

# Package ‘HaDeX2’

May 7, 2026

**Title** Analysis and Visualisation of Hydrogen/Deuterium Exchange Mass Spectrometry Data

**Version** 1.0.0

**Description** Processing, analysis and visualization of Hydrogen Deuterium eXchange monitored by Mass Spectrometry experiments (HDX-MS). 'HaDeX2' introduces a new standardized and reproducible workflow for the analysis of the HDX-MS data, including uncertainty propagation, data aggregation and visualization on 3D structure. Additionally, it covers data exploration, quality control and generation of publication-quality figures. All functionalities are also available in the accompanying 'shiny' app.

**Depends** R (>= 3.5)

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.3.3

**Imports** checkmate, data.table, dplyr, ggplot2, glue, gridExtra, magick, purrr, readr, readxl, r3dmol (>= 0.1.2), remotes, stringi, tidyr, ggiraph

**Suggests** bookdown, digest, knitr, magrittr, pander, renv, rmarkdown, microbenchmark, testthat, vdiff, scales, shiny, spelling

**VignetteBuilder** knitr

**Language** en-US

**URL** <https://hadexversum.github.io/HaDeX2/>

**NeedsCompilation** no

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**Date/Publication** 2026-02-09 13:30:26 UTC

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HaDeX2-package

*HaDeX2***Description**

The HaDeX2 package is a toolbox for the analysis of HDX-MS data.

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**See Also**

Useful links:

- <https://hadexversum.github.io/HaDeX2/>

---

add\_stat\_dependency     *Calculates confidence limits*

---

**Description**

Returns relation with confidence limits for each peptide.

**Usage**

```
add_stat_dependency(  
  calc_dat,  
  confidence_level = 0.98,  
  theoretical = FALSE,  
  fractional = TRUE  
)
```

**Arguments**

calc_dat	data produced by <a href="#">calculate_diff_uptake</a> function.
confidence_level	confidence limit - from range [0, 1].
theoretical	logical, determines if values are theoretical.
fractional	logical, determines if values are fractional.

**Details**

This function checks if the values are statistically significant based on provided criteria using Houde test.

**Value**

calc\_dat extended by column specifying if given peptide is relevant in given confidence limit. The value of the confidence limit is added as an attribute - as well as parameters used to calculate (theoretical/fractional).

**Examples**

```
calc_dat <- calculate_diff_uptake(alpha_dat)
result <- add_stat_dependency(calc_dat)
head(result)
```

---

alpha\_dat

*Elongation factor eEF1B subunit alpha*

---

**Description**

Originally published in: Bondarchuk, T. V., Shalak, V. F., Lozhko, D. M., Fatalaska, A., Szczepanowski, R. H., Liudkovska, V., Tsuvariev, O. Y., Dadlez, M., El'skaya, A. V., & Negrutskii, B. S. (2022). Quaternary organization of the human eEF1B complex reveals unique multi-GEF domain assembly. *Nucleic Acids Research*, 50(16), 9490–9504. <doi:/10.1093/nar/gkac685>

**Author(s)**

Bondarchuk, T. V., Shalak, V. F., Lozhko, D. M., Fatalaska, A., Szczepanowski, R. H., Liudkovska, V., Tsuvariev, O. Y., Dadlez, M., El'skaya, A. V., & Negrutskii, B. S.

**References**

[doi:10.1093/nar/gkac685](https://doi.org/10.1093/nar/gkac685)

---

calculate\_aggregated\_diff\_uptake

*Calculates aggregated deuterium uptake difference for one time point*

---

**Description**

Function aggregates the differential deuterium uptake values from peptide level into single-amino resolution using ‘weighted approach’ (defined in ‘vignette("datafiles")’). For visualization use [plot\\_aggregated\\_uptake](#)

**Usage**

```
calculate_aggregated_diff_uptake(diff_uptake_dat, time_t)
```

**Arguments**

diff\_uptake\_dat

differential uptake data, product of e.g. [create\\_diff\\_uptake\\_dataset](#)

time\_t

chosen time point

**Value**

a `data.frame` object

**Examples**

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
calculate_aggregated_diff_uptake(diff_uptake_dat, time_t = 5)
```

---

```
calculate_aggregated_test_results
      Aggregate test result
```

---

**Description**

Function aggregates peptide-level information into residue level. Significance method indicates if the difference is significant or not - if the number of significant peptides over this residue is bigger than the number of insignificant.

**Usage**

```
calculate_aggregated_test_results(
  p_diff_uptake_conf_dat,
  method = c("significance", "weiss"),
  time_t = 1,
  skip_amino = 1
)
```

**Arguments**

<code>p_diff_uptake_conf_dat</code>	uptake data with confidence, as produced by <a href="#">create_p_diff_uptake_dataset_with_confidence</a> function
<code>method</code>	method of aggregation: significance or weiss method
<code>time_t</code>	chosen time point
<code>skip_amino</code>	integer, indicator how many amino acids from the N-terminus should be omitted in visualization

**Details**

Only peptides without modification are aggregated.

**Value**

a `data.frame` object

**See Also**[read\\_hdx](#)**Examples**

```
p_diff_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat)
p_diff_uptake_conf_dat <- create_p_diff_uptake_dataset_with_confidence(p_diff_uptake_dat)
calculate_aggregated_test_results(p_diff_uptake_conf_dat, method = "significance")
calculate_aggregated_test_results(p_diff_uptake_conf_dat, method = "weiss")
```

---

`calculate_aggregated_uptake`*Calculates aggregated deuterium uptake for one time point*

---

**Description**

Function aggregates the deuterium uptake values from peptide level into single-amino resolution using ‘weighted approach’ (defined in ‘vignette("datafiles)")’. For visualization use [plot\\_aggregated\\_uptake](#)

**Usage**

```
calculate_aggregated_uptake(
  kin_dat,
  state = unique(kin_dat[["State"]])[1],
  time_t
)
```

**Arguments**

<code>kin_dat</code>	single state uptake data, product of e.g. <a href="#">create_uptake_dataset</a>
<code>state</code>	state included in calculations
<code>time_t</code>	chosen time point

**Value**

a `data.frame` object

**Examples**

```
# disabled due to long execution time

kin_dat <- create_uptake_dataset(alpha_dat, states = "Alpha_KSCN")
head(create_aggregated_uptake_dataset(kin_dat))
```

---

`calculate_auc`*Calculate Area Under the Curve*

---

### Description

Calculates area under the deuterium uptake curve

### Usage

```
calculate_auc(  
  uptake_dat,  
  protein = uptake_dat[["Protein"]][1],  
  state = uptake_dat[["State"]][1],  
  preserve_values = FALSE  
)
```

### Arguments

<code>uptake_dat</code>	data with deuterium uptake values, calculated e.g. by <a href="#">create_uptake_dataset</a>
<code>protein</code>	chosen protein
<code>state</code>	state included in calculations
<code>preserve_values</code>	indicator if the original columns from <code>uptake_dat</code> should be preserve in the result

### Details

The AUC is calculated on the data normalized to unit square by division by maximum values of exposure time and deuterium uptake, respectively.

### Value

a [data.frame](#) object

### See Also

[read\\_hdx](#) [create\\_uptake\\_dataset](#)

### Examples

```
uptake_dat <- create_uptake_dataset(alpha_dat)  
head(calculate_auc(uptake_dat))
```

---

`calculate_back_exchange`*Back exchange estimation*

---

### Description

Calculates back-exchange for a state

### Usage

```
calculate_back_exchange(  
  dat,  
  protein = dat[["Protein"]][1],  
  states = unique(dat[["State"]]),  
  time_100 = max(dat[["Exposure"]])  
)
```

### Arguments

<code>dat</code>	data imported by the <a href="#">read_hdx</a> function
<code>protein</code>	selected protein
<code>states</code>	selected biological states for given protein
<code>time_100</code>	time point of measurement for fully deuterated sample

### Details

Back-exchange is a reverse exchange phenomenon, important to acknowledge when working with HDX data. This function calculates back-exchange values for one biological state of the selected protein. For visualization of back-exchange data use [plot\\_coverage\\_heatmap](#) with displayed value specified as 'back-exchange'. For the definition of back-exchange see the 'vignette("datafiles")'.

### Value

a [data.frame](#) object

### See Also

[read\\_hdx](#) [plot\\_coverage\\_heatmap](#)

### Examples

```
head(calculate_back_exchange(alpha_dat))
```

---

`calculate_confidence_limit_values`*Calculate the value of confidence limit*

---

**Description**

Calculates confidence limit values for prepared provided, based on chosen parameters.

**Usage**

```
calculate_confidence_limit_values(  
  diff_uptake_dat,  
  confidence_level = 0.98,  
  theoretical = FALSE,  
  fractional = TRUE,  
  n_rep = NULL  
)
```

**Arguments**

<code>diff_uptake_dat</code>	differential data calculated using <code>calculate_diff_uptake</code> function
<code>confidence_level</code>	confidence level for the test, from range [0, 1]
<code>theoretical</code>	logical, determines if values are theoretical
<code>fractional</code>	logical, determines if values are fractional
<code>n_rep</code>	number of replicates

**Details**

Function `calculate_confidence_limit_values` calculates confidence limit using Houde test. The confidence limits are calculated on whole provided dataset. If the user wishes to calculate confidence limit for one, two or more time points, the provided data should be adjusted accordingly.

**Value**

range of confidence limit interval

**References**

Houde, D., Berkowitz, S.A., and Engen, J.R. (2011). The Utility of Hydrogen/Deuterium Exchange Mass Spectrometry in Biopharmaceutical Comparability Studies. *J Pharm Sci* 100, 2071–2086.

**See Also**

[read\\_hdx](#) [calculate\\_diff\\_uptake](#) [create\\_diff\\_uptake\\_dataset](#)

## Examples

```
diff_uptake_dat <- calculate_diff_uptake(alpha_dat)
calculate_confidence_limit_values(diff_uptake_dat)
```

---

calculate\_diff\_uptake *Calculate differential uptake*

---

## Description

Calculates differential deuterium uptake between two selected biological states.

## Usage

```
calculate_diff_uptake(  
  dat,  
  protein = unique(dat[["Protein"]][1]),  
  states = unique(dat[["State"]])[1:2],  
  time_0 = min(dat[["Exposure"]]),  
  time_t = unique(dat[["Exposure"]])[3],  
  time_100 = max(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

## Arguments

dat	data imported by the <a href="#">read_hdx</a> function
protein	chosen protein
states	vector of two states for chosen protein. Order is important, as the deuterium uptake difference is calculated as state_1 - state_2
time_0	minimal exchange control time point of measurement [min]
time_t	time point of the measurement for which the calculations are done [min]
time_100	maximal exchange control time point of measurement [min]
deut_part	deuterium percentage in solution used in experiment, value from range [0, 1]

## Details

Function [calculate\\_diff\\_uptake](#) calculates differential values based on provided criteria for peptides for chosen protein in selected states. The methods of calculation of deuterium uptake difference, fractional deuterium uptake difference with respect to minimal/maximal exchange controls or theoretical tabular values are thoroughly described in the ‘Data processing’ article, as well as law of propagation of uncertainty, used to calculate uncertainty.

## Value

a [data.frame](#) object.

**See Also**

[read\\_hdx](#) [calculate\\_state\\_uptake](#)

**Examples**

```
diff_dat <- calculate_diff_uptake(alpha_dat)
head(diff_dat)
```

---

calculate\_exp\_masses *Calculate measured mass, aggregated from the replicates of the experiment*

---

**Description**

Calculate the measured mass (with the uncertainty of the measurement) as aggregated data from the replicates of the experiment.

**Usage**

```
calculate_exp_masses(dat)
```

**Arguments**

dat                    data as imported by the [read\\_hdx](#) function

**Details**

Each measurement is repeated at least three times to obtain reliable result and to calculate uncertainty of the measurement. For more information on how the data is aggregated or how the uncertainty is calculated, see the documentation.

**Value**

a [data.frame](#) object.

**See Also**

[read\\_hdx](#) [calculate\\_exp\\_masses\\_per\\_replicate](#) [calculate\\_state\\_uptake](#)

**Examples**

```
calculate_exp_masses(alpha_dat)
```

---

`calculate_exp_masses_per_replicate`*Calculate measured mass for each replicate of the experiment*

---

**Description**

Calculate the measured mass from partial results, per each replicate of the experiment.

**Usage**

```
calculate_exp_masses_per_replicate(dat)
```

**Arguments**

`dat` data as imported by the [read\\_hdx](#) function

**Details**

Each replicate of the experiment generates measurements of the mass for obtained charge values for the peptide. This is an effect of the properties of mass spectrometry, that measures the mass to charge ratio (learn more about Mass Spectrometry in the documentation). The possible charge values depend on the sequence of the peptide. The separate measurement (for each replicate in given state in given time point) can be distinguished by the 'File' value.

**Value**

a `data.frame` object.

**See Also**

[read\\_hdx](#) [calculate\\_exp\\_masses](#) [calculate\\_state\\_uptake](#)

**Examples**

```
head(calculate_exp_masses_per_replicate(alpha_dat))
```

---

calculate\_kinetics      *Calculate kinetics data*

---

### Description

Calculate kinetics of the hydrogen-deuteration exchange for given peptide in given state.

### Usage

```
calculate_kinetics(  
  dat,  
  protein = dat[["Protein"]][1],  
  sequence = dat[["Sequence"]][1],  
  state = dat[["State"]][1],  
  start = dat[["Start"]][1],  
  end = dat[["End"]][1],  
  time_0 = min(dat[["Exposure"]]),  
  time_100 = max(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

### Arguments

dat	dat data imported by the <a href="#">read_hdx</a> function.
protein	protein chosen protein.
sequence	sequence of chosen peptide.
state	biological state of chosen peptide.
start	start position of chosen peptide.
end	end position of chosen peptide.
time_0	minimal exchange control time point of measurement.
time_100	maximal exchange control time point of measurement.
deut_part	deuterium percentage in solution used in experiment, value from range [0, 1].

### Details

The function calculates deuteration data for all available data points for given peptide in chosen biological state.. All four variants (relative & theoretical combinations) of deuterium uptake computations are supported. Manual correction of percentage of deuterium the protein was exposed to during the exchange in theoretical calculations is provided. To visualize obtained data we recommend using [plot\\_uptake\\_curve](#) function. The first version doesn't support filled Modification and Fragment columns. IMPORTANT! The kinetic data is often described as deuterium uptake curve data. We use this terms interchangeable.

### Value

a [data.frame](#) object.

**See Also**

[read\\_hdx](#) [calculate\\_state\\_uptake](#) [plot\\_uptake\\_curve](#)

**Examples**

```
# by default: for the first peptide
calculate_kinetics(alpha_dat)
```

---

calculate_MHP	<i>Calculate MHP of the peptide</i>
---------------	-------------------------------------

---

**Description**

Calculate the mass of the singly charged monoisotopic (or not) molecular ion of for given peptide.

**Usage**

```
calculate_MHP(Sequence, mono = FALSE)
```

**Arguments**

Sequence	sequence of the peptide (string) or vector of sequences. Each letter of the sequence of the peptide represents different amino acid (three letter representation not allowed)
mono	logical value to determine if the mass should be monoisotopic or not. FALSE by default

**Details**

This function calculates the mass of the singly charged monoisotopic (or not) molecular ion for given peptide. It is the sum of the residue masses plus the masses of the terminating group (H and OH). The source of the masses can be found here: [http://www.matrixscience.com/help/aa\\_help.html](http://www.matrixscience.com/help/aa_help.html). Keep in mind that this function returns the value of an unmodified peptide.

**Value**

vector of numeric MHP values of provided Sequences

**See Also**

[read\\_hdx](#) [calculate\\_state\\_uptake](#)

**Examples**

```
calculate_MHP("CHERICHERILADY")
calculate_MHP("CHERICHERILADY", mono = TRUE)
```

---

`calculate_peptide_kinetics`*Calculate kinetics dataset*

---

### Description

Calculate kinetics of the hydrogen-deuteration exchange for given peptide in multiple biological states.

### Usage

```
calculate_peptide_kinetics(  
  dat,  
  protein = dat[["Protein"]][1],  
  sequence = dat[["Sequence"]][1],  
  states = unique(dat[["State"]]),  
  start = dat[["Start"]][1],  
  end = dat[["End"]][1],  
  time_0 = min(dat[["Exposure"]]),  
  time_100 = max(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

### Arguments

<code>dat</code>	dat data imported by the <a href="#">read_hdx</a> function.
<code>protein</code>	chosen protein.
<code>sequence</code>	sequence of chosen peptide.
<code>states</code>	vector of states (for chosen protein), for which the calculations are done.
<code>start</code>	start position of chosen peptide.
<code>end</code>	end position of chosen peptide.
<code>time_0</code>	minimal exchange control time point of measurement.
<code>time_100</code>	maximal exchange control time point of measurement.
<code>deut_part</code>	deuterium percentage in solution used in experiment, value from range [0, 1].

### Details

Function [calculate\\_peptide\\_kinetics](#) calculates kinetic data for chosen peptide in chosen biological states. It is a wrapper for [calculate\\_kinetics](#) but for multiple states. The output of this function can be visualized using [plot\\_uptake\\_curve](#). IMPORTANT! The kinetic data is often described as deuterium uptake curve data. We use this terms interchangeable.

### Value

a [data.frame](#) object.

**See Also**

[calculate\\_kinetics](#) [calculate\\_state\\_uptake](#) [plot\\_uptake\\_curve](#)

**Examples**

```
# by default calculated for the first peptide from the peptide pool
calculate_peptide_kinetics(alpha_dat)
```

---

calculate_p_value	<i>Create p-value dataset</i>
-------------------	-------------------------------

---

**Description**

Create p-value dataset

**Usage**

```
calculate_p_value(
  dat,
  protein = unique(dat[["Protein"]])[1],
  state_1 = unique(dat[["State"]])[1],
  state_2 = unique(dat[["State"]])[2],
  p_adjustment_method = "none",
  confidence_level = 0.98
)
```

**Arguments**

dat	data imported by the <a href="#">read_hdx</a> function.
protein	chosen protein.
state_1	biological state for chosen protein. From this state values the second state values are subtracted to get the deuterium uptake difference.
state_2	biological state for chosen protein. This state values are subtracted from the first state values to get the deuterium uptake difference.
p_adjustment_method	method of adjustment P-values for multiple comparisons. Possible methods: "BH" (Benjamini & Hochberg correction), "bonferroni" (Bonferroni correction) and "none" (default).
confidence_level	confidence level for the t-test.

## Details

This function calculates P-value based on the supplied data. Unpaired t-Student test (with supplied parameters) is used to establish if the null hypothesis (there is no difference between measured mass values between two selected biological states) can be rejected, based on the experimental mass values from replicates of the experiment - for peptide in given time point of measurement. For the peptides that have only one replicate of the measurement (in any state) the P-value cannot be calculated and is assigned with NA value.

## Value

a `data.frame` object.

## See Also

[read\\_hdx](#) [calculate\\_exp\\_masses\\_per\\_replicate](#) [plot\\_volcano](#) [create\\_diff\\_uptake\\_dataset](#)  
[create\\_p\\_diff\\_uptake\\_dataset](#)

## Examples

```
p_dat <- calculate_p_value(alpha_dat)
head(p_dat)
```

---

calculate\_state\_uptake

*Calculate deuterium uptake*

---

## Description

Calculates deuteration uptake based on supplied parameters.

## Usage

```
calculate_state_uptake(
  dat,
  protein = unique(dat[["Protein"]])[1],
  state = unique(dat[["State"]])[1],
  time_0 = min(dat[dat[["Exposure"]] > 0, ][["Exposure"]]),
  time_t = unique(dat[["Exposure"]])[3],
  time_100 = max(dat[["Exposure"]]),
  deut_part = 0.9
)
```

### Arguments

dat	data as imported by the <a href="#">read_hdx</a> function
protein	chosen protein
state	state included in calculations
time_0	minimal exchange control
time_t	chosen time point
time_100	maximal exchange control
deut_part	percentage of deuterium the protein was exposed to, value in range [0, 1]

### Details

The function `calculate_state_uptake` calculates deuterium uptake (in different forms) for all of the peptides in given protein in given state based on supplied parameters: ‘time\_0’, ‘time\_100’ and ‘time\_t’. All four variants (fractional) are supplied (mean values and uncertainty). Manual correction of percentage of deuterium the protein was exposed to during the exchange in theoretical calculations is provided.

Methods of calculation and uncertainty are profoundly discussed in the vignette.

### Value

a `data.frame` object

### See Also

[read\\_hdx](#) [create\\_uptake\\_dataset](#) [calculate\\_confidence\\_limit\\_values](#) [add\\_stat\\_dependency](#)

### Examples

```
head(calculate_state_uptake(alpha_dat))
```

---

```
create_aggregated_diff_uptake_dataset
```

*Calculates aggregated uptake difference for peptide pool*

---

### Description

This is a wrapper function for [calculate\\_aggregated\\_diff\\_uptake](#), which calculates aggregated differential uptake for only one time point. This function returns values for all time points from ‘diff\_uptake\_dat’.

### Usage

```
create_aggregated_diff_uptake_dataset(diff_uptake_dat)
```

**Arguments**

diff\_uptake\_dat  
differential uptake data, product of e.g. [create\\_diff\\_uptake\\_dataset](#)

**Value**

a `data.frame` object

**Examples**

```
# disabled due to long execution time

diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
head(create_aggregated_diff_uptake_dataset(diff_uptake_dat))
```

---

create\_aggregated\_uptake\_dataset

*Calculates the aggregated uptake for peptide pool*

---

**Description**

This is a wrapper function for [calculate\\_aggregated\\_uptake](#), which calculates aggregated uptake for only one time point. This function returns values for all time points from 'kin\_dat'.

**Usage**

```
create_aggregated_uptake_dataset(kin_dat)
```

**Arguments**

kin\_dat            single state uptake data, product of e.g. [create\\_uptake\\_dataset](#)

**Value**

a `data.frame` object

**Examples**

```
# disabled due to long execution time

kin_dat <- create_uptake_dataset(alpha_dat, states = "Alpha_KSCN")
create_aggregated_uptake_dataset(kin_dat)
```

---

```
create_control_dataset
```

*Create dataset with control*

---

### Description

This function adds selected experimental maximal exchange control as a measurement for all biological states.

### Usage

```
create_control_dataset(  
  dat,  
  control_protein = dat[["Protein"]][1],  
  control_state = dat[["State"]][1],  
  control_exposure = max(dat[["Exposure"]])  
)
```

### Arguments

`dat` data imported by the [read\\_hdx](#) function.

`control_protein` maximal exchange control protein, from `dat`.

`control_state` maximal exchange control state, from `dat`.

`control_exposure` maximal exchange control exposure (time point of measurement), from `dat`.

### Details

Function [create\\_control\\_dataset](#) creates a dataset (similar to the output of [read\\_hdx](#) function), with maximal exchange control for all the states, based on provided parameters. The other functions are operating within a state, so the control is prepared for each state. The chosen maximal exchange control is distinguishable by the value '99999' in 'Exposure' control.

### Value

a `data.frame` object.

### See Also

[read\\_hdx](#) [calculate\\_state\\_uptake](#)

### Examples

```
head(create_control_dataset(alpha_dat))
```

---

```
create_diff_uptake_dataset
```

*Generate differential dataset*

---

## Description

Calculates differential deuterium uptake values between two states.

## Usage

```
create_diff_uptake_dataset(  
  dat,  
  protein = unique(dat[["Protein"]])[1],  
  state_1 = unique(dat[["State"]])[1],  
  state_2 = unique(dat[["State"]])[2],  
  time_0 = min(dat[["Exposure"]]),  
  time_100 = max(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

## Arguments

dat	data imported by the <a href="#">read_hdx</a> function.
protein	chosen protein.
state_1	biological state for chosen protein. From this state values the second state values are subtracted to get the deuterium uptake difference.
state_2	biological state for chosen protein. This state values are subtracted from the first state values to get the deuterium uptake difference.
time_0	minimal exchange control time point of measurement [min].
time_100	maximal exchange control time point of measurement [min].
deut_part	deuterium percentage in solution used in experiment, value from range [0, 1].

## Details

The function [create\\_diff\\_uptake\\_dataset](#) generates a dataset with differential values between given biological states (state\_1 - state\_2). For each peptide of chosen protein for time points of measurement between minimal and maximal control time points of measurement deuterium uptake difference, fractional deuterium uptake difference with respect to controls or theoretical tabular values are calculated, with combined and propagated uncertainty. Each peptide has an ID, based on its start position. Function output can be visualized as a differential (Woods) plot, butterfly differential plot or chiclet differential plot.

## Value

a [data.frame](#) object.

**See Also**

[read\\_hdx](#) [calculate\\_diff\\_uptake](#) [plot\\_differential\\_butterfly](#) [plot\\_differential\\_chiclet](#)  
[plot\\_differential](#)

**Examples**

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
head(diff_uptake_dat)
```

---

```
create_kinetic_dataset
```

*Create kinetics dataset for a list of peptides and their states*

---

**Description**

Generates the data set of deuterium uptake between selected time points based on supplied peptide list.

**Usage**

```
create_kinetic_dataset(
  dat,
  peptide_list,
  protein = dat[["Protein"]][1],
  time_0 = min(dat[["Exposure"]]),
  time_100 = max(dat[["Exposure"]]),
  deut_part = 0.9
)
```

**Arguments**

<code>dat</code>	dat data imported by the <a href="#">read_hdx</a> function.
<code>peptide_list</code>	list of peptides for the calculation.
<code>protein</code>	chosen protein.
<code>time_0</code>	minimal exchange control time point of measurement.
<code>time_100</code>	maximal exchange control time point of measurement.
<code>deut_part</code>	deuterium percentage in solution used in experiment, value from range [0, 1].

**Details**

This is a wrapper for [calculate\\_kinetics](#), but for the peptide list instead of one peptide.

**Value**

a `data.frame` object.

**See Also**

[calculate\\_kinetics](#) [calculate\\_state\\_uptake](#) [plot\\_uptake\\_curve](#)

**Examples**

```
peptide_list <- data.frame(Sequence = c("GFGDLKSPAGL", "FGDLKSPAGL"),
                          state = c("ALPHA_Gamma", "ALPHA_Gamma"),
                          start = c(1, 2), end = c(11, 11))
create_kinetic_dataset(alpha_dat, peptide_list)
```

---

```
create_overlap_distribution_dataset
      Show overlap distribution data
```

---

**Description**

Generates the data of frequency of overlap of each amino in the protein sequence.

**Usage**

```
create_overlap_distribution_dataset(
  dat,
  protein = dat[["Protein"]][1],
  state = dat[["State"]][1],
  start = min(dat[["Start"]]),
  end = max(dat[["End"]]),
  protein_sequence = reconstruct_sequence(dat)
)
```

**Arguments**

dat	data imported by the <a href="#">read_hdx</a> function.
protein	chosen protein.
state	biological state for chosen protein.
start	start position of chosen protein.
end	end position of chosen protein.
protein_sequence	data produced by <a href="#">reconstruct_sequence</a> function.

**Details**

This data frame presents how many times (by how many peptides) a amino position in protein sequence is covered. This data is available in the GUI.

**Value**

a `data.frame` object.

**See Also**

[read\\_hdx](#) [reconstruct\\_sequence](#)

**Examples**

```
create_overlap_distribution_dataset(alpha_dat)
```

---

```
create_p_diff_uptake_dataset
```

*Create differential uptake dataset with p-value*

---

**Description**

Creates differential deuterium uptake dataset with P-value from t-Student test for selected two biological states.

**Usage**

```
create_p_diff_uptake_dataset(  
  dat,  
  diff_uptake_dat = NULL,  
  protein = unique(dat[["Protein"]])[1],  
  state_1 = unique(dat[["State"]])[1],  
  state_2 = unique(dat[["State"]])[2],  
  p_adjustment_method = "none",  
  confidence_level = 0.98,  
  time_0 = min(dat[["Exposure"]]),  
  time_100 = max(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

**Arguments**

<code>dat</code>	data imported by the <a href="#">read_hdx</a> function.
<code>diff_uptake_dat</code>	differential uptake data
<code>protein</code>	chosen protein.
<code>state_1</code>	biological state for chosen protein. From this state values the second state values are subtracted to get the deuterium uptake difference.
<code>state_2</code>	biological state for chosen protein. This state values are subtracted from the first state values to get the deuterium uptake difference.

p_adjustment_method	method of adjustment P-values for multiple comparisons. Possible methods: "BH" (Benjamini & Hochberg correction), "bonferroni" (Bonferroni correction) and "none" (default).
confidence_level	confidence level for the t-test.
time_0	minimal exchange control time point of measurement [min].
time_100	maximal exchange control time point of measurement [min].
deut_part	deuterium percentage in solution used in experiment, value from range [0, 1].

### Details

For peptides in all of the time points of measurement (except for minimal and maximal exchange control) the deuterium uptake difference between state\_1 and state\_2 is calculated, with its uncertainty (combined and propagated as described in 'Data processing' article). For each peptide in time point the P-value is calculated using unpaired t-test. The deuterium uptake difference is calculated as the difference of measured masses in a given time point for two states. The tested hypothesis is that the mean masses for states from the replicates of the experiment are similar. The P-values indicates if the null hypothesis can be rejected - rejection of the hypothesis means that the difference between states is statistically significant at provided confidence level. The P-values can be adjusted using the provided method.

### Value

a [data.frame](#) object with calculated deuterium uptake difference in different forms with their uncertainty, P-value and  $-\log(P\text{-value})$  for the peptides from the provided data.

### References

Hageman, T. S. & Weis, D. D. Reliable Identification of Significant Differences in Differential Hydrogen Exchange-Mass Spectrometry Measurements Using a Hybrid Significance Testing Approach. *Anal Chem* 91, 8008–8016 (2019).

### See Also

[read\\_hdx](#) [calculate\\_exp\\_masses\\_per\\_replicate](#)

### Examples

```
p_diff_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat)
head(p_diff_uptake_dat)
```

---

```
create_p_diff_uptake_dataset_with_confidence
```

*Create differential dataset with statistical validity*

---

## Description

Create differential dataset with statistical validity

## Usage

```
create_p_diff_uptake_dataset_with_confidence(  
  p_diff_uptake_dat,  
  houde_interval = NULL,  
  houde_interval_times = NULL,  
  theoretical = FALSE,  
  fractional = FALSE  
)
```

## Arguments

<code>p_diff_uptake_dat</code>	differential uptake data alongside the P-value as calculated by <a href="#">create_p_diff_uptake_dataset</a>
<code>houde_interval</code>	interval value, as calculated by <a href="#">calculate_confidence_limit_values</a>
<code>houde_interval_times</code>	specified time points to be used in calculation
<code>theoretical</code>	logical, determines if values are theoretical
<code>fractional</code>	logical, determines if values are fractional

## Details

Function provides additional information on statistical relevance based on supplied data.

## Value

a `data.frame` object.

## See Also

[read\\_hdx](#) [create\\_p\\_diff\\_uptake\\_dataset](#) [calculate\\_confidence\\_limit\\_values](#)

## Examples

```
p_diff_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat)  
p_diff_uptake_dat_confidence <- create_p_diff_uptake_dataset_with_confidence(p_diff_uptake_dat)  
head(p_diff_uptake_dat_confidence)
```

---

```
create_quality_control_dataset
    Experiment quality control
```

---

### Description

Checks how the uncertainty changes in a function of maximal exchange control time of measurement.

### Usage

```
create_quality_control_dataset(  
  dat,  
  protein = dat[["Protein"]][1],  
  state_1 = unique(dat[["State"]])[1],  
  state_2 = unique(dat[["State"]])[2],  
  time_t = unique(dat[["Exposure"]])[2],  
  time_0 = min(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

### Arguments

<code>dat</code>	data imported by the <a href="#">read_hdx</a> function.
<code>protein</code>	chosen protein.
<code>state_1</code>	first biological state.
<code>state_2</code>	second biological state.
<code>time_t</code>	time point of the measurement for which the calculations are done.
<code>time_0</code>	minimal exchange control time point of measurement.
<code>deut_part</code>	deuterium percentage in solution used in experiment, value from range [0, 1].

### Details

The function calculates mean uncertainty of all peptides and its uncertainty (standard error) based on given 'time\_0' and 'time\_t' as a function of 'time\_100'. Both theoretical and experimental results for each state and their difference are supplied for comparison but only experimental calculations depends on 'time\_100' variable. The results are either in form of fractional or absolute values depending on the 'fractional' parameter supplied by the user. This data can be useful for general overview of the experiment and analyze of the chosen time parameters.

### Value

[data.frame](#) object with mean uncertainty per different 'time\_100' value. The values are shown as percentages.

**See Also**

[read\\_hdx](#) [calculate\\_state\\_uptake](#) [plot\\_quality\\_control](#) [show\\_quality\\_control\\_data](#)

**Examples**

```
create_quality_control_dataset(alpha_dat)
```

---

create\_replicate\_dataset

*Create replicates data*

---

**Description**

Create replicate data set suitable for replicate histogram, for one or multiple time points of measurement.

**Usage**

```
create_replicate_dataset(  
  dat,  
  time_t = NULL,  
  protein = unique(dat[["Protein"]])[1],  
  state = dat[["State"]][1]  
)
```

**Arguments**

dat	data as imported by the <a href="#">read_hdx</a> function.
time_t	optional, for only one time point of measurement. If value is NULL, all time point from dat are preserved.
protein	chosen protein.
state	biological state for chosen protein.

**Details**

The function [create\\_replicate\\_dataset](#) creates a dataset with information about how many replicates are for peptides in specific state in time points of measurement.

**Value**

a [data.frame](#) object.

**See Also**

[plot\\_replicate\\_histogram](#) [show\\_replicate\\_histogram\\_data](#)

## Examples

```
create_replicate_dataset(alpha_dat)
```

---

```
create_state_comparison_dataset
```

*Creates comparison uptake dataset*

---

## Description

Calculates deuterium uptake values for selected biological states in selected time point of measurements.

## Usage

```
create_state_comparison_dataset(  
  dat,  
  protein = unique(dat[["Protein"]])[1],  
  states = unique(dat[["State"]]),  
  time_0 = min(dat[["Exposure"]]),  
  time_t = unique(dat[["Exposure"]])[3],  
  time_100 = max(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

## Arguments

<code>dat</code>	data imported by the <a href="#">read_hdx</a> function.
<code>protein</code>	chosen protein.
<code>states</code>	vector of states (for chosen protein), for which the calculations are done.
<code>time_0</code>	minimal exchange control time point of measurement [min].
<code>time_t</code>	time point of the measurement for which the calculations are done [min].
<code>time_100</code>	maximal exchange control time point of measurement [min].
<code>deut_part</code>	deuterium percentage in solution used in experiment, value from range [0, 1].

## Details

Function [create\\_state\\_comparison\\_dataset](#) is a wrapper for [calculate\\_state\\_uptake](#) function, calls this function for all (default) or chosen states in states vector.

## Value

a `data.frame` object.

**See Also**

[read\\_hdx](#) [calculate\\_state\\_uptake](#)

**Examples**

```
comparison_dat <- create_state_comparison_dataset(alpha_dat)
head(comparison_dat)
```

---

```
create_state_uptake_dataset
```

*Create uptake dataset for chosen state*

---

**Description**

Calculates deuterium uptake values for one biological state.

**Usage**

```
create_state_uptake_dataset(  
  dat,  
  protein = unique(dat[["Protein"]])[1],  
  state = (dat[["State"]])[1],  
  time_0 = min(dat[dat[["Exposure"]] > 0, ][["Exposure"]]),  
  time_100 = max(dat[["Exposure"]]),  
  deut_part = 0.9  
)
```

**Arguments**

dat	data imported by the <a href="#">read_hdx</a> function.
protein	chosen protein.
state	biological state for chosen protein.
time_0	minimal exchange control time point of measurement [min].
time_100	maximal exchange control time point of measurement [min].
deut_part	deuterium percentage in solution used in experiment, value from range [0, 1].

**Details**

The function [create\\_uptake\\_dataset](#) generates a dataset with deuterium uptake values in different forms. For each peptide in chosen protein in chosen state for time points of measurement between minimal and maximal control time points of measurement deuterium uptake, fractional deuterium uptake with respect to controls or theoretical tabular values are calculated, with combined and propagated uncertainty. Each peptide has an ID, based on its start position. This data can be presented in a form of comparison plot, butterfly plot or chiclet plot.

**Value**

a `data.frame` object.

**See Also**

[read\\_hdx](#) [calculate\\_state\\_uptake](#)

**Examples**

```
state_uptake_dat <- create_state_uptake_dataset(alpha_dat)
head(state_uptake_dat)
```

---

`create_uptake_dataset` *Create uptake dataset for multiple states*

---

**Description**

Calculates deuterium uptake values for selected biological states in multiple time points of measurements.

**Usage**

```
create_uptake_dataset(
  dat,
  protein = unique(dat[["Protein"]])[1],
  states = unique(dat[["State"]]),
  time_0 = min(dat[["Exposure"]]),
  time_100 = max(dat[["Exposure"]]),
  deut_part = 0.9
)
```

**Arguments**

<code>dat</code>	data imported by the <a href="#">read_hdx</a> function.
<code>protein</code>	chosen protein.
<code>states</code>	list of biological states for chosen protein.
<code>time_0</code>	minimal exchange control time point of measurement [min].
<code>time_100</code>	maximal exchange control time point of measurement [min].
<code>deut_part</code>	deuterium percentage in solution used in experiment, value from range [0, 1].

**Details**

Function `create_uptake_dataset` generates a dataset with deuterium uptake values in different forms. For each peptide in chosen protein in chosen states for time points of measurement between minimal and maximal control time points of measurement deuterium uptake, fractional deuterium uptake with respect to controls or theoretical tabular values are calculated, with combined and propagated uncertainty. Each peptide has an ID, based on its start position. This function is a wrapper for `create_state_uptake_dataset` but for multiple states. The output of this function can be presented in a form of comparison plot.

**Value**

a `data.frame` object.

**See Also**

`read_hdx` `calculate_state_uptake` `create_state_uptake_dataset`

**Examples**

```
uptake_dat <- create_uptake_dataset(alpha_dat, states = c("Alpha_KSCN", "ALPHA_Gamma"))
head(uptake_dat)
```

---

get_n_replicates	<i>Get number of replicates</i>
------------------	---------------------------------

---

**Description**

Calculates the number of replicates from the experimental data.

**Usage**

```
get_n_replicates(dat, protein = dat[["Protein"]][1])
```

**Arguments**

dat	data imported by the <code>read_hdx</code> function.
protein	chosen protein

**Details**

Calculate the number of replicates of experiment.

**Value**

a `numeric` value.

**See Also**[read\\_hdx](#)**Examples**

```
get_n_replicates(alpha_dat)
```

---

```
get_peptide_sequence
```

*Get peptide sequence based on the position*

---

**Description**

Gets the peptide sequence based on selected parameters (start and end position, modification).

**Usage**

```
get_peptide_sequence(  
  dat,  
  protein = dat[["Protein"]][1],  
  start,  
  end,  
  modification = FALSE  
)
```

**Arguments**

dat	any data frame that contains following information: protein, sequence, start, end, modification.
protein	chosen protein.
start	start position of the peptide of interest.
end	end position of the peptide of interest.
modification	logical value to indicate if peptide of interest has modification or not.

**Details**

Function returns peptide sequence for selected parameters. Peptide sequence is often required to properly identify peptide of interest, and to avoid mistakes sequence is returned by the function. Moreover, function uses the modification value to select peptide sequence.

**Value**

a [character](#) value.

**See Also**[read\\_hdx](#)

**Examples**

```
get_peptide_sequence(alpha_dat, start = 1, end = 11)
```

---

get\_protein\_coverage *Get protein coverage*

---

**Description**

Calculate protein coverage by the peptides in selected biological state or states.

**Usage**

```
get_protein_coverage(  
  dat,  
  protein = dat[["Protein"]][1],  
  states = unique(dat[["State"]]),  
  protein_length = NULL  
)
```

**Arguments**

dat	data imported by the <a href="#">read_hdx</a> function
protein	selected protein
states	selected biological states
protein_length	<a href="#">numeric</a> , indicates the length of the protein. If not provided, the maximal end value from the file is used.

**Details**

Function [get\\_protein\\_coverage](#) calculates the percentage coverage of the protein sequence, rounded to two decimal places.

**Value**

a [numeric](#) percentage value (rounded to two decimal places).

**See Also**

[read\\_hdx](#)

**Examples**

```
get_protein_coverage(alpha_dat)  
get_protein_coverage(alpha_dat, protein_length = 230)
```

---

`get_protein_redundancy`*Get protein redundancy*

---

**Description**

Calculates the protein redundancy in the whole experiment (all biological states).

**Usage**

```
get_protein_redundancy(dat, protein_length = NULL)
```

**Arguments**

`dat` data imported by the [read\\_hdx](#) function.

`protein_length` [numeric](#), indicates the length of the protein. If not provided, the maximal end value from the file is used.

**Details**

Function [get\\_protein\\_redundancy](#) calculates the redundancy of the protein, based on provided experimental data.

**Value**

a [numeric](#) value.

**See Also**

[read\\_hdx](#)

**Examples**

```
get_protein_redundancy(alpha_dat)
```

---

get\_replicate\_list\_sd *Get replicates sd*

---

## Description

Get list of peptides with their standard deviation.

## Usage

```
get_replicate_list_sd(  
  dat,  
  protein = dat[["Protein"]][1],  
  state = dat[["State"]][1],  
  time_t = unique(dat[["Exposure"]])[3]  
)
```

## Arguments

dat	data as imported by the <a href="#">read_hdx</a> function.
protein	chosen protein.
state	biological state for chosen protein.
time_t	time point of the measurement.

## Details

Function gets the peptide list in selected state with their standard deviation of measurement, calculated from the technical replicates. It is used for selection for measurement variability plot.

## Value

a [data.frame](#) object.

## See Also

[create\\_replicate\\_dataset](#)

## Examples

```
get_replicate_list_sd(alpha_dat)
```

---

get\_residue\_positions *Get residue positions*

---

**Description**

Get residue positions

**Usage**

```
get_residue_positions(dat)
```

**Arguments**

dat                    data imported by the [read\\_hdx](#) function.

**Details**

Prepares data table for high-resolution values. It creates a long data frame with each position in the sequence accompanied by the amino acid placed in this position.

**Value**

a data.frame object.

**Examples**

```
# disabled due to long execution time  
get_residue_positions(alpha_dat)
```

---

get\_structure\_color *Get color palette for 3D structure*

---

**Description**

This function provides a set of color codes associated with aggregated values to be presented on 3D structure.

**Usage**

```
get_structure_color(  
  aggregated_dat,  
  differential = FALSE,  
  fractional = TRUE,  
  theoretical = FALSE,  
  time_t  
)
```

**Arguments**

aggregated\_dat aggregated data, either for single uptake or differential  
differential indicator if the aggregated\_dat contains differential results  
fractional indicator if fractional values are used  
theoretical indicator if theoretical values are used  
time\_t time point from aggregated\_dat to be shown on the structure

**Value**

a list

**Examples**

```
# disabled due to long execution time

diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
diff_aggregated_dat <- create_aggregated_diff_uptake_dataset(diff_uptake_dat)
get_structure_color(diff_aggregated_dat,
                    differential = TRUE,
                    time_t = 1)
```

---

HaDeXify

*HaDeX customized ggplot theme*

---

**Description**

This function HaDeXifies plot. It adds HaDeX logo and ggplot theme.

**Usage**

```
HaDeXify(plt)
```

**Arguments**

plt ggplot object. Plot to HaDeXify.

**Details**

Function adds the logo of HaDeX package in the left down corner of the plot and the HaDeX theme.

**Value**

a `ggplot` object.

**See Also**

[read\\_hdx](#) [plot\\_differential](#) [plot\\_butterfly](#)

**Examples**

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
HaDeXify(plot_differential(diff_uptake_dat))
```

---

HaDeX\_GUI

*HaDeX Graphical User Interface*

---

**Description**

Shows how to launch graphical user interface from HaDeXGUI package. If the GUI package is not installed, it asks user whether to install it.

**Usage**

```
HaDeX_GUI()
```

**Value**

No return value, called for side effects

**Warning**

Any ad-blocking software may cause malfunctions.

---

install\_GUI

*Installs GUI package from GitHub*

---

**Description**

Installs GUI package from GitHub

**Usage**

```
install_GUI()
```

**Value**

No return value, called for side effects

---

is_GUI_installed	<i>Checks if GUI package is installed</i>
------------------	---

---

**Description**

Indicates presence or absence of HaDeXGUI package.

**Usage**

```
is_GUI_installed()
```

**Value**

logical value indicating availability of GUI package

---

plot_aggregated_differential_uptake	<i>Plots aggregated uptake difference</i>
-------------------------------------	---

---

**Description**

Plots aggregated uptake difference

**Usage**

```
plot_aggregated_differential_uptake(
  aggregated_diff_dat,
  fractional = TRUE,
  theoretical = FALSE,
  time_100 = max(unique(aggregated_diff_dat[["Exposure"]])),
  panels = FALSE,
  interactive = FALSE
)
```

**Arguments**

aggregated_diff_dat	aggregated differential uptake data as calculated by <a href="#">create_aggregated_diff_uptake_dataset</a>
fractional	logical, determines if values are fractional
theoretical	logical, determines if values are theoretical
time_100	maximal exchange control time point of measurement [min]
panels	logical, indicator if plot should be divided into panels or not
interactive	logical, whether plot should have an interactive layer created with with <code>ggigraph</code> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app)

**Value**

a `ggplot` object

**See Also**

[create\\_diff\\_uptake\\_dataset](#) [create\\_aggregated\\_diff\\_uptake\\_dataset](#)

**Examples**

```
# disabled due to long execution time

diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
aggregated_diff_dat <- create_aggregated_diff_uptake_dataset(diff_uptake_dat)
plot_aggregated_differential_uptake(aggregated_diff_dat, panels = FALSE)
plot_aggregated_differential_uptake(aggregated_diff_dat, fractional = FALSE,
                                   theoretical = TRUE, panels = FALSE)
plot_aggregated_differential_uptake(aggregated_diff_dat, theoretical = TRUE,
                                   panels = TRUE)
```

---

plot\_aggregated\_uptake

*Plots aggregated uptake*

---

**Description**

Plots aggregated uptake

**Usage**

```
plot_aggregated_uptake(
  aggregated_dat,
  fractional = TRUE,
  theoretical = FALSE,
  time_100 = max(unique(aggregated_dat[["Exposure"]])),
  panels = FALSE,
  interactive = FALSE
)
```

**Arguments**

aggregated_dat	aggregated differential uptake data as calculated by <a href="#">create_aggregated_uptake_dataset</a>
fractional	logical, determines if values are fractional
theoretical	logical, determines if values are theoretical
time_100	maximal exchange control time point of measurement [min]
panels	logical, indicator if plot should be divided into panels or not

`interactive` logical, whether plot should have an interactive layer created with `gginter`, which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

### Value

a `ggplot` object

### Examples

```
# disabled due to long execution time

kin_dat <- create_uptake_dataset(alpha_dat, states = "Alpha_KSCN")
aggregated_dat <- create_aggregated_uptake_dataset(kin_dat)
plot_aggregated_uptake(aggregated_dat, panels = FALSE)
plot_aggregated_uptake(aggregated_dat, fractional = FALSE, panels = FALSE)
plot_aggregated_uptake(aggregated_dat, fractional = FALSE, theoretical = TRUE, panels = TRUE)
```

---

plot\_aggregated\_uptake\_structure

*Plot aggregated uptake on the 3D structure*

---

### Description

Function plots the aggregated data (either one state deuterium uptake or differential deuterium uptake) on the 3d structure.

### Usage

```
plot_aggregated_uptake_structure(
  aggregated_dat,
  differential = FALSE,
  fractional = TRUE,
  theoretical = FALSE,
  time_t,
  pdb_file_path
)
```

### Arguments

`aggregated_dat` aggregated data, either for single uptake or differential  
`differential` indicator if the `aggregated_dat` contains differential results  
`fractional` indicator if fractional values are used  
`theoretical` indicator if theoretical values are used  
`time_t` time point from `aggregated_dat` to be shown on the structure  
`pdb_file_path` file path to the pdb file

**Value**

a r3dmol object.

**Examples**

```
library(shiny)
# disabled due to its long time and producing 3rdmol object

diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
diff_aggregated_dat <- create_aggregated_diff_uptake_dataset(diff_uptake_dat)
pdb_file_path <- system.file(package = "HaDeX2", "HaDeX/data/Model_eEF1Balpha.pdb")

plot_aggregated_uptake_structure(diff_aggregated_dat,
                                differential = TRUE,
                                time_t = 1,
                                pdb_file_path = pdb_file_path)

kin_dat <- create_uptake_dataset(alpha_dat, states = "Alpha_KSCN" )
aggregated_dat <- create_aggregated_uptake_dataset(kin_dat)
plot_aggregated_uptake_structure(aggregated_dat,
                                differential = FALSE,
                                time_t = 1,
                                pdb_file_path = pdb_file_path)
```

---

```
plot_amino_distribution
      generate_amino_distribution
```

---

**Description**

Generates amino distribution based on the protein sequence and shows if the amino acid is hydrophobic or hydrophylic.

**Usage**

```
plot_amino_distribution(
  position_in_sequence,
  hydro_properties,
  protein,
  charge_colors,
  interactive = getOption("hadex_use_interactive_plots")
)
```

**Arguments**

position_in_sequence	custom format
hydro_properties	data with hydrophobic properties
protein	chosen protein
charge_colors	vector of desired colors
interactive	logical, whether plot should have an interactive layer created with with ggigraph, which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

**Details**

The data for this function is not packaged yet.

**Value**

a `ggplot` object.

---

plot_butterfly	<i>Butterfly deuterium uptake plot</i>
----------------	--

---

**Description**

Butterfly plot of deuterium uptake values in time for one biological state.

**Usage**

```
plot_butterfly(
  uptake_dat,
  theoretical = FALSE,
  fractional = FALSE,
  uncertainty_type = "ribbon",
  interactive = getOption("hadex_use_interactive_plots")
)
```

**Arguments**

uptake_dat	data produced by <code>create_state_uptake_dataset</code> function.
theoretical	logical, determines if values are theoretical.
fractional	logical, determines if values are fractional.
uncertainty_type	type of presenting uncertainty, possible values: "ribbon", "bars" or "bars + line".
interactive	logical, whether plot should have an interactive layer created with with ggigraph, which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

**Details**

Function `plot_butterfly` generates butterfly plot based on provided data and parameters. On X-axis there is peptide ID. On the Y-axis there is deuterium uptake in chosen form. Data from multiple time points of measurement is presented.

**Value**

a `ggplot` object.

**See Also**

`create_state_uptake_dataset`

**Examples**

```
state_uptake_dat <- create_state_uptake_dataset(alpha_dat)
plot_butterfly(state_uptake_dat)
```

---

plot\_chiclet

*Chiclet deuterium uptake plot*

---

**Description**

Chiclet plot of deuterium uptake values in time for one biological state.

**Usage**

```
plot_chiclet(
  uptake_dat,
  theoretical = FALSE,
  fractional = FALSE,
  show_uncertainty = FALSE,
  interactive = getOption("hadex_use_interactive_plots")
)
```

**Arguments**

<code>uptake_dat</code>	produced by <code>create_state_uptake_dataset</code> function.
<code>theoretical</code>	logical, determines if values are theoretical.
<code>fractional</code>	logical, determines if values are fractional.
<code>show_uncertainty</code>	logical, determines if the uncertainty is shown.
<code>interactive</code>	logical, whether plot should have an interactive layer created with <code>gginter</code> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

**Details**

Function `plot_chiclet` produces a chiclet plot based on the same dataset as butterfly plot, as it is the different form of presenting the same data. On X-axis there is a peptide ID. On Y-axis are time points of measurement. Each tile for a peptide in time has a color value representing the deuterium uptake, in a form based on provided criteria (e.g. fractional). Each tile has a plus sign, which size represent the uncertainty of measurement for chosen value.

**Value**

a `ggplot` object.

**See Also**

`create_state_uptake_dataset`

**Examples**

```
state_uptake_dat <- create_state_uptake_dataset(alpha_dat)
plot_chiclet(state_uptake_dat)
```

---

plot\_coverage

*Peptide coverage*

---

**Description**

Plot the peptide coverage of the protein sequence

**Usage**

```
plot_coverage(
  dat,
  protein = dat[["Protein"]][1],
  states = NULL,
  show_blanks = TRUE,
  interactive = getOption("hadex_use_interactive_plots")
)
```

**Arguments**

dat	data imported by the <code>read_hdx</code> function
protein	selected protein
states	selected biological states for given protein
show_blanks	logical, indicator if the non-covered regions of the sequence are indicated in red
interactive	logical, whether plot should have an interactive layer created with with <code>ggigraph</code> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app)

### Details

The function `plot_coverage` generates sequence coverage plot based on experimental data for selected protein in selected biological states. Only non-duplicated peptides are shown on the plot, next to each other.

The aim of this plot is to see the dependence between position of the peptide on the protein sequence. Their position on y-axis does not contain any information.

### Value

a `ggplot` object

### See Also

[read\\_hdx\\_plot\\_position\\_frequency](#)

### Examples

```
plot_coverage(alpha_dat)
plot_coverage(alpha_dat, show_blanks = FALSE)

diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
plot_coverage(diff_uptake_dat)
```

---

plot\_coverage\_heatmap *Coverage heatmap*

---

### Description

Coverage heatmap with color indicating specific value

### Usage

```
plot_coverage_heatmap(
  x_dat,
  protein = x_dat[["Protein"]][1],
  state = NULL,
  value = NULL,
  time_t = NULL,
  interactive = getOption("hadex_use_interactive_plots")
)
```

### Arguments

<code>x_dat</code>	data created using <code>calculate_</code> or <code>create_</code> function
<code>protein</code>	selected protein
<code>state</code>	selected biological state

value	value to be presented
time_t	chosen time point
interactive	logical, whether plot should have an interactive layer created with with ggigraph, which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app)

### Details

Plots standard protein coverage but colored with desired value.

### Value

a [ggplot](#) object

### See Also

[read\\_hdx\\_plot\\_coverage](#)

### Examples

```
uptake_dat <- create_uptake_dataset(alpha_dat, states = "Alpha_KSCN")
plot_coverage_heatmap(uptake_dat)
plot_coverage_heatmap(x_dat = uptake_dat, value = "frac_deut_uptake", time_t = 0.167)
plot_coverage_heatmap(uptake_dat, value = "err_frac_deut_uptake", time_t = 0.167)

diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
plot_coverage_heatmap(diff_uptake_dat)
plot_coverage_heatmap(diff_uptake_dat, value = "diff_frac_deut_uptake")
plot_coverage_heatmap(diff_uptake_dat, value = "diff_frac_deut_uptake", time_t = 0.167)
plot_coverage_heatmap(diff_uptake_dat, value = "err_diff_frac_deut_uptake", time_t = 0.167)

auc_dat <- calculate_auc(create_uptake_dataset(alpha_dat))
plot_coverage_heatmap(auc_dat, value = "auc")

bex_dat <- calculate_back_exchange(alpha_dat, state = "Alpha_KSCN")
plot_coverage_heatmap(bex_dat, value = "back_exchange")
```

---

plot\_differential      *Differential plot*

---

### Description

Woods plot of differential deuterium uptake values between two biological states in time point.

**Usage**

```

plot_differential(
  diff_uptake_dat = NULL,
  diff_p_uptake_dat = NULL,
  skip_amino = 0,
  time_t = NULL,
  theoretical = FALSE,
  fractional = FALSE,
  show_houde_interval = FALSE,
  hide_houde_insignificant = FALSE,
  show_tstud_confidence = FALSE,
  hide_tstud_insignificant = FALSE,
  confidence_level = 0.98,
  all_times = FALSE,
  line_size = 1.5,
  interactive = getOption("hadex_use_interactive_plots")
)

```

**Arguments**

diff_uptake_dat	produced by <a href="#">calculate_diff_uptake</a> or <a href="#">create_diff_uptake_dataset</a> function.
diff_p_uptake_dat	produced by <a href="#">create_p_diff_uptake_dataset</a> function.
skip_amino	integer, indicator how many amino acids from the N-terminus should be omitted
time_t	time point of measurement, if only one should be displayed on the plot.
theoretical	logical, determines if values are theoretical.
fractional	logical, determines if values are fractional.
show_houde_interval	logical, determines if houde interval is shown.
hide_houde_insignificant	logical, determines if statistically insignificant values using Houde test are hidden on the plot.
show_tstud_confidence	logical, determines if t-Student test validity is shown.
hide_tstud_insignificant	logical, determines if statistically insignificant values using t-Student test are hidden on the plot.
confidence_level	confidence level for the test, from range [0, 1].
all_times	logical, determines if all the time points from the supplied data should be displayed on the plots next to each other.
line_size	line size of the lines displayed on the plot.

`interactive` logical, whether plot should have an interactive layer created with `gginter`, which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

## Details

Function `plot_differential` presents provided data in a form of differential (Woods) plot. The plot shows difference in exchange for two biological states, selected in generation of dataset at one time point of measurement. On X-axis there is a position in a sequence, with length of a segment of each peptide representing its length. On Y-axis there is deuterium uptake difference in chosen form. Error bars represents the combined and propagated uncertainty. For Woods Plot there is available Houde test and t-Student test to see the statistically significant peptides. Selecting both of them simultaneously results in hybrid testing, as described in Weis et al. The statistically significant values are in color (red if the difference is positive, blue if negative), and the insignificant values are grey.

## Value

a `[ggplot2::ggplot()]` object.

## References

Hageman, T. S. & Weis, D. D. Reliable Identification of Significant Differences in Differential Hydrogen Exchange-Mass Spectrometry Measurements Using a Hybrid Significance Testing Approach. *Anal Chem* 91, 8008–8016 (2019).

## See Also

[create\\_diff\\_uptake\\_dataset](#) [calculate\\_diff\\_uptake](#) [show\\_diff\\_uptake\\_data](#)

## Examples

```
# disabled due to long execution time

diff_uptake_dat <- calculate_diff_uptake(alpha_dat,
                                       states = c("ALPHA_Gamma", "Alpha_KSCN"), time_t = 0.167)
plot_differential(diff_uptake_dat = diff_uptake_dat, time_t = 0.167)
plot_differential(diff_uptake_dat = diff_uptake_dat, time_t = 0.167, show_houde_interval = TRUE)
plot_differential(diff_uptake_dat = diff_uptake_dat, time_t = 0.167, skip_amino = 0)
plot_differential(diff_uptake_dat = diff_uptake_dat, time_t = 0.167, line_size = 1)
plot_differential(diff_uptake_dat = diff_uptake_dat, all_times = TRUE)
plot_differential(diff_uptake_dat = diff_uptake_dat, all_times = TRUE, show_houde_interval = TRUE)
plot_differential(diff_uptake_dat = diff_uptake_dat, all_times = TRUE, show_houde_interval = TRUE,
                 hide_houde_insignificant = TRUE)

diff_p_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat, state_1 = "Alpha_KSCN",
                                                state_2 = "ALPHA_Gamma")
plot_differential(diff_p_uptake_dat = diff_p_uptake_dat, all_times = TRUE,
                 show_tstud_confidence = TRUE)
plot_differential(diff_p_uptake_dat = diff_p_uptake_dat, all_times = FALSE,
                 time_t = 0.167, show_tstud_confidence = TRUE, show_houde_interval = TRUE)
```

```

plot_differential(diff_p_uptake_dat = diff_p_uptake_dat, show_tstud_confidence = TRUE,
                 show_houde_interval = TRUE, all_times = FALSE)
plot_differential(diff_p_uptake_dat = diff_p_uptake_dat, show_tstud_confidence = TRUE,
                 show_houde_interval = TRUE, all_times = FALSE, hide_houde_insignificant = TRUE)
plot_differential(diff_p_uptake_dat = diff_p_uptake_dat, show_tstud_confidence = TRUE,
                 show_houde_interval = TRUE, all_times = FALSE, hide_houde_insignificant = TRUE,
                 hide_tstud_insignificant = TRUE)

```

---

plot\_differential\_butterfly

*Butterfly differential deuterium uptake plot*

---

### Description

Butterfly plot of differential deuterium uptake values between two biological states in time.

### Usage

```

plot_differential_butterfly(
  diff_uptake_dat = NULL,
  diff_p_uptake_dat = NULL,
  theoretical = FALSE,
  fractional = FALSE,
  show_houde_interval = FALSE,
  show_tstud_confidence = FALSE,
  uncertainty_type = "ribbon",
  confidence_level = 0.98,
  interactive = getOption("hadex_use_interactive_plots")
)

```

### Arguments

diff_uptake_dat	data produced by <a href="#">calculate_diff_uptake</a> or <a href="#">create_diff_uptake_dataset</a> function
diff_p_uptake_dat	differential uptake data alongside the P-value as calculated by <a href="#">create_p_diff_uptake_dataset</a>
theoretical	logical, determines if values are theoretical
fractional	logical, determines if values are fractional
show_houde_interval	logical, determines if houde interval is shown
show_tstud_confidence	logical, determines if t-Student test validity is shown
uncertainty_type	type of presenting uncertainty, possible values: "ribbon", "bars" or "bars + line"

confidence_level	confidence level for the test, from range [0, 1] Important if selected show_confidence_limit
interactive	logical, whether plot should have an interactive layer created with with ggigraph, which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

### Details

Function `plot_differential_butterfly` generates differential butterfly plot based on provided data and parameters. On X-axis there is peptide ID. On the Y-axis there is deuterium uptake difference in chosen form. Data from multiple time points of measurement is presented. If chosen, there are confidence limits based on Houde test on provided confidence level.

### Value

a `[ggplot2::ggplot()]` object.

### References

Houde, D., Berkowitz, S.A., and Engen, J.R. (2011). The Utility of Hydrogen/Deuterium Exchange Mass Spectrometry in Biopharmaceutical Comparability Studies. *J Pharm Sci* 100, 2071–2086.

### See Also

[read\\_hdx](#) [create\\_diff\\_uptake\\_dataset](#) [calculate\\_diff\\_uptake](#)

### Examples

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
plot_differential_butterfly(diff_uptake_dat = diff_uptake_dat)
plot_differential_butterfly(diff_uptake_dat = diff_uptake_dat, show_houde_interval = TRUE)

diff_p_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat)
plot_differential_butterfly(diff_p_uptake_dat = diff_p_uptake_dat, show_tstud_confidence = TRUE)
```

---

plot\_differential\_chiclet

*Chiclet differential deuterium uptake plot*

---

### Description

Chiclet plot of differential deuterium uptake values between two biological states in time.

**Usage**

```
plot_differential_chiclet(
  diff_uptake_dat = NULL,
  diff_p_uptake_dat = NULL,
  theoretical = FALSE,
  fractional = FALSE,
  show_houde_interval = FALSE,
  show_tstud_confidence = FALSE,
  confidence_level = 0.98,
  show_uncertainty = FALSE,
  interactive = getOption("hadex_use_interactive_plots")
)
```

**Arguments**

diff_uptake_dat	data produced by <a href="#">calculate_diff_uptake</a> or <a href="#">create_diff_uptake_dataset</a> function
diff_p_uptake_dat	differential uptake data alongside the P-value as calculated by <a href="#">create_p_diff_uptake_dataset</a>
theoretical	logical, determines if values are theoretical
fractional	logical, determines if values are fractional
show_houde_interval	logical, determines if houde interval is shown
show_tstud_confidence	logical, determines if t-Student test validity is shown
confidence_level	confidence level for the test, from range [0, 1] Important if selected show_confidence_limit
show_uncertainty	logical, determines if the uncertainty is shown
interactive	logical, whether plot should have an interactive layer created with with <a href="#">ggigraph</a> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

**Details**

Function [plot\\_differential\\_chiclet](#) generates chiclet differential plot based on provided data and parameters. On X-axis there is a peptide ID. On Y-axis are time points of measurement. Each tile for a peptide in time has a color value representing the deuterium uptake difference between chosen states, in a form based on provided criteria (e.g. fractional). Each tile has a plus sign, which size represent the uncertainty of measurement for chosen value.

**Value**

a `[ggplot2::ggplot()]` object.

**See Also**

[create\\_diff\\_uptake\\_dataset](#) [calculate\\_diff\\_uptake](#)

**Examples**

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
plot_differential_chiclet(diff_uptake_dat)
plot_differential_chiclet(diff_uptake_dat, show_houde_interval = TRUE)

diff_p_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat)
plot_differential_chiclet(diff_p_uptake_dat = diff_p_uptake_dat,
                        show_tstud_confidence = TRUE)
plot_differential_chiclet(diff_p_uptake_dat = diff_p_uptake_dat,
                        show_tstud_confidence = TRUE, show_houde_interval = TRUE)
```

---

```
plot_differential_uptake_curve
      Plot differential uptake curve
```

---

**Description**

Differential uptake curve for one peptide between two biological states.

**Usage**

```
plot_differential_uptake_curve(
  diff_uptake_dat = NULL,
  diff_p_uptake_dat = NULL,
  sequence = NULL,
  theoretical = FALSE,
  fractional = FALSE,
  uncertainty_type = "ribbon",
  log_x = TRUE,
  show_houde_interval = FALSE,
  show_tstud_confidence = FALSE,
  interactive = getOption("hadex_use_interactive_plots")
)
```

**Arguments**

diff_uptake_dat	produced by <a href="#">calculate_diff_uptake</a> or <a href="#">create_diff_uptake_dataset</a> function
diff_p_uptake_dat	differential uptake data alongside the P-value as calculated by <a href="#">create_p_diff_uptake_dataset</a>
sequence	sequence of chosen peptide

<code>theoretical</code>	logical, determines if plot shows theoretical values
<code>fractional</code>	logical, determines if plot shows fractional values
<code>uncertainty_type</code>	type of presenting uncertainty, possible values: "ribbon", "bars" or "bars + line"
<code>log_x</code>	logical, determines if x axis shows logarithmic values
<code>show_houde_interval</code>	logical, determines if houde interval is shown
<code>show_tstud_confidence</code>	logical, determines if t-Student test validity is shown
<code>interactive</code>	logical, whether plot should have an interactive layer created with with <code>ggigraph</code> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

### Details

This plot shows the differential deuterium uptake between two biological states for selected peptides in different time points. The possibility to plot multiple differences (between state and mutant) for the peptide will be added soon.

### Value

a `ggplot` object.

### See Also

[read\\_hdx](#) [create\\_diff\\_uptake\\_dataset](#) [calculate\\_diff\\_uptake](#)

### Examples

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
plot_differential_uptake_curve(diff_uptake_dat = diff_uptake_dat, sequence = "GDLKSPAGL")

diff_p_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat)
plot_differential_uptake_curve(diff_p_uptake_dat = diff_p_uptake_dat,
                             sequence = "GDLKSPAGL", show_houde_interval = TRUE)
plot_differential_uptake_curve(diff_p_uptake_dat = diff_p_uptake_dat,
                             sequence = "GDLKSPAGL", show_houde_interval = TRUE,
                             show_tstud_confidence = TRUE)
plot_differential_uptake_curve(diff_p_uptake_dat = diff_p_uptake_dat,
                             sequence = "GDLKSPAGL", show_tstud_confidence = TRUE)
```

---

plot_manhattan	<i>Manhattan plot</i>
----------------	-----------------------

---

### Description

Manhattan plot with p-values from the t-Student test and peptide position.

### Usage

```
plot_manhattan(
  p_dat,
  skip_amino = 0,
  plot_title = NULL,
  separate_times = TRUE,
  times = NULL,
  confidence_level = NULL,
  show_confidence_limit = TRUE,
  show_peptide_position = FALSE,
  interactive = getOption("hadex_use_interactive_plots")
)
```

### Arguments

p_dat	data produced by the <a href="#">create_p_diff_uptake_dataset</a> function.
skip_amino	integer, indicator how many amino acids from the N-terminus should be omitted
plot_title	title for the plot. If not provided, it is constructed in a form: "Differences between state_1 and state_2"
separate_times	logical, indicates if the data should be seen on the same plot, or on separate plots for each time point of measurement.
times	vector of time points of measurements to be included in the plot.
confidence_level	confidence level for the test, from range [0, 1].
show_confidence_limit	logical, indicates if the hybrid testing confidence intervals are shown.
show_peptide_position	logical, indicates if the peptide length and position in the sequence is shown. Otherwise, the peptides are represented by their ID.
interactive	logical, whether plot should have an interactive layer created with with <a href="#">ggigraph</a> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

## Details

The manhattan plot presents the P-values from t-student test, to see the regions of the protein with statistically significant changes between two biological states. On X-axis there is a position in a sequence, with length of a segment of each peptide representing its length. On Y-axis there is P-value from t-Student test.

## Value

a [ggplot](#) object.

## References

Hageman, T. S. & Weis, D. D. Reliable Identification of Significant Differences in Differential Hydrogen Exchange-Mass Spectrometry Measurements Using a Hybrid Significance Testing Approach. *Anal Chem* 91, 8008–8016 (2019).

## See Also

[read\\_hdx](#) [create\\_p\\_diff\\_uptake\\_dataset](#)

## Examples

```
p_dat <- create_p_diff_uptake_dataset(alpha_dat)
plot_manhattan(p_dat)
plot_manhattan(p_dat, separate_times = FALSE)
plot_manhattan(p_dat, show_peptide_position = TRUE, separate_times = FALSE)
plot_manhattan(p_dat, separate_times = FALSE, show_confidence_limit = FALSE)
```

---

plot\_overlap

*Plot overlap data*

---

## Description

Generates overlapping peptide plot based on supplied data and parameters.

## Usage

```
plot_overlap(dat, protein = dat[["Protein"]][1], state = dat[["State"]][1])
```

## Arguments

dat	data imported by the <a href="#">read_hdx</a> function.
protein	protein included in calculations
state	state included in calculations

**Details**

The overlap plot presents all the peptides in given state on the protein sequence. This plot is visible in GUI.

**Value**

a `ggplot` object.

**See Also**

`read_hdx show_overlap_data`

**Examples**

```
plot_overlap(alpha_dat)
```

---

plot\_overlap\_distribution  
*Plot overlap distribution*

---

**Description**

Generates overlap distribution plot based on supplied data and parameters.

**Usage**

```
plot_overlap_distribution(  
  overlap_dist_dat,  
  start = 1,  
  end = max(overlap_dist_dat[["pos"]]),  
  interactive = getOption("hadex_use_interactive_plots")  
)
```

**Arguments**

overlap_dist_dat	produced by <code>create_overlap_distribution_dataset</code> function
start	start position of chosen protein.
end	end position of chosen protein.
interactive	logical, whether plot should have an interactive layer created with <code>gginter</code> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

**Details**

This plot presents how many times (by how many peptides) a amino position in protein sequence is covered. This plot is visible in GUI.

**Value**

a `ggplot` object.

**See Also**

[read\\_hdx](#) [reconstruct\\_sequence](#) [create\\_overlap\\_distribution\\_dataset](#)

**Examples**

```
overlap_dist_dat <- create_overlap_distribution_dataset(alpha_dat)
plot_overlap_distribution(overlap_dist_dat)
```

---

plot\_peptide\_charge\_measurement

*Plot peptide charge measurement*

---

**Description**

Plot the charge measurements from replicates for peptide in specific time point.

**Usage**

```
plot_peptide_charge_measurement(
  dat,
  protein = dat[["Protein"]][1],
  state = dat[["State"]][1],
  sequence = dat[["Sequence"]][1],
  time_t = unique(dat[["Exposure"]])[3]
)
```

**Arguments**

<code>dat</code>	data as imported by the <a href="#">read_hdx</a> function.
<code>protein</code>	chosen protein.
<code>state</code>	biological state for chosen protein.
<code>sequence</code>	sequence of chosen peptide.
<code>time_t</code>	time point of the measurement.

**Details**

This function shows the measurements of charge from different replicates for specific peptide in specific state in specific time point of measurement on the plot.

**Value**

a `[ggplot2::ggplot()]` object.

**See Also**

[read\\_hdx show\\_peptide\\_charge\\_measurement](#)

**Examples**

```
plot_peptide_charge_measurement(alpha_dat)
```

---

```
plot_peptide_mass_measurement
Plot peptide mass measurement
```

---

**Description**

Plot the mass measurements from replicates for peptide in specific time point.

**Usage**

```
plot_peptide_mass_measurement(
  dat,
  protein = dat[["Protein"]][1],
  state = dat[["State"]][1],
  sequence = dat[["Sequence"]][1],
  show_charge_values = TRUE,
  time_t = unique(dat[["Exposure"]])[3],
  interactive = getOption("hadex_use_interactive_plots")
)
```

**Arguments**

dat	data produced by <a href="#">read_hdx</a> function
protein	chosen protein
state	biological state for chosen protein
sequence	sequence of chosen peptide
show_charge_values	logical, indicator if the data is shown for replicates or for charge values within the replicates
time_t	chosen time point
interactive	logical, whether plot should have an interactive layer created with with <a href="#">ggigraph</a> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app)

### Details

This function shows the measurements of mass from different replicates for specific peptide in specific state in specific time point of measurement on the plot. Moreover, on the plot is shown the average mass from the replicates, used later in calculations. The ribbon next to the dotted average mass indicates the uncertainty.

### Value

a `[ggplot2::ggplot()]` object.

### See Also

[read\\_hdx](#) [calculate\\_exp\\_masses\\_per\\_replicate](#) [calculate\\_exp\\_masses](#) [calculate\\_state\\_uptake](#)  
[calculate\\_diff\\_uptake](#)

### Examples

```
plot_peptide_mass_measurement(alpha_dat, sequence = "FGSDDEEESEEAKRLRE")  
plot_peptide_mass_measurement(alpha_dat, sequence = "FGSDDEEESEEAKRLRE", show_charge_values = FALSE)
```

---

plot\_position\_frequency  
*Position frequency*

---

### Description

Plot the frequency of coverage for protein sequence

### Usage

```
plot_position_frequency(  
  dat,  
  protein = dat[["Protein"]][1],  
  state = dat[["State"]][1]  
)
```

### Arguments

dat	data as imported by the <a href="#">read_hdx</a> function
protein	selected protein
state	selected biological state for given protein

### Details

The function `plot_position_frequency` generates a histogram of the coverage frequency in selected biological states for selected protein. The position frequency plot presents how many times each position of the sequence is covered by different peptides.

**Value**

a [ggplot](#) object.

**See Also**

[read\\_hdx](#) [plot\\_coverage](#)

**Examples**

```
plot_position_frequency(alpha_dat)
```

---

`plot_quality_control` *Plot quality control data*

---

**Description**

Generates quality control plot based on supplied data.

**Usage**

```
plot_quality_control(qc_dat)
```

**Arguments**

`qc_dat` data produced by [create\\_quality\\_control\\_dataset](#) function, scaled if necessary.

**Details**

This plot presents the mean uncertainty in function of selected maximal exchange control time of measurement. This plot is visible in GUI.

**Value**

a [`ggplot2::ggplot()`] object.

**See Also**

[create\\_quality\\_control\\_dataset](#) [show\\_quality\\_control\\_data](#)

**Examples**

```
qc_dat <- create_quality_control_dataset(alpha_dat)
plot_quality_control(qc_dat)
```

---

plot\_replicate\_histogram  
*Plot replicates histogram*

---

### Description

Plot histogram on number of replicates per peptide in one or multiple time point of measurement.

### Usage

```
plot_replicate_histogram(  
  rep_dat,  
  time_points = FALSE,  
  interactive = getOption("hadex_use_interactive_plots")  
)
```

### Arguments

rep_dat	replicate data, created by <a href="#">create_replicate_dataset</a> function.
time_points	logical, indicator if the histogram should show values aggregated for time points of measurements.
interactive	logical, whether plot should have an interactive layer created with <code>gggraph</code> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

### Details

The function shows three versions of replicate histogram, based on supplied `rep_dat` and `time_points`. If `time_points` is selected, the histogram shows the number of replicates for time points of measurement, to spot if there were troubles with samples for specific time point of measurement. Then, on the X-axis is Exposure (in minutes) and on the Y-axis number of replicates. If `time_points` is not selected, on the X-axis there are peptide ID, and on the Y-axis there are numbers of replicates. If `rep_dat` contains data from one time point of measurement, the histogram colors reflect the number of replicates to highlight the outliers. If `rep_dat` contains multiple time point of measurement, the colors help to distinguish between them.

### Value

a `[ggplot2::ggplot()]` object.

### See Also

[create\\_replicate\\_dataset](#) [show\\_replicate\\_histogram\\_data](#)

## Examples

```
rep_dat <- create_replicate_dataset(alpha_dat)
plot_replicate_histogram(rep_dat)

plot_replicate_histogram(rep_dat, time_points = TRUE)

rep_dat <- create_replicate_dataset(alpha_dat, time_t = 0.167)
plot_replicate_histogram(rep_dat)
```

---

plot\_replicate\_mass\_uptake

*Replicate mass uptake curve*

---

## Description

Plot the mass uptake curve for selected peptide to see the difference between replicates.

## Usage

```
plot_replicate_mass_uptake(
  dat,
  protein = dat[["Protein"]][1],
  state = dat[["State"]][1],
  sequence = dat[["Sequence"]][1],
  aggregated = FALSE,
  log_x = TRUE,
  interactive = getOption("hadex_use_interactive_plots")
)
```

## Arguments

dat	data imported by the <a href="#">read_hdx</a> function
protein	selected protein
state	selected biological state for given protein
sequence	selected peptide sequence for given protein in given biological state
aggregated	logical, indicator if presented data is aggregated on replicate level
log_x	logical, indicator if the X axis values are transformed to log10
interactive	logical, whether plot should have an interactive layer created with <a href="#">ggigraph</a> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app)

### Details

The function `plot_replicate_mass_uptake` generates a plot showing the mass uptake for selected protein in replicates of the experiments. The values can be presented in two ways: as aggregated values for each replicate, or before aggregation - measured values for charge values within a replicate. The mass uptake is generated by subtracting the MHP mass of a peptide from measured mass and the mass uptake is presented in Daltons.

### Value

a `ggplot` object.

### See Also

`read_hdx` `calculate_exp_masses_per_replicate`

### Examples

```
plot_replicate_mass_uptake(alpha_dat)
plot_replicate_mass_uptake(alpha_dat, aggregated = TRUE)
```

---

plot\_state\_comparison *State deuterium uptake comparison*

---

### Description

Comparison plot of deuterium uptake values in time point for biological states.

### Usage

```
plot_state_comparison(
  uptake_dat,
  skip_amino = 0,
  theoretical = FALSE,
  fractional = FALSE,
  line_size = 1.5,
  all_times = FALSE,
  time_t = NULL,
  interactive = getOption("hadex_use_interactive_plots")
)
```

### Arguments

<code>uptake_dat</code>	data produced by <code>calculate_state_uptake</code> function
<code>skip_amino</code>	integer, indicator how many amino acids from the N-terminus should be omitted
<code>theoretical</code>	logical, indicator if values are calculated using theoretical controls

fractional	logical, indicator if values are shown in fractional form
line_size	line size for the plot
all_times	logical, indicator is all all time should be plotted next to each other
time_t	chosen time point
interactive	logical, whether plot should have an interactive layer created with with ggigraph, which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app)

### Details

The function `plot_state_comparison` generates a comparison plot, presenting deuterium uptake values of peptides from selected protein in selected biological states at one time point of measurement at once. On X-axis there is a position in a sequence, with length of a segment of each peptide representing its length. On Y-axis there is deuterium uptake in selected form. Error bars represents the combined and propagated uncertainty.

### Value

a ggplot object

### See Also

[read\\_hdx](#) [calculate\\_state\\_uptake](#)

### Examples

```
uptake_dat <- calculate_state_uptake(alpha_dat)
plot_state_comparison(uptake_dat)

plot_state_comparison(uptake_dat, all_times = TRUE)
plot_state_comparison(uptake_dat, fractional = TRUE, all_times = TRUE)
```

---

plot\_uncertainty

*Uncertainty of the peptide measurements*

---

### Description

Plot the uncertainty of the mass measurements - for aggregated data or before aggregation - to see if there is a region with uncertainty higher than acceptable

## Usage

```
plot_uncertainty(  
  dat,  
  protein = dat[["Protein"]][1],  
  state = dat[["State"]][1],  
  skip_amino = 0,  
  aggregated = TRUE,  
  separate_times = TRUE,  
  show_threshold = TRUE,  
  interactive = getOption("hadex_use_interactive_plots")  
)
```

## Arguments

dat	data imported by the <a href="#">read_hdx</a> function
protein	selected protein
state	selected biological state for given protein
skip_amino	integer, indicator how many amino acids from the N-terminus should be omitted
aggregated	logical, indicator if presented data is aggregated on replicate level
separate_times	logical, indicator if the values for different time points are presented separately
show_threshold	logical, indicator if the threshold of significance is shown
interactive	logical, whether plot should have an interactive layer created with <a href="#">ggigraph</a> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app)

## Details

The function [plot\\_uncertainty](#) generates a plot of uncertainty of mass measurement of each peptide from selected protein in selected biological state. The values can be presented in two ways: as aggregated values for each replicate, or before aggregation - measured values for charge values within a replicate. On X-axis there is a position in a sequence, with length of a segment of each peptide representing its length. On Y-axis there is uncertainty of the measurement in Daltons. The threshold is set to 1 Da, as this value is associated with exchange.

## Value

a [ggplot](#) object.

## See Also

[read\\_hdx](#) [calculate\\_exp\\_masses](#)

**Examples**

```

plot_uncertainty(alpha_dat)
plot_uncertainty(alpha_dat, aggregated = FALSE)
plot_uncertainty(alpha_dat, aggregated = FALSE, separate_times = FALSE)
plot_uncertainty(alpha_dat, skip_amino = 1)

```

---

plot_uptake_curve	<i>Deuterium uptake curve</i>
-------------------	-------------------------------

---

**Description**

Plot deuterium uptake curve for selected peptides

**Usage**

```

plot_uptake_curve(
  uc_dat,
  theoretical = FALSE,
  fractional = FALSE,
  uncertainty_type = "ribbon",
  log_x = TRUE,
  interactive = getOption("hadex_use_interactive_plots")
)

```

**Arguments**

uc_dat	data produced by <a href="#">calculate_kinetics</a> or <a href="#">calculate_peptide_kinetics</a> or <a href="#">create_kinetic_dataset</a> functions
theoretical	logical, indicator if values are calculated using theoretical controls
fractional	logical, indicator if values are shown in fractional form
uncertainty_type	type of uncertainty presentation, possible values: "ribbon", "bars" or "bars + line"
log_x	logical, indicator if the X axis values are transformed to log10
interactive	logical, whether plot should have an interactive layer created with with <a href="#">ggigraph</a> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

**Details**

The function [plot\\_uptake\\_curve](#) generates the deuterium uptake curve for selected peptides from selected protein. On X-axis there are time points of measurements. On Y-axis there is deuterium uptake in selected form. The combined and propagated uncertainty can be presented as ribbons or error bars.

**Value**

a `ggplot` object.

**See Also**

`read_hdx` `calculate_kinetics` `calculate_peptide_kinetics` `create_kinetic_dataset`

**Examples**

```
uc_dat <- calculate_kinetics(alpha_dat, protein = "db_eEF1Ba",
                           sequence = "GFGDLKSPAGL",
                           state = "Alpha_KSCN",
                           start = 1, end = 11,
                           time_0 = 0, time_100 = 1440)
plot_uptake_curve(uc_dat = uc_dat,
                  theoretical = FALSE,
                  fractional = TRUE)
```

---

plot\_volcano

*Volcano plot*

---

**Description**

Volcano plot for differential deuterium uptake between two biological states

**Usage**

```
plot_volcano(
  p_dat,
  state_1 = "",
  state_2 = "",
  adjust_axes = TRUE,
  show_confidence_limits = FALSE,
  confidence_level = 0.98,
  color_times = TRUE,
  show_insignificant_grey = FALSE,
  hide_insignificant = FALSE,
  fractional = FALSE,
  theoretical = FALSE,
  interactive = getOption("hadex_use_interactive_plots")
)
```

## Arguments

p_dat	data produced by the <a href="#">create_p_diff_uptake_dataset</a> function
state_1	selected biological state for given protein
state_2	selected biological state for given protein
adjust_axes	<a href="#">logical</a> , indicator if the X-axis is symmetrical in relation to 0
show_confidence_limits	<a href="#">logical</a> , indicates if the hybrid test confidence levels are shown
confidence_level	confidence level for the test, from range [0, 1]. It should be the same as used to prepare p_dat
color_times	<a href="#">logical</a> , indicator if different time points are distinguishable by color
show_insignificant_grey	<a href="#">logical</a> , indicator if the values not passing the test are shown in grey
hide_insignificant	<a href="#">logical</a> , indicator if the values not passing the test are hidden
fractional	<a href="#">logical</a> , indicator if values are shown in fractional form
theoretical	<a href="#">logical</a> , indicator if values are calculated using theoretical controls
interactive	<a href="#">logical</a> , whether plot should have an interactive layer created with with <a href="#">ggigraph</a> , which would add tooltips to the plot in an interactive display (HTML/Markdown documents or shiny app).

## Details

The function [plot\\_volcano](#) generates the volcano plot based on supplied p\_dat. On X-axis there is differential deuterium uptake in selected form. On Y-axis there is the P-value from t-Student test between two biological states. Based on selected confidence level, the confidence limits are calculated to indicate statistically significant values - shown as red dotted lines. The values that are in upper left and right corners pass the hybrid test.

## Value

a [ggplot](#) object.

## References

Hageman, T. S. & Weis, D. D. Reliable Identification of Significant Differences in Differential Hydrogen Exchange-Mass Spectrometry Measurements Using a Hybrid Significance Testing Approach. *Anal Chem* 91, 8008–8016 (2019).

Houde, D., Berkowitz, S.A., and Engen, J.R. (2011). The Utility of Hydrogen/Deuterium Exchange Mass Spectrometry in Biopharmaceutical Comparability Studies. *J Pharm Sci* 100, 2071–2086.

## See Also

[create\\_p\\_diff\\_uptake\\_dataset](#)

## Examples

```
p_dat <- create_p_diff_uptake_dataset(alpha_dat)
plot_volcano(p_dat, show_confidence_limits = TRUE)

plot_volcano(p_dat, show_confidence_limits = TRUE, show_insignificant_grey = TRUE)
plot_volcano(p_dat, show_confidence_limits = TRUE, hide_insignificant = TRUE)
```

---

```
prepare_hdxviewer_export
```

*Prepares data export for HDX-Viewer*

---

## Description

This function produces the data in format suitable for HDX-Viewer. If necessary, this result can be downloaded as the csv file with download indicator set to true.

## Usage

```
prepare_hdxviewer_export(  
  x_dat,  
  differential = FALSE,  
  fractional = TRUE,  
  theoretical = FALSE,  
  download = FALSE,  
  file_path = tempdir()  
)
```

## Arguments

x_dat	one state deuterium uptake data or differential uptake data
differential	indicator of x_dat type
fractional	indicator if fractional values are used
theoretical	indicator if theoretical values are used
download	indicator if the result should be downloaded as csv file
file_path	path for saving downloaded file

## Value

a `data.frame` object

## Examples

```
# disabled due to long execution time

kin_dat <- create_uptake_dataset(alpha_dat, states = "Alpha_KSCN" )
aggregated_dat <- create_aggregated_uptake_dataset(kin_dat)
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
diff_aggregated_dat <- create_aggregated_diff_uptake_dataset(diff_uptake_dat)
prepare_hdxviewer_export(aggregated_dat, differential = FALSE)
# prepare_hdxviewer_export(aggregated_dat, differential = TRUE) # shouldnt work
prepare_hdxviewer_export(diff_aggregated_dat, differential = TRUE)
# prepare_hdxviewer_export(aggregated_dat, differential = FALSE, download = TRUE) # writes a file
```

---

read\_hdx

*Read HDX-MS data file*

---

## Description

Import HDX-MS datafile and validate its content

## Usage

```
read_hdx(filename, separator = ",")
```

## Arguments

filename            a file supplied by the user. Formats allowed: .csv, .xlsx and .xls.  
separator           a value separating the columns.

## Details

The function `read_hdx` generates a dataset read from the supplied datafile. The files produced by DynamX 3.0 or 2.0 in 'cluster data' format and 'tables' file from HDeXaminer are handled. Moreover, the data should include at least two replicates of the experiment to calculate the uncertainty of the measurement. For the files of HDeXaminer origin, the rows with no complete information (e.g. missing 'Exp Cent' value) are removed. The 'Confidence' column is preserved as the user should have impact on accepting rows based on their Confidence flag. Moreover, those files need action from the user - to confirm data processing (e.g. FD time point), choose accepted confidence values and make some change of the labels using `update_hdexaminer_file` function. For further information check the documentation. IMPORTANT! The files of HDeXaminer origin MUST be processed by hand or by `update_hdexaminer_file` function to fit the input of processing functions e.g. `calculate_state_uptake` or `calculate_diff_uptake`.

## Value

a `data.frame` object

**See Also**

[update\\_hdexaminer\\_file](#) [create\\_control\\_dataset](#) [calculate\\_state\\_uptake](#)

**Examples**

```
dat <- read_hdx(system.file(package = "HaDeX2",
                           "HaDeX/data/alpha.csv"))
head(dat)
```

---

reconstruct\_sequence    *Reconstruct protein sequence*

---

**Description**

Reconstruct protein sequence from experimental data

**Usage**

```
reconstruct_sequence(  
  dat,  
  protein = dat[["Protein"]][1],  
  states = unique(dat[["State"]]),  
  end = NULL  
)
```

**Arguments**

dat	data read by <a href="#">read_hdx</a>
protein	selected protein
states	selected biological states for given protein
end	<a href="#">numeric</a> , end position of the protein

**Details**

The function [reconstruct\\_sequence](#) generates protein sequence from supplied experimental data. For a position not covered, letter x is shown. If the C-terminus of protein is not covered, there is a possibility to manually fix the protein length by passing a value to the ‘end’ parameter.

**Value**

a [character](#) object.

**See Also**

[read\\_hdx](#)

## Examples

```
reconstruct_sequence(alpha_dat)
```

---

```
show_aggregated_uptake_data
```

*Show aggregated values in friendly form*

---

## Description

Function plots the aggregated uptake data with regard to submitted parameters in a friendly form. Designed for GUI.

## Usage

```
show_aggregated_uptake_data(  
  aggregated_dat,  
  differential = FALSE,  
  fractional = TRUE,  
  theoretical = FALSE  
)
```

## Arguments

aggregated_dat	aggregated uptake data
differential	indicator if the aggregated_dat is single-state or differential
fractional	logical, determines if values are fractional
theoretical	logical, determines if values are theoretical

## Value

a `data.frame` object

## Examples

```
# disabled due to long execution time  
  
kin_dat <- create_uptake_dataset(alpha_dat, states = "Alpha_KSCN")  
aggregated_dat <- create_aggregated_uptake_dataset(kin_dat)  
show_aggregated_uptake_data(aggregated_dat)  
  
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)  
aggregated_diff_dat <- create_aggregated_diff_uptake_dataset(diff_uptake_dat)  
show_aggregated_uptake_data(aggregated_diff_dat, differential = TRUE)
```

---

```
show_coverage_heatmap_data
```

*Coverage heatmap data*

---

**Description**

This function prepares the data used in coverage heatmap to be shown in user-friendly way.

**Usage**

```
show_coverage_heatmap_data(x_dat, value = NULL)
```

**Arguments**

x_dat	data created using calculate_ or create_ function
value	value to be presented

**Value**

a `data.table` object

**Examples**

```
# auc data
auc_dat <- calculate_auc(create_uptake_dataset(alpha_dat))
show_coverage_heatmap_data(auc_dat, value = "auc")

# back-exchange
bex_dat <- calculate_back_exchange(alpha_dat)
show_coverage_heatmap_data(bex_dat, value = "back_exchange")
```

---

```
show_diff_uptake_data
```

*Differential deuterium uptake data*

---

**Description**

Present differential deuterium uptake values in selected form

**Usage**

```
show_diff_uptake_data(
  diff_uptake_dat,
  theoretical = FALSE,
  fractional = FALSE,
  renamed = TRUE
)
```

**Arguments**

diff_uptake_dat	data produced by <a href="#">create_diff_uptake_dataset</a> function
theoretical	logical, indicator if values are calculated using theoretical controls
fractional	logical, indicator if values are shown in fractional form
renamed	logical, indicator if the names of the columns are renamed to user-friendly ones. Currently FALSE not implemented.

**Details**

The function [show\\_uptake\\_data](#) generates a subsets of the diff\_uptake\_dat based on selected parameters. The numerical values are rounded to 4 places. The names of columns are changed to user-friendly ones.

**Value**

a `data.frame` object

**See Also**

[read\\_hdx](#) [create\\_diff\\_uptake\\_dataset](#)

**Examples**

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
head(show_diff_uptake_data(diff_uptake_dat))
```

---

show\_diff\_uptake\_data\_confidence

*Differential uptake data with confidence*

---

**Description**

Present differential deuterium uptake values in selected form, accompanied by the significance

**Usage**

```
show_diff_uptake_data_confidence(
  diff_uptake_dat,
  theoretical = FALSE,
  fractional = FALSE,
  confidence_level = 0.98,
  hybrid = FALSE
)
```

### Arguments

diff_uptake_dat	data produced by <a href="#">create_diff_uptake_dataset</a> function
theoretical	logical, indicator if values are calculated using theoretical controls
fractional	logical, indicator if values are shown in fractional form
confidence_level	confidence level for the test, from range [0, 1].
hybrid	logical, indicator if the hybrid testing was applied in diff_uptake_dat.

### Details

The function [show\\_uptake\\_data](#) generates a subsets of the uptake dat based on selected parameters. It contains the information if the value is statistically significant at selected confidence level. The numerical values are rounded to 4 places. The names of columns are changed to user-friendly ones.

### Value

a `data.frame` object

### See Also

[create\\_diff\\_uptake\\_dataset](#) [plot\\_differential](#)

### Examples

```
diff_uptake_dat <- create_diff_uptake_dataset(alpha_dat)
show_diff_uptake_data_confidence(diff_uptake_dat)
```

---

show\_overlap\_data      *Show data on peptide overlap*

---

### Description

Presents peptide overlap on protein sequence data, based on the supplied parameters.

### Usage

```
show_overlap_data(
  dat,
  protein = dat[["Protein"]][1],
  state = dat[["State"]][1],
  start = min(dat[["Start"]]),
  end = max(dat[["End"]])
)
```

**Arguments**

<code>dat</code>	data imported by the <a href="#">read_hdx</a> function.
<code>protein</code>	chosen protein.
<code>state</code>	biological state for chosen protein.
<code>start</code>	start position of chosen protein.
<code>end</code>	end position of chosen protein.

**Details**

The data frame presents all the peptides in given state, with its start and end position on the protein sequence. This data is available in the GUI.

**Value**

a `data.frame` object.

**See Also**

[read\\_hdx\\_plot\\_overlap](#)

**Examples**

```
show_overlap_data(alpha_dat)
```

---

```
show_peptide_charge_measurement
```

*Show peptide charge measurement*

---

**Description**

Show the charge measurements from replicates for peptide in specific time point.

**Usage**

```
show_peptide_charge_measurement(  
  dat,  
  protein = dat[["Protein"]][1],  
  state = dat[["State"]][1],  
  sequence = dat[["Sequence"]][1],  
  time_t = unique(dat[["Exposure"]])[3]  
)
```

**Arguments**

dat	data as imported by the <a href="#">read_hdx</a> function.
protein	chosen protein.
state	biological state for chosen protein.
sequence	sequence of chosen peptide.
time_t	time point of the measurement.

**Details**

This function shows the measurements of charge from different replicates for specific peptide in specific state in specific time point of measurement.

**Value**

a `data.frame` object.

**See Also**

[read\\_hdx](#) [plot\\_peptide\\_charge\\_measurement](#)

**Examples**

```
show_peptide_charge_measurement(alpha_dat)
```

---

```
show_peptide_mass_measurement
```

*Show peptide mass measurement*

---

**Description**

Show the mass measurements from replicates for peptide in specific time point.

**Usage**

```
show_peptide_mass_measurement(  
  rep_mass_dat,  
  protein = rep_mass_dat[["Protein"]][1],  
  state = rep_mass_dat[["State"]][1],  
  sequence = rep_mass_dat[["Sequence"]][1],  
  time_t = unique(rep_mass_dat[["Exposure"]])[3]  
)
```

**Arguments**

rep_mass_dat	data produced by <a href="#">calculate_exp_masses_per_replicate</a> function.
protein	chosen protein.
state	biological state for chosen protein.
sequence	sequence of chosen peptide.
time_t	time point of the measurement.

**Details**

This function shows the measurements of mass from different replicates for specific peptide in specific state in specific time point of measurement.

**Value**

a `data.frame` object.

**See Also**

[read\\_hdx](#) [calculate\\_exp\\_masses\\_per\\_replicate](#) [calculate\\_exp\\_masses](#) [calculate\\_state\\_uptake](#)  
[calculate\\_diff\\_uptake](#)

**Examples**

```
rep_mass_dat <- calculate_exp_masses_per_replicate(alpha_dat)
show_peptide_mass_measurement(rep_mass_dat)
```

---

show\_p\_diff\_uptake\_data

*Differential deuterium uptake data*

---

**Description**

Present differential deuterium uptake values in selected form

**Usage**

```
show_p_diff_uptake_data(  
  p_diff_uptake_dat,  
  theoretical = FALSE,  
  fractional = FALSE,  
  renamed = TRUE  
)
```

**Arguments**

p_diff_uptake_dat	data produced by <a href="#">create_p_diff_uptake_dataset</a> function
theoretical	logical, indicator if values are calculated using theoretical controls
fractional	logical, indicator if values are shown in fractional form
renamed	logical, indicator if the names of the columns are renamed to user-friendly ones. Currently FALSE not implemented.

**Details**

The function [show\\_uptake\\_data](#) generates a subsets of the diff\_uptake\_dat based on selected parameters. The numerical values are rounded to 4 places. The names of columns are changed to user-friendly ones.

**Value**

a `data.frame` object

**See Also**

[read\\_hdx](#) [create\\_diff\\_uptake\\_dataset](#)

**Examples**

```
p_diff_uptake_dat <- create_p_diff_uptake_dataset(alpha_dat)
head(show_p_diff_uptake_data(p_diff_uptake_dat))
```

---

```
show_quality_control_data
```

*Show quality control data*

---

**Description**

Generates quality control data, based on the supplied parameters.

**Usage**

```
show_quality_control_data(qc_dat)
```

**Arguments**

qc_dat	data produced by <a href="#">create_quality_control_dataset</a> function, scaled if necessary.
--------	--

**Details**

This data frame presents the mean uncertainty in function of selected maximal exchange control time of measurement. This data is available in the GUI.

**Value**

a `data.frame` object.

**See Also**

[create\\_quality\\_control\\_dataset](#) [plot\\_quality\\_control](#)

**Examples**

```
qc_dat <- create_quality_control_dataset(alpha_dat)
show_quality_control_data(qc_dat)
```

---

```
show_replicate_histogram_data
```

*Show replicate data*

---

**Description**

Show histogram data on number of replicates per peptide in one or multiple time point of measurement.

**Usage**

```
show_replicate_histogram_data(rep_dat)
```

**Arguments**

`rep_dat` replicate data, created by [create\\_replicate\\_dataset](#) function.

**Details**

The function shows the information about number of replicates for peptides in one or multiple time point of measurement, depends on supplied data.

**Value**

a `data.frame` object.

**See Also**

[create\\_replicate\\_dataset](#) [plot\\_replicate\\_histogram](#)

## Examples

```
rep_dat <- create_replicate_dataset(alpha_dat)
show_replicate_histogram_data(rep_dat)
```

---

show_summary_data	<i>Summary data</i>
-------------------	---------------------

---

## Description

Show summary table

## Usage

```
show_summary_data(
  dat,
  confidence_level = "",
  protein_length = max(dat[["End"]])
)
```

## Arguments

`dat` data imported by the [read\\_hdx](#) function

`confidence_level` confidence level for the test, from range [0, 1].

`protein_length` the length of the protein sequence

## Details

The format in the table is suggested by the community and should be provided for experimental data.

## Value

a `data.table` object

## References

Masson, G.R., Burke, J.E., Ahn, N.G., Anand, G.S., Borchers, C., Brier, S., Bou-Assaf, G.M., Engen, J.R., Englander, S.W., Faber, J., et al. (2019). Recommendations for performing, interpreting and reporting hydrogen deuterium exchange mass spectrometry (HDX-MS) experiments. *Nat Methods* 16, 595–602

## See Also

[read\\_hdx](#)

**Examples**

```
show_summary_data(alpha_dat)
```

---

show_uc_data	<i>Deuterium uptake curve data</i>
--------------	------------------------------------

---

**Description**

Present deuterium uptake curve data

**Usage**

```
show_uc_data(uc_dat, theoretical = FALSE, fractional = FALSE)
```

**Arguments**

uc_dat	calculated kinetic data by <a href="#">calculate_kinetics</a> or <a href="#">calculate_peptide_kinetics</a> or <a href="#">create_kinetic_dataset</a> function
theoretical	logical, indicator if values are calculated using theoretical controls
fractional	logical, indicator if values are shown in fractional form

**Details**

The function [show\\_uptake\\_data](#) generates a subsets of the uc\_dat based on selected parameters. The numerical values are rounded to 4 places. The names of columns are changed to user-friendly ones.

**Value**

a [data.frame](#) object

**See Also**

[read\\_hdx](#) [calculate\\_kinetics](#) [calculate\\_peptide\\_kinetics](#)

**Examples**

```
uc_dat <- calculate_kinetics(alpha_dat,
                             sequence = "GFGDLKSPAGL",
                             state = "ALPHA_Gamma",
                             start = 1, end = 11)
show_uc_data(uc_dat)
```

---

show_uptake_data	<i>Deuterium uptake data</i>
------------------	------------------------------

---

### Description

Present deuterium uptake values in selected form

### Usage

```
show_uptake_data(  
  uptake_dat,  
  theoretical = FALSE,  
  fractional = FALSE,  
  renamed = TRUE  
)
```

### Arguments

uptake_dat	data produced by <a href="#">create_uptake_dataset</a> function or <a href="#">create_state_uptake_dataset</a>
theoretical	logical, indicator if values are calculated using theoretical controls
fractional	logical, indicator if values are shown in fractional form
renamed	logical, indicator if the names of the columns are renamed to user-friendly ones. Currently FALSE not implemented.

### Details

The function [show\\_uptake\\_data](#) generates a subsets of the uptake\_dat based on selected parameters. The numerical values are rounded to 4 places. The names of columns are changed to user-friendly ones.

### Value

a [data.frame](#) object

### See Also

[read\\_hdx](#) [create\\_uptake\\_dataset](#)

### Examples

```
uptake_dat <- create_uptake_dataset(alpha_dat)  
show_uptake_data(uptake_dat)
```

---

`update_hdexaminer_file`*Update HDeXaminer datafile*

---

## Description

Update data from HDeXaminer file

## Usage

```
update_hdexaminer_file(  
  dat,  
  fd_time,  
  old_protein_name = NULL,  
  new_protein_name = NULL,  
  old_state_name = NULL,  
  new_state_name = NULL,  
  confidence = c("High", "Medium")  
)
```

## Arguments

<code>dat</code>	data imported by the <a href="#">read_hdx</a> function
<code>fd_time</code>	time point [min] for fully deuterated sample
<code>old_protein_name</code>	protein name to be changed
<code>new_protein_name</code>	new name for <code>old_protein_name</code>
<code>old_state_name</code>	state names to be changed
<code>new_state_name</code>	new names for <code>old_state_name</code>
<code>confidence</code>	vector of accepted confidence values (internal flag from HDeXaminer). By default only accepted values are 'Medium' and 'High', with 'Low' excluded

## Details

The function [update\\_hdexaminer\\_file](#) changes the data read from HDeXaminer file. Data from HDeXaminer is condensed and automated data retrieving may be corrected by the user. The original file has a mark "FD" for fully deuterated data instead of numerical value for time point (provided in minutes) that is not consistent for workflow and not enough for precise data description. Moreover, the data about both protein and state is included in one column and for detailed information function [update\\_hdexaminer\\_file](#) allows to change them.

## Value

a `data.frame` object

**See Also**

[read\\_hdx](#) [calculate\\_kinetics](#) [plot\\_coverage](#) [plot\\_position\\_frequency](#) [reconstruct\\_sequence](#)

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