

Package ‘ampir’

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Type Package

Title Predict Antimicrobial Peptides

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Description

A toolkit to predict antimicrobial peptides from protein sequences on a genome-wide scale. It incorporates two support vector machine models (“precursor” and “mature”) trained on publicly available antimicrobial peptide data using calculated physico-chemical and compositional sequence properties described in Meher et al. (2017) <[doi:10.1038/srep42362](https://doi.org/10.1038/srep42362)>.

In order to support genome-wide analyses, these models are designed to accept any type of protein as input and calculation of compositional properties has been optimised for high-throughput use. For best results it is important to select the model that accurately represents your sequence type: for full length proteins, it is recommended to use the default “precursor” model. The alternative, “mature”, model is best suited for mature peptide sequences that represent the final antimicrobial peptide sequence after post-translational processing. For details see Fingerhut et al. (2020) <[doi:10.1093/bioinformatics/btaa653](https://doi.org/10.1093/bioinformatics/btaa653)>.

The ‘ampir’ package is also available via a Shiny based GUI at <<https://ampir.marine-omics.net/>>.

URL <https://github.com/Legana/ampir>

License GPL-2

Encoding UTF-8

Depends R (>= 3.5.0)

Imports Peptides, caret (>= 6.0.0), kernlab, Rcpp, parallel

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aaseq_is_valid	<i>Check protein sequences for non-standard amino acids</i>
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Description

Any proteins that contains an amino acid that is not one of the 20 standard amino acids is flagged as invalid

Usage

```
aaseq_is_valid(seq)
```

Arguments

seq	A vector of protein sequences
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Value

A logical vector where TRUE indicates a valid protein sequence and FALSE indicates a sequence with invalid amino acids

calculate_features *Calculate a set of numerical features from protein sequences*

Description

This function calculates set physicochemical and compositional features from protein sequences in preparation for supervised model learning

Usage

```
calculate_features(df, min_len = 10)
```

Arguments

df	A dataframe which contains protein sequence names as the first column and amino acid sequence as the second column
min_len	Minimum length sequence for which features can be calculated. It is an error to provide sequences with length shorter than this

Value

A dataframe containing numerical values related to the protein features of each given protein

Note

This function depends on the Peptides package

References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. The R Journal. 7(1), 4–14 (2015).

Examples

```
my_protein_df <- read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))

calculate_features(my_protein_df)
## Output (showing the first six output columns)
#   seq_name      Amphiphilicity Hydrophobicity      pI      Mw      Charge      ....
# [1] G1P6H5_MYOLU      0.4145847      0.4373494      8.501312      9013.757      4.53015      ....
```

calc_amphiphilicity *Calculate amphiphilicity (or hydrophobic moment)*

Description

Calculate amphiphilicity (or hydrophobic moment)

Usage

```
calc_amphiphilicity(seq)
```

Arguments

seq A protein sequence

References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal*. 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

calc_hydrophobicity *Calculate the hydrophobicity*

Description

Calculate the hydrophobicity

Usage

```
calc_hydrophobicity(seq)
```

Arguments

seq A protein sequence

References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal*. 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

calc_mw	<i>Calculate the molecular weight</i>
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Description

Calculate the molecular weight

Usage

```
calc_mw(seq)
```

Arguments

seq	A protein sequence
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References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal*. 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

calc_net_charge	<i>Calculate the net charge</i>
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Description

Calculate the net charge

Usage

```
calc_net_charge(seq)
```

Arguments

seq	A protein sequence
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References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal*. 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

calc_pI *Calculate the isoelectric point (pI)*

Description

Calculate the isoelectric point (pI)

Usage

```
calc_pI(seq)
```

Arguments

seq pI

References

Osorio, D., Rondon-Villarreal, P. & Torres, R. Peptides: A package for data mining of antimicrobial peptides. *The R Journal*. 7(1), 4–14 (2015). The imported function originates from the Peptides package (<https://github.com/dosorio/Peptides/>).

calc_pseudo_comp *Calculate the pseudo amino acid composition*

Description

This function is adapted from the extractPAAC function from the protr package (<https://github.com/nanxstats/protr>)

Usage

```
calc_pseudo_comp(seq, lambda_min = 4, lambda_max = 19)
```

Arguments

seq A vector of protein sequences as character strings

lambda_min Minimum allowable lambda. It is an error to provide a protein sequence shorter than lambda_min+1

lambda_max For each sequence lambda will be set to one less than the sequence length or lambda_max, whichever is smaller

References

Nan Xiao, Dong-Sheng Cao, Min-Feng Zhu, and Qing-Song Xu. (2015). protr/ProtrWeb: R package and web server for generating various numerical representation schemes of protein sequences. *Bioinformatics* 31 (11), 1857-1859.

chunk_rows	<i>Determine row breakpoints for dividing a dataset into chunks for parallel processing</i>
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Description

Determine row breakpoints for dividing a dataset into chunks for parallel processing

Usage

```
chunk_rows(nrows, n_cores)
```

Arguments

nrows	The number of rows in the dataset to be chunked
n_cores	The number of cores that will be used for parallel processing

Value

A list of integer vectors consisting of the rows in each chunk

df_to_faa	<i>Save a dataframe in FASTA format</i>
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Description

This function writes a dataframe out as a FASTA format file

Usage

```
df_to_faa(df, file = "")
```

Arguments

df	a dataframe containing two columns: the sequence name and amino acid sequence itself
file	file path to save the named file to

Value

A FASTA file where protein sequences are represented in two lines: The protein name preceded by a greater than symbol, and a new second line that contains the protein sequence

Examples

```
my_protein <- read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))

# Write a dataframe to a FASTA file
df_to_faa(my_protein, tempfile("my_protein.fasta", tempdir()))
```

predict_amps	<i>Predict the antimicrobial peptide probability of a protein</i>
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Description

This function predicts the probability of a protein to be an antimicrobial peptide

Usage

```
predict_amps(faa_df, min_len = 5, n_cores = 1, model = "precursor")
```

Arguments

faa_df	A dataframe obtained from read_faa containing two columns: the sequence name (seq_name) and amino acid sequence (seq_aa)
min_len	The minimum protein length for which predictions will be generated
n_cores	On multicore machines split the task across this many processors. This option does not work on Windows
model	Either a string with the name of a built-in model (mature, precursor), OR, A train object suitable for passing to the predict.train function in the caret package. If omitted the default model will be used.

Value

The original input data.frame with a new column added called prob_AMP with the probability of that sequence to be an antimicrobial peptide. Any sequences that are too short or which contain invalid amino acids will have NA in this column

Examples

```
my_bat_faa_df <- read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))

predict_amps(my_bat_faa_df)
#   seq_name   prob_AMP
# [1] G1P6H5_MYOLU  0.9723796
```

read_faa	<i>Read FASTA amino acids file into a dataframe</i>
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Description

This function reads a FASTA amino acids file into a dataframe

Usage

```
read_faa(file = NULL)
```

Arguments

file file path to the FASTA format file containing the protein sequences

Value

Dataframe containing the sequence name (seq_name) and sequence (seq_aa) columns

Note

This function was adapted from ‘read.fasta.R’ by Jinlong Zhang (jinlongzhang01@gmail.com) for the phylotools package (<http://github.com/helixcn/phylotools>)

Examples

```
read_faa(system.file("extdata/bat_protein.fasta", package = "ampir"))
```

```
## Output
#            seq_name                    seq_aa
# [1] G1P6H5_MYOLU  MALTVRIQAACLLLLLLASLTSYSL....
```

remove_nonstandard_aa	<i>Remove non standard amino acids from protein sequences</i>
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Description

This function removes anything that is not one of the 20 standard amino acids in protein sequences

Usage

```
remove_nonstandard_aa(df)
```

Arguments

df A dataframe which contains protein sequence names as the first column and amino acid sequence as the second column

Value

a dataframe like the input dataframe but with removed proteins that contained non standard amino acids

Examples

```
non_standard_df <- readRDS(system.file("extdata/non_standard_df.rds", package = "ampir"))

# non_standard_df
#   seq_name          seq_aa
# [1] G1P6H5_MYOLU  MALTVRIQAACLLLLLLASLSYLLLSQTTQLADLQTQ...
# [2] fake_sequence  MKVTHEUSYR$GXMBIJIDG*M80-%

remove_nonstandard_aa(non_standard_df)
#   seq_name          seq_aa
# [1] G1P6H5_MYOLU  MALTVRIQAACLLLLLLASLSYLLLSQTTQLADLQTQ...
```

remove_stop_codon	<i>Remove stop codon at end of sequence</i>
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Description

Stop codons at the end of the amino acid sequences are removed

Usage

```
remove_stop_codon(faa_df)
```

Arguments

faa_df A dataframe containing two columns: the sequence name and amino acid sequence

Value

The input dataframe without the stop codons at the end of sequences

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