (>= 4.1.0)

Imports bslib, cluster, data.table, DT, future, ggplot2, glmnet,
htmltools, openxlsx, promises, rcdk, rlang, shiny (>= 1.8.1),
shinyhelper, shinyjs, withr, xgboost

Suggests callr, caret, cli, devtools, knitr, languageserver, linter,
pkgdown, pkgbuild, pkgload, rmarkdown, servr, tibble, testthat
(>= 3.0.0), toscutil, usethis

LazyData true

Config/testthat/edition 3

Config/testthat/parallel true

Config/testthat/start-first getCDs, parLapply2, get_predictors,
plot_frm, train_frm-lasso, train_frm-gbtree, fit_gbtree

NeedsCompilation no

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Repository CRAN

Date/Publication 2025-12-17 23:40:02 UTC

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adjust_frm	<i>Adjust an existing FastRet model for use with a new column</i>
------------	---

Description

The goal of this function is to train a model that predicts RT_ADJ (retention time measured on a new, adjusted column) from RT (retention time measured on the original column) and to attach this adjustment model to an existing FastRet model.

Usage

```
adjust_frm(
  frm,
  new_data,
  predictors = 1:6,
  nfolds = 5,
  verbose = 1,
  seed = NULL,
  do_cv = TRUE,
  adj_type = "lm",
  add_cds = NULL
)
```

Arguments

frm	An object of class frm as returned by <code>train_frm()</code> .
new_data	Data frame with required columns "RT", "NAME", "SMILES"; optional "INCHIKEY". "RT" must be the retention time measured on the adjusted column. Each row must match at least one row in <code>frm\$df</code> . The exact matching behavior is described in 'Details'.
predictors	Numeric vector specifying which transformations to include in the model. Available options are: 1=RT, 2=RT ² , 3=RT ³ , 4=log(RT), 5=exp(RT), 6=sqrt(RT). Note that predictor 1 (RT) is always included, even if not specified explicitly.
nfolds	The number of folds for cross validation.
verbose	Show progress messages?
seed	An integer value to set the seed for random number generation to allow for reproducible results.
do_cv	A logical value indicating whether to perform cross-validation. If FALSE, the cv element in the returned adjustment object will be NULL.
adj_type	A string representing the adjustment model type. Either "lm", "lasso", "ridge", or "gbtree".
add_cds	A logical value indicating whether to add chemical descriptors as predictors to new data. Default is TRUE if adj_type is "lasso", "ridge" or "gbtree" and FALSE if adj_type is "lm".

Details

Matching is done via "SMILES"+"INCHIKEY" if both datasets have non-missing INCHIKEYs for all rows; otherwise via "SMILES"+"NAME". If multiple rows in `frm$df` match the same row in `new_data`, their RT values are averaged first, and this average is used for training the adjustment model.

Example: if `frm$df` equals data.frame OLD shown below and `new_data` equals data.frame NEW, then the resulting, paired data.frame will look like PAIRED.

```
OLD <- data.frame(
```

```

NAME = c("A", "B", "B", "C" ),
SMILES = c("C", "CC", "CC", "CCC"),
RT = c(5.0, 8.0, 8.2, 9.0 )
)
NEW <- data.frame(
  NAME = c("A", "B", "B", "B"),
  SMILES = c("C", "CC", "CC", "CC"),
  RT = c(2.5, 5.5, 5.7, 5.6)
)
PAIRED <- data.frame(
  NAME = c("A", "B", "B", "B"),
  SMILES = c("C", "CC", "CC", "CC"),
  RT = c(5.0, 8.1, 8.1, 8.1), # Average of OLD$RT[2:3]
  RT_ADJ = c(2.5, 5.5, 5.7, 5.6) # Taken from NEW
)

```

If `do_cv` is `TRUE`, the adjustment procedure is evaluated in cross-validation. However, care must be taken when interpreting the CV results, as the model performance depends on both the adjustment layer and the original model, which was trained on the full base dataset. Therefore, the observed CV metrics should be read as "expected performance when predicting RTs for molecules that were part of the base-model training but not part of the adjustment set" instead of "expected performance when predicting RTs for completely new molecules".

Value

An object of class `frm`, as returned by `train_frm()`, but with an additional element `adj` containing the adjustment model. Components of `adj` are:

- `model`: The fitted adjustment model. Class depends on `adj_type` and is one of `lm`, `glmnet`, or `xgb.Booster`.
- `df`: The data frame used for training the adjustment model. Including columns "NAME", "SMILES", "RT", "RT_ADJ" and optionally "INCHIKEY", as well as any additional predictors specified via the `predictors` argument.
- `cv`: A named list containing the cross validation results (see 'Details'), or `NULL` if `do_cv = FALSE`. When not `NULL`, elements are:
 - `folds`: A list of integer vectors specifying the samples in each fold.
 - `models`: A list of adjustment models trained on each fold.
 - `stats`: A list of vectors with RMSE, Rsquared, MAE, pBelow1Min per fold. Added with v1.3.0.
 - `preds`: Retention time predictions obtained during CV by applying the adjustment model to the hold-out data.
 - `preds_adjonly`: Removed (i.e. `NULL`) since v1.3.0.
- `args`: Function arguments used for adjustment (excluding `frm`, `new_data` and `verbose`). Added with v1.3.0.
- `version`: The version of the `FastRet` package used to train the adjustment model. Added with v1.3.0.

Examples

```
frm <- read_rp_lasso_model_rds()
new_data <- read_rpadj_xlsx()
frm_adj <- adjust_frm(frm, new_data, verbose = 0)
```

clip_predictions *Clip predictions to observed range*

Description

Clips predicted retention times by fitting a log-normal distribution to the observed training RTs and bounding predictions to the central 99.99% interval. All observed RTs must be positive to estimate the distribution. If the estimated lower bound would be negative, it is replaced by 1% of the observed minimum RT instead.

Usage

```
clip_predictions(yhat, y)
```

Arguments

yhat Numeric vector of predicted retention times.
y Numeric vector of observed retention times used to derive bounds.

Value

Numeric vector of clipped (bounded) predictions.

Examples

```
# Draw only a few samples (10) and clip based on these. The allowed range will  
# be much bigger than the observed range.
```

```
set.seed(42)
y <- rlnorm(n = 1000, meanlog = 2, sdlog = 0.1)
yhat <- y
yhat[1] <- -100 # way too low to be realistic
yhat[2] <- 1000 # way too high to be realistic
yhat <- clip_predictions(yhat, y)
range(y) # [ 6.18, 8.93]
yhat[1:2] # [ 4.96, 10.61] # Limited by theoretical bounds
```

```
# Draw more samples (1000) and clip based on these. The allowed range will  
# be almost identical to the observed range.
```

```
set.seed(42)
y <- rnorm(n = 100, mean = 100, sd = 5)
```

```
yhat <- y
yhat[1] <- -100
yhat[2] <- 1000
yhat <- clip_predictions(yhat, y)
range(y) # 83.14, 117.47
yhat[1:2] # 83.14, 117.72
```

fastret_app

The FastRet GUI

Description

Creates the FastRet GUI

Usage

```
fastret_app(port = 8080, host = "0.0.0.0", reload = FALSE, nsw = 1)
```

Arguments

port	The port the application should listen on
host	The address the application should listen on
reload	Whether to reload the application when the source code changes
nsw	The number of subworkers each worker is allowed to start. The higher this number, the faster individual tasks like model fitting can be processed.

Value

An object of class `shiny.appobj`.

Examples

```
x <- fastret_app()
if (interactive()) shiny::runApp(x)
```

getCDs	<i>Get Chemical Descriptors for a list of molecules</i>
--------	---

Description

Calculate Chemical Descriptors (CDs) for a list of molecules. Molecules can appear multiple times in the list.

Usage

```
getCDs(df, verbose = 1, nw = 1, keepdf = TRUE)
```

Arguments

df	dataframe with two mandatory columns: "NAME" and "SMILES"
verbose	0: no output, 1: progress, 2: more progress and warnings
nw	number of workers for parallel processing
keepdf	If TRUE, cbind(df, CDs) is returned. Else CDs.

Value

A dataframe with all input columns (if keepdf is TRUE) and chemical descriptors as remaining columns.

Examples

```
cds <- getCDs(head(RP, 3), verbose = 1, nw = 1)
```

plot_frm	<i>Plot predictions for a FastRet model</i>
----------	---

Description

Creates scatter plots of measured vs. predicted retention times (RT) for a FastRet Model (FRM). Supports plotting cross-validation (CV) predictions and fitted predictions on the training set, as well as their adjusted variants when the model has been adjusted via [adjust_frm\(\)](#). Coloring highlights points within 1 minute of the identity line and simple outliers.

Usage

```
plot_frm(frm = train_frm(verbose = 1), type = "scatter.cv", trafo = "identity")
```

Arguments

frm	An object of class frm as returned by <code>train_frm()</code> .
type	Plot type. One of: <ul style="list-style-type: none"> • "scatter.cv": CV predictions for the training set • "scatter.cv.adj": CV predictions for the adjustment set (requires frm\$adj) • "scatter.train": Model predictions for the training set • "scatter.train.adj": Adjusted model predictions for the adjustment set (requires frm\$adj)
trafo	Transformation applied for display. One of: <ul style="list-style-type: none"> • "identity": no transformation • "log2": apply log2 transform to axes (metrics are computed on raw values)

Value

NULL, called for its side effect of plotting.

Examples

```
frm <- read_rp_lasso_model_rds()
plot_frm(frm, type = "scatter.cv")
```

predict.frm

Predict retention times using a FastRet Model

Description

Predict retention times for new data using a FastRet Model (FRM).

Usage

```
## S3 method for class 'frm'
predict(
  object = train_frm(),
  df = object$df,
  adjust = NULL,
  verbose = 0,
  clip = TRUE,
  impute = TRUE,
  ...
)
```

Arguments

object	An object of class frm as returned by <code>train_frm()</code> .
df	A data.frame with the same columns as the training data.
adjust	If object was adjusted using <code>adjust_frm()</code> , it will contain a property <code>object\$adj</code> . If <code>adjust</code> is TRUE, <code>object\$adj</code> will be used to adjust predictions obtained from <code>object\$model</code> . If FALSE <code>object\$adj</code> will be ignored. If NULL, <code>object\$model</code> will be used, if available.
verbose	A logical value indicating whether to print progress messages.
clip	Clip predictions to be within RT range of training data?
impute	Impute missing predictor values using column means of training data?
...	Not used. Required to match the generic signature of <code>predict()</code> .

Value

A numeric vector with the predicted retention times.

See Also

`train_frm()`, `adjust_frm()`

Examples

```
object <- read_rp_lasso_model_rds()
df <- head(RP)
yhat <- predict(object, df)
```

preprocess_data	<i>Preprocess data</i>
-----------------	------------------------

Description

Preprocess data so they can be used as input for `train_frm()`.

Usage

```
preprocess_data(
  data,
  degree_polynomial = 1,
  interaction_terms = FALSE,
  verbose = 1,
  nw = 1,
  rm_near_zero_var = TRUE,
  rm_na = TRUE,
  add_cds = TRUE,
  rm_ucs = TRUE,
  rt_terms = 1,
  mandatory = c("NAME", "RT", "SMILES")
)
```

Arguments

data	Dataframe with following columns: <ul style="list-style-type: none"> • Mandatory: NAME, RT and SMILES. • Recommended: INCHIKEY. • Optional: Any of the chemical descriptors listed in CDFeatures. All other columns will be removed. See 'Details'.
degree_polynomial	Add predictors with polynomial terms up to the specified degree, e.g. 2 means "add squares", 3 means "add squares and cubes". Set to 1 to leave descriptors unchanged.
interaction_terms	Add interaction terms? Polynomial terms are not included in the generation of interaction terms.
verbose	0: no output, 1: show progress, 2: progress and warnings.
nw	Number of workers to use for parallel processing.
rm_near_zero_var	Remove near zero variance predictors?
rm_na	Remove NA values?
add_cds	Add chemical descriptors using getCDS() ? See 'Details'.
rm_ucs	Remove unsupported columns?
rt_terms	Which retention-time transformations to append as extra predictors. Supply a numeric vector referencing predefined <code>rt_terms</code> (1=RT, 2=I(RT ²), 3=I(RT ³), 4=log(RT), 5=exp(RT), 6=sqrt(RT)) or a character vector with the explicit transformation terms. Character values are passed to <code>model.frame()</code> , so they must use valid formula syntax (e.g. "I(RT ²)" rather than "RT ² ").
mandatory	Character vector of mandatory columns that must be present in data. If any of these columns are missing, an error is raised.

Details

If `add_cds = TRUE`, chemical descriptors are added using [getCDS\(\)](#). If **all** chemical descriptors listed in [CDFeatures](#) are already present in the input data object, [getCDS\(\)](#) will leave them unchanged. If one or more chemical descriptors are missing, **all** chemical descriptors will be recalculated and existing ones will be overwritten.

Value

A dataframe with the preprocessed data.

Examples

```
data <- head(RP, 3)
pre <- preprocess_data(data, verbose = 0)
```

read_retip_hilic_data *Read the HILIC dataset from the Retip package*

Description

Reads the Retip::HILIC dataset (CC BY 4.0) from the Retip package or, if Retip is not installed, downloads the dataset directly from the [Retip GitHub repository](#). Before returning the dataset, SMILES strings are canonicalized and the original tibble object is converted to a base R data.frame.

Usage

```
read_retip_hilic_data(verbose = 1)
```

Arguments

verbose Verbosity. 1 for messages, 0 to suppress them.

Details

Attribution as required by CC BY 4.0:

- Original dataset by: Paolo Bonini, Tobias Kind, Hiroshi Tsugawa, Dinesh Kumar Barupal, and Oliver Fiehn as part of the Retip project.
- Source repository: <https://github.com/oloBion/Retip>
- Original file: <https://github.com/oloBion/Retip/raw/master/data/HILIC.RData>
- License: CC BY 4.0 (<https://creativecommons.org/licenses/by/4.0/>)
- Modifications in FastRet:
 - converted tibble to data.frame
 - canonicalized SMILES using `as_canonical()`
 - renamed column 'INCHKEY' to 'INCHIKEY'

Value

A data frame with 970 rows and the following columns:

- NAME: Molecule name
- INCHIKEY: InChIKey
- SMILES: Canonical SMILES string
- RT: Retention time in Minutes

Source

<https://github.com/oloBion/Retip/raw/master/data/HILIC.RData>

References

Retip: Retention Time Prediction for Compound Annotation in Untargeted Metabolomics
Paolo Bonini, Tobias Kind, Hiroshi Tsugawa, Dinesh Kumar Barupal, and Oliver Fiehn
Analytical Chemistry 2020 92 (11), 7515-7522 DOI: 10.1021/acs.analchem.9b05765

Examples

```
df <- read_retip_hilic_data(verbose = 0)
```

read_rpadj_xlsx	<i>Hypothetical retention times</i>
-----------------	-------------------------------------

Description

Subset of the data from [read_rp_xlsx\(\)](#) with some slight modifications to simulate changes in temperature and/or flowrate.

Usage

```
read_rpadj_xlsx()
```

Value

A dataframe with 25 rows (metabolites) and 3 columns: RT, SMILES and NAME.

Examples

```
x <- read_rpadj_xlsx()
```

read_rp_lasso_model_rds	<i>LASSO Model trained on RP dataset</i>
-------------------------	--

Description

Read a LASSO model trained on the [RP](#) dataset using [train_frm\(\)](#).

Usage

```
read_rp_lasso_model_rds()
```

Value

A frm object.

Examples

```
frm <- read_rp_lasso_model_rds()
```

read_rp_xlsx	<i>Read retention times (RT) measured on a reverse phase (RP) column</i>
--------------	--

Description

Reads retention times from a reverse phase liquid chromatography experiment, performed at 35°C and a flow rate of 0.3 mL/min. The data is also available as a dataframe in the package; to access it directly, use [RP](#).

Usage

```
read_rp_xlsx()
```

Value

A dataframe of 442 metabolites with columns RT, SMILES and NAME.

Source

Measured by the Institute of Functional Genomics at the University of Regensburg.

See Also

[RP](#)

Examples

```
x <- read_rp_xlsx()
all.equal(x, RP)
```


4. Scaling the chemical descriptors by coefficients of the Ridge Regression model.
5. Clustering the entire dataset, which includes the scaled chemical descriptors and the retention times.
6. Returning the clustering results, which include the cluster assignments, the medoid indicators, and the raw data.

Usage

```
selective_measuring(  
  raw_data,  
  k_cluster = 25,  
  verbose = 1,  
  seed = NULL,  
  rt_coef = "max_ridge_coef"  
)
```

Arguments

<code>raw_data</code>	The raw data to be processed. Must be a dataframe with columns NAME, RT and SMILES.
<code>k_cluster</code>	The number of clusters for PAM clustering.
<code>verbose</code>	The level of verbosity.
<code>seed</code>	An optional random seed for reproducibility, set at the beginning of the function.
<code>rt_coef</code>	Which coefficient to use for scaling RT before clustering. Options are: <ul style="list-style-type: none">• <code>max_ridge_coef</code>: scale with the maximum absolute coefficient obtained in ridge regression. I.e., RT will have approximately the same weight as the most important chemical descriptor.• <code>1</code>: do not scale RT any further, i.e., use standardized RT. The effect of leaving RT unscaled is kind of unpredictable, as the ridge coefficients depend on the dataset. If the maximum absolute coefficient is much smaller than 1, RT will dominate the clustering. If it is much larger than 1, RT will have little influence on the clustering.• <code>0</code>: exclude RT from the clustering.

Value

A list containing the following elements:

- `clustering`: A data frame with columns RT, SMILES, NAME, CLUSTER and IS_MEDOID.
- `clobj`: The clustering object. The object returned by the clustering function. Depends on the method parameter.
- `coefs`: The coefficients from the Ridge Regression model.
- `model`: The Ridge Regression model.
- `df`: The preprocessed data.
- `dfz`: The standardized features.
- `dfzb`: The features scaled by the coefficients (betas) of the Ridge Regression model.

Examples

```
x <- selective_measuring(RP[1:50, ], k = 5, verbose = 0)
# For the sake of a short runtime, only the first 50 rows of the RP dataset
# were used in this example. In practice, you should always use the entire
# dataset to find the optimal subset for re-measurement.
```

start_gui	<i>Start the FastRet GUI</i>
-----------	------------------------------

Description

Starts the FastRet GUI

Usage

```
start_gui(port = 8080, host = "0.0.0.0", reload = FALSE, nw = 2, nsw = 1)
```

Arguments

port	The port the application should listen on
host	The address the application should listen on
reload	Whether to reload the application when the source code changes
nw	The number of worker processes started. The first worker always listens for user input from the GUI. The other workers are used for handling long running tasks like model fitting or clustering. If nw is 1, the same process is used for both tasks, which means that the GUI will become unresponsive during long running tasks.
nsw	The number of subworkers each worker is allowed to start. The higher this number, the faster individual tasks like model fitting can be processed. A value of 1 means that all subprocesses will run sequentially.

Details

If you set `nw = 3` and `nsw = 4`, you should have at least 16 cores, one core for the shiny main process. Three cores for the three worker processes and 12 cores ($3 * 4$) for the subworkers. For the default case, `nworkers = 2` and `nsw = 1`, you only need 3 cores, as `nsw = 1` means that all subprocesses will run sequentially.

Value

A shiny app. This function returns a shiny app that can be run to interact with the model.

Examples

```
if (interactive()) start_gui()
```

`train_frm`*Train a new FastRet model (FRM) for retention time prediction*

Description

Trains a new model from molecule SMILES to predict retention times (RT) using the specified method.

Usage

```
train_frm(  
  df,  
  method = "lasso",  
  verbose = 1,  
  nfolds = 5,  
  nw = 1,  
  degree_polynomial = 1,  
  interaction_terms = FALSE,  
  rm_near_zero_var = TRUE,  
  rm_na = TRUE,  
  rm_ns = FALSE,  
  seed = NULL,  
  do_cv = TRUE  
)
```

Arguments

<code>df</code>	A dataframe with columns "NAME", "RT", "SMILES" and optionally a set of chemical descriptors. If no chemical descriptors are provided, they are calculated using the function preprocess_data() .
<code>method</code>	A string representing the prediction algorithm. Either "lasso", "ridge", "gbtree", "gbtreeDefault" or "gbtreeRP". Method "gbtree" is an alias for "gbtreeDefault".
<code>verbose</code>	A logical value indicating whether to print progress messages.
<code>nfolds</code>	An integer representing the number of folds for cross validation.
<code>nw</code>	An integer representing the number of workers for parallel processing.
<code>degree_polynomial</code>	An integer representing the degree of the polynomial. Polynomials up to the specified degree are included in the model.
<code>interaction_terms</code>	A logical value indicating whether to include interaction terms in the model.
<code>rm_near_zero_var</code>	A logical value indicating whether to remove near zero variance predictors.
<code>rm_na</code>	A logical value indicating whether to remove NA values before training. Highly recommended to avoid issues during model fitting. Setting this to FALSE with <code>method = "lasso"</code> will most likely lead to errors.

rm_ns	A logical value indicating whether to remove chemical descriptors that were considered as not suitable for linear regression based on a previous analysis of an independent dataset. Currently not used.
seed	An integer value to set the seed for random number generation to allow for reproducible results.
do_cv	A logical value indicating whether to perform cross-validation. If FALSE, the cv element in the returned object will be NULL.

Value

A 'FastRet Model', i.e., an object of class frm. Components are:

- model: The fitted base model. This can be an object of class glmnet (for Lasso or Ridge regression) or xgb.Booster (for GBTree models).
- df: The data frame used for training the model. The data frame contains all user-provided columns (including mandatory columns RT, SMILES and NAME) as well the calculated chemical descriptors. (But no interaction terms or polynomial features, as these can be recreated within a few milliseconds).
- cv: A named list containing the cross validation results, or NULL if do_cv = FALSE. When not NULL, elements are:
 - folds: A list of integer vectors specifying the samples in each fold.
 - models: A list of models trained on each fold.
 - stats: A list of vectors with RMSE, Rsquared, MAE, pBelow1Min per fold.
 - preds: Retention time predictions obtained in CV as numeric vector.
- seed: The seed used for random number generation.
- version: The version of the FastRet package used to train the model.
- args: The value of function arguments besides df as named list.

Examples

```
m <- train_frm(df = RP[1:40, ], method = "lasso", nfolds = 2, verbose = 0)
# For the sake of a short runtime, only the first 40 rows of the RP dataset
# are used in this example. In practice, you should always use the entire
# training dataset for model training.
```

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