

# Package ‘LFApp’

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**Title** Shiny Apps for Lateral Flow Assays

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**Description** Shiny apps for the quantitative analysis of images from lateral flow assays (LFAs). The images are segmented and background corrected and color intensities are extracted. The apps can be used to import and export intensity data and to calibrate LFAs by means of linear, loess, or gam models. The calibration models can further be saved and applied to intensity data from new images for determining concentrations.

**License** LGPL-3

**Depends** R (>= 4.0.0)

**Imports** stats, utils, graphics, methods, mgecv, shiny, shinyjs,  
shinythemes, shinyFiles, shinyMobile (>= 0.9), EBImage, DT,  
ggplot2, fs

**Suggests** knitr, rmarkdown, remotes

**VignetteBuilder** knitr

**Encoding** UTF-8

**URL** <https://github.com/fpaskali/LFApp>

**BugReports** <https://github.com/fpaskali/LFApp/issues>

**NeedsCompilation** no

**Repository** CRAN

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LFApp-package	<i>Shiny Apps for Lateral Flow Assays.</i>
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### Description

Shiny apps for the quantitative analysis of images from lateral flow assays (LFAs). The images are segmented and background corrected and color intensities are extracted. The apps can be used to import and export intensity data and to calibrate LFAs by means of linear, loess, or gam models. The calibration models can further be saved and applied to intensity data from new images for determining concentrations.

### Details

library(LFApp)

### Author(s)

Filip Paskali, Weronika Schary, Matthias Kohl  
 Maintainer: Filip Paskali <Filip.Paskali@gmail.com>

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run_functions	<i>Run Analysis Shiny Apps</i>
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### Description

Function start the Analysis Shiny App.

### Usage

```
run_analysis()
run_cal()
run_core()
run_quan()
run_mobile_analysis()
run_mobile_cal()
run_mobile_core()
run_mobile_quan()
```

**Details**

The functions start the various shiny apps included in the package.

**Value**

An object that represents the app. Printing the object will run the app.

**Author(s)**

Filip Paskali <F.Paskali@hs-furtwangen.de>, Weronika Schary <W.Schary@hs-furtwangen.de>, Matthias Kohl <Matthias.Kohl@stamats.de>

**Examples**

```
if(interactive()){  
  ## start full analysis app  
  run_analysis()  
  ## start mobile version of full analysis app  
  run_mobile_analysis()  
}
```

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threshold\_li

*Li Thresholding Algorithm*

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**Description**

The function computes a background threshold of an image by using Li's iterative minimum cross entropy method.

**Usage**

```
threshold_li(image, tolerance = NULL, initial_guess = NULL, iter_callback = NULL)
```

**Arguments**

image            object of class Image from package EBImage.  
tolerance        optional tolerance threshold.  
initial\_guess    optional initial value for the minimization.  
iter\_callback    optional function applied to the minimization criterion.

**Details**

For more details about the method see Li and Lee (1993) as well as Li and Tam (1998).

**Value**

numeric vector with the computed threshold.

**Author(s)**

Filip Paskali <Filip.Paskali@gmail.de>

**References**

C.H. Li and C.K. Lee (1993). Minimum cross entropy thresholding. *Pattern Recognition* **26** (4): 617-25. [https://doi.org/10.1016/0031-3203\(93\)90115-D](https://doi.org/10.1016/0031-3203(93)90115-D).

C.H. Li and P.K.S. Tam (1998). An iterative algorithm for minimum cross entropy thresholding. *Pattern Recognition Letters* **19** (8): 771-76. [https://doi.org/10.1016/S0167-8655\(98\)00057-9](https://doi.org/10.1016/S0167-8655(98)00057-9).

**Examples**

```
library(EBImage)
x <- readImage(system.file("images", "sample.TIF", package="LFApp"))
threshold_li(x)
```

---

triangle

*Triangle Thresholding Algorithm*

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**Description**

The function computes a background threshold of an image using the triangle algorithm.

**Usage**

```
triangle(image, offset = 0.2, breaks = 256)
```

**Arguments**

image	object of class Image from package EBImage.
offset	numeric, additional offset added to the computed threshold.
breaks	integer, number of breaks used in the histogram.

**Details**

The Triangle method is based on the histogram of the intensities. Based on the range of intensities and the maximum peak a threshold is determined. The method was proposed in Zack et al. (1977).

**Value**

numeric vector with the computed threshold.

**Author(s)**

Matthias Kohl <Matthias.Kohl@stamats.de>

## References

G.W. Zack, W. E. Rogers, and S. A. Latt (1977). Automatic measurement of sister chromatid exchange frequency. *The journal of histochemistry and cytochemistry: official journal of the Histochemistry Society* **25** (7): 741-53. <https://doi.org/10.1177/25.7.70454>.

## Examples

```
library(EImage)
x <- readImage(system.file("images", "sample.TIF", package="LFApp"))
triangle(x)
```

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