

Package ‘fitdistrplus’

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Title Help to Fit of a Parametric Distribution to Non-Censored or Censored Data

Version 1.2-6

Description Extends the `fitdistr()` function (of the MASS package) with several functions to help the fit of a parametric distribution to non-censored or censored data. Censored data may contain left censored, right censored and interval censored values, with several lower and upper bounds. In addition to maximum likelihood estimation (MLE), the package provides moment matching (MME), quantile matching (QME), maximum goodness-of-fit estimation (MGE) and maximum spacing estimation (MSE) methods (available only for non-censored data). Weighted versions of MLE, MME, QME and MSE are available. See e.g. Casella & Berger (2002), Statistical inference, Pacific Grove, for a general introduction to parametric estimation.

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<https://lbbe.univ-lyon1.fr/fr/fitdistrplus>,
<https://github.com/lbbe-software/fitdistrplus>

BugReports <https://github.com/lbbe-software/fitdistrplus/issues>

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Description

The idea of this package emerged in 2008 from a collaboration between J.B. Denis, R. Pouillot and M.L. Delignette who at this time worked in the area of quantitative risk assessment. The implementation of this package was a part of a more general project named "Risk assessment with R" gathering different packages and hosted in [R-forge](#).

The **fitdistrplus** package was first written by M.L. Delignette-Muller and made available in [CRAN](#) on 2009 and presented at the [2009 useR conference](#) in Rennes. A few months after, C. Dutang joined the project by starting to participate to the implementation of the **fitdistrplus** package. The package has also been presented at the [2011 useR conference](#) at the 2eme rencontres R in 2013 (<https://r2013-lyon.sciencesconf.org/>), and the [2019 useR conference](#). Since 2020, A. Siberchicot helps the development of **fitdistrplus** and maintains the package.

Four vignettes are available within the package:

- a [general overview](#) of the package published in the Journal of Statistical Software ([doi:10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04)),
- a document answering the most [Frequently Asked Questions](#),
- a document presenting a [benchmark of optimization algorithms](#) when finding parameters,
- a document about [starting values](#).

The **fitdistrplus** package is a general package that aims at helping the fit of univariate parametric distributions to censored or non-censored data. The two main functions are `fitdist` for fit on non-censored data and `fitdistcens` for fit on censored data.

The choice of candidate distributions to fit may be helped using functions `descdist` and `plotdist` for non-censored data and `plotdistcens` for censored data).

Using functions `fitdist` and `fitdistcens`, different methods can be used to estimate the distribution parameters:

- maximum likelihood estimation by default (`mledist`),
- moment matching estimation (`mmedist`),
- quantile matching estimation (`qmedist`),
- maximum goodness-of-fit estimation (`mgedist`).

For classical distributions initial values are automatically calculated if not provided by the user. Graphical functions `plotdist` and `plotdistcens` can be used to help a manual calibration of initial values for parameters of non-classical distributions. Function `prefit` is proposed to help the definition of good starting values in the special case of constrained parameters. In the case where maximum likelihood is chosen as the estimation method, function `llplot` enables to visualize log-likelihood contours.

The goodness-of-fit of fitted distributions (a single fit or multiple fits) can be explored using different graphical functions (`cdfcomp`, `denscomp`, `qqcomp` and `ppcomp` for non-censored data and

`cdfcompdens` for censored data). Goodness-of-fit statistics are also provided for non-censored data using function `gofstat`.

Bootstrap is proposed to quantify the uncertainty on parameter estimates (functions `bootdist` and `bootdistcens`) and also to quantify the uncertainty on CDF or quantiles estimated from the fitted distribution (`quantile` and `CIcdfplot`).

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

bootdist

Bootstrap simulation of uncertainty for non-censored data

Description

Uses parametric or nonparametric bootstrap resampling in order to simulate uncertainty in the parameters of the distribution fitted to non-censored data.

Usage

```
bootdist(f, bootmethod = "param", niter = 1001, silent = TRUE,
         parallel = c("no", "snow", "multicore"), ncpus)
## S3 method for class 'bootdist'
print(x, ...)
## S3 method for class 'bootdist'
plot(x, main = "Bootstrapped values of parameters", enhance = FALSE,
      trueval = NULL, rampcol = NULL, nbgrid = 100, nbcol = 100, ...)
## S3 method for class 'bootdist'
summary(object, ...)
## S3 method for class 'bootdist'
density(..., bw = nrd0, adjust = 1, kernel = "gaussian")
## S3 method for class 'density.bootdist'
plot(x, mar=c(4,4,2,1), lty=NULL, col=NULL, lwd=NULL, ...)
## S3 method for class 'density.bootdist'
print(x, ...)
```

Arguments

<code>f</code>	An object of class "fitdist", output of the <code>fitdist</code> function.
<code>bootmethod</code>	A character string coding for the type of resampling : "param" for a parametric resampling and "nonparam" for a nonparametric resampling of data.
<code>niter</code>	The number of samples drawn by bootstrap.
<code>silent</code>	A logical to remove or show warnings and errors when bootstrapping.
<code>parallel</code>	The type of parallel operation to be used, "snow" or "multicore" (the second one not being available on Windows), or "no" if no parallel operation.

ncpus	Number of processes to be used in parallel operation : typically one would fix it to the number of available CPUs.
x	An object of class "bootdist" or "density.bootdist".
object	An object of class "bootdist".
main	an overall title for the plot: see title , default to "Bootstrapped values of parameters".
enhance	a logical to get an enhanced plot.
trueval	when relevant, a numeric vector with the true value of parameters (for backfitting purposes).
rampcol	colors to interpolate; must be a valid argument to colorRampPalette() .
nbgrid	Number of grid points in each direction. Can be scalar or a length-2 integer vector.
nbcol	An integer argument, the required number of colors
...	Further arguments to be passed to generic methods or "bootdist" objects for density.
bw, adjust, kernel	resp. the smoothing bandwidth, the scaling factor, the kernel used, see density .
mar	A numerical vector of the form c(bottom, left, top, right), see par .
lty, col, lwd	resp. the line type, the color, the line width, see par .

Details

Samples are drawn by parametric bootstrap (resampling from the distribution fitted by [fitdist](#)) or nonparametric bootstrap (resampling with replacement from the data set). On each bootstrap sample the function [mledist](#) (or [mmedist](#), [qmedist](#), [mgedist](#) according to the component `f$method` of the object of class "fitdist") is used to estimate bootstrapped values of parameters. When that function fails to converge, NA values are returned. Medians and 2.5 and 97.5 percentiles are computed by removing NA values. The medians and the 95 percent confidence intervals of parameters (2.5 and 97.5 percentiles) are printed in the summary. If inferior to the whole number of iterations, the number of iterations for which the function converges is also printed in the summary.

By default (when `enhance=FALSE`), the plot of an object of class "bootdist" consists in a scatterplot or a matrix of scatterplots of the bootstrapped values of parameters. It uses the function [stripchart](#) when the fitted distribution is characterized by only one parameter, the function [plot](#) when there are two parameters and the function [pairs](#) in other cases. In these last cases, it provides a representation of the joint uncertainty distribution of the fitted parameters.

When `enhance=TRUE`, a personalized plot version of [pairs](#) is used where upper graphs are scatterplots and lower graphs are heatmap image using [image](#) based on a kernel based estimator for the 2D density function (using [kde2d](#) from MASS package). Arguments `rampcol`, `nbgrid`, `nbcol` can be used to customize the plots. Defaults values are `rampcol=c("green", "yellow", "orange", "red")`, `nbcol=100` (see [colorRampPalette\(\)](#)), `nbgrid=100` (see [kde2d](#)). In addition, when fitting parameters on simulated datasets for backtesting purposes, an additional argument `trueval` can be used to plot a cross at the true value.

It is possible to accelerate the bootstrap using parallelization. We recommend you to use `parallel = "multicore"`, or `parallel = "snow"` if you work on Windows, and to fix `ncpus` to the number of available processors.

density computes the empirical density of bootdist objects using the [density](#) function (with Gaussian kernel by default). It returns an object of class `density.bootdist` for which `print` and `plot` methods are provided.

Value

bootdist returns an object of class "bootdist", a list with 6 components,

estim	a data frame containing the bootstrapped values of parameters.
converg	a vector containing the codes for convergence obtained if an iterative method is used to estimate parameters on each bootstrapped data set (and 0 if a closed formula is used).
method	A character string coding for the type of resampling : "param" for a parametric resampling and "nonparam" for a nonparametric resampling.
nbboot	The number of samples drawn by bootstrap.
CI	bootstrap medians and 95 percent confidence percentile intervals of parameters.
fitpart	The object of class "fitdist" on which the bootstrap procedure was applied.

Generic functions:

`print` The print of a "bootdist" object shows the bootstrap parameter estimates. If inferior to the whole number of bootstrap iterations, the number of iterations for which the estimation converges is also printed.

`summary` The summary provides the median and 2.5 and 97.5 percentiles of each parameter. If inferior to the whole number of bootstrap iterations, the number of iterations for which the estimation converges is also printed in the summary.

`plot` The plot shows the bootstrap estimates with [stripchart](#) function for univariate parameters and [plot](#) function for multivariate parameters.

`density` The density computes empirical densities and return an object of class `density.bootdist`.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

Cullen AC and Frey HC (1999), *Probabilistic techniques in exposure assessment*. Plenum Press, USA, pp. 181-241.

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, [doi:10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [fitdistrplus](#) for an overview of the package. [fitdist](#), [mledist](#), [qmedist](#), [mmedist](#), [mgedist](#), [quantile.bootdist](#) for another generic function to calculate quantiles from the fitted distribution and its bootstrap results and [CIcdfplot](#) for adding confidence intervals on quantiles to a CDF plot of the fitted distribution.

Please visit the [Frequently Asked Questions](#).

Examples

```
# We choose a low number of bootstrap replicates in order to satisfy CRAN running times
# constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.

set.seed(123) # here just to make random sampling reproducible

# (1) Fit of a gamma distribution to serving size data
# using default method (maximum likelihood estimation)
# followed by parametric bootstrap
#
data(groundbeef)
x1 <- groundbeef$serving
f1 <- fitdist(x1, "gamma")
b1 <- bootdist(f1, niter=51)
print(b1)
plot(b1)
plot(b1, enhance=TRUE)
summary(b1)
quantile(b1)
CIcdfplot(b1, CI.output = "quantile")
density(b1)
plot(density(b1))

# (2) non parametric bootstrap on the same fit
#
b1b <- bootdist(f1, bootmethod="nonparam", niter=51)
summary(b1b)
quantile(b1b)

# (3) Fit of a normal distribution on acute toxicity values of endosulfan in log10 for
# nonarthropod invertebrates, using maximum likelihood estimation
# to estimate what is called a species sensitivity distribution
# (SSD) in ecotoxicology, followed by estimation of the 5 percent quantile value of
# the fitted distribution, what is called the 5 percent hazardous concentration (HC5)
# in ecotoxicology, with its two-sided 95 percent confidence interval calculated by
# parametric bootstrap
#
data(endosulfan)
ATV <- subset(endosulfan, group == "NonArthroInvert")$ATV
log10ATV <- log10(subset(endosulfan, group == "NonArthroInvert")$ATV)
f1n <- fitdist(log10ATV, "norm")
b1n <- bootdist(f1n, bootmethod = "param", niter=51)
quantile(b1n, probs = c(0.05, 0.1, 0.2))

# (4) comparison of sequential and parallel versions of bootstrap
# to be tried with a greater number of iterations (1001 or more)
#

niter <- 1001
data(groundbeef)
```

```

x1 <- groundbeef$serving
f1 <- fitdist(x1, "gamma")

# sequential version
ptm <- proc.time()
summary(bootdist(f1, niter = niter))
proc.time() - ptm

# parallel version using snow
require("parallel")
ptm <- proc.time()
summary(bootdist(f1, niter = niter, parallel = "snow", ncpus = 2))
proc.time() - ptm

# parallel version using multicore (not available on Windows)
ptm <- proc.time()
summary(bootdist(f1, niter = niter, parallel = "multicore", ncpus = 2))
proc.time() - ptm

```

bootdistcens

Bootstrap simulation of uncertainty for censored data

Description

Uses nonparametric bootstrap resampling in order to simulate uncertainty in the parameters of the distribution fitted to censored data.

Usage

```

bootdistcens(f, niter = 1001, silent = TRUE,
             parallel = c("no", "snow", "multicore"), ncpus)
## S3 method for class 'bootdistcens'
print(x, ...)
## S3 method for class 'bootdistcens'
plot(x, ...)
## S3 method for class 'bootdistcens'
summary(object, ...)
## S3 method for class 'bootdistcens'
density(..., bw = nrd0, adjust = 1, kernel = "gaussian")
## S3 method for class 'density.bootdistcens'
plot(x, mar=c(4,4,2,1), lty=NULL, col=NULL, lwd=NULL, ...)
## S3 method for class 'density.bootdistcens'
print(x, ...)

```

Arguments

<code>f</code>	An object of class "fitdiscens", output of the fitdiscens function.
<code>niter</code>	The number of samples drawn by bootstrap.
<code>silent</code>	A logical to remove or show warnings and errors when bootstrapping.
<code>parallel</code>	The type of parallel operation to be used, "snow" or "multicore" (the second one not being available on Windows), or "no" if no parallel operation.
<code>ncpus</code>	Number of processes to be used in parallel operation : typically one would fix it to the number of available CPUs.
<code>x</code>	An object of class "bootdiscens".
<code>object</code>	An object of class "bootdiscens".
<code>...</code>	Further arguments to be passed to generic methods or "bootdiscens" objects for density.
<code>bw, adjust, kernel</code>	resp. the smoothing bandwidth, the scaling factor, the kernel used, see density .
<code>mar</code>	A numerical vector of the form <code>c(bottom, left, top, right)</code> , see par .
<code>lty, col, lwd</code>	resp. the line type, the color, the line width, see par .

Details

Samples are drawn by nonparametric bootstrap (resampling with replacement from the data set). On each bootstrap sample the function [mledist](#) is used to estimate bootstrapped values of parameters. When [mledist](#) fails to converge, NA values are returned. Medians and 2.5 and 97.5 percentiles are computed by removing NA values. The medians and the 95 percent confidence intervals of parameters (2.5 and 97.5 percentiles) are printed in the summary. If inferior to the whole number of iterations, the number of iterations for which [mledist](#) converges is also printed in the summary.

The plot of an object of class "bootdiscens" consists in a scatterplot or a matrix of scatterplots of the bootstrapped values of parameters. It uses the function [stripchart](#) when the fitted distribution is characterized by only one parameter, and the function [plot](#) in other cases. In these last cases, it provides a representation of the joint uncertainty distribution of the fitted parameters.

It is possible to accelerate the bootstrap using parallelization. We recommend you to use `parallel = "multicore"`, or `parallel = "snow"` if you work on Windows, and to fix `ncpus` to the number of available processors.

`density` computes the empirical density of `bootdiscens` objects using the [density](#) function (with Gaussian kernel by default). It returns an object of class `density.bootdiscens` for which `print` and `plot` methods are provided.

Value

`bootdiscens` returns an object of class "bootdiscens", a list with 6 components,

<code>estim</code>	a data frame containing the bootstrapped values of parameters.
<code>converg</code>	a vector containing the codes for convergence of the iterative method used to estimate parameters on each bootstrapped data set.
<code>method</code>	A character string coding for the type of resampling : in this case "nonparam" as it is the only available method for censored data.

nbboot	The number of samples drawn by bootstrap.
CI	bootstrap medians and 95 percent confidence percentile intervals of parameters.
fitpart	The object of class "fitdistcens" on which the bootstrap procedure was applied.

Generic functions:

- `print` The print of a "bootdistcens" object shows the bootstrap parameter estimates. If inferior to the whole number of bootstrap iterations, the number of iterations for which the estimation converges is also printed.
- `summary` The summary provides the median and 2.5 and 97.5 percentiles of each parameter. If inferior to the whole number of bootstrap iterations, the number of iterations for which the estimation converges is also printed in the summary.
- `plot` The plot shows the bootstrap estimates with the `stripchart` function for univariate parameters and `plot` function for multivariate parameters.
- `density` The density computes empirical densities and return an object of class `density.bootdistcens`.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

- Cullen AC and Frey HC (1999), *Probabilistic techniques in exposure assessment*. Plenum Press, USA, pp. 181-241.
- Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, [doi:10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See `fitdistrplus` for an overview of the package. `fitdistcens`, `mledist`, `quantile.bootdistcens` for another generic function to calculate quantiles from the fitted distribution and its bootstrap results and `CIcdfplot` for adding confidence intervals on quantiles to a CDF plot of the fitted distribution.

Please visit the [Frequently Asked Questions](#).

Examples

```
# We choose a low number of bootstrap replicates in order to satisfy CRAN running times
# constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.

set.seed(123) # here just to make random sampling reproducible

# (1) Fit of a normal distribution to fluazinam data in log10
# followed by nonparametric bootstrap and calculation of quantiles
# with 95 percent confidence intervals
#
data(fluazinam)
```

```
(d1 <-log10(fluazinam))
f1 <- fitdistcens(d1, "norm")
b1 <- bootdistcens(f1, niter = 51)
b1
summary(b1)
plot(b1)
quantile(b1)
CIcdfplot(b1, CI.output = "quantile")
plot(density(b1))

# (2) Estimation of the mean of the normal distribution
# by maximum likelihood with the standard deviation fixed at 1
# using the argument fix.arg
# followed by nonparametric bootstrap
# and calculation of quantiles with 95 percent confidence intervals
#
f1b <- fitdistcens(d1, "norm", start = list(mean = 1),fix.arg = list(sd = 1))
b1b <- bootdistcens(f1b, niter = 51)
summary(b1b)
plot(b1b)
quantile(b1b)

# (3) comparison of sequential and parallel versions of bootstrap
# to be tried with a greater number of iterations (1001 or more)
#

niter <- 1001
data(fluazinam)
d1 <-log10(fluazinam)
f1 <- fitdistcens(d1, "norm")

# sequential version
ptm <- proc.time()
summary(bootdistcens(f1, niter = niter))
proc.time() - ptm

# parallel version using snow
require("parallel")
ptm <- proc.time()
summary(bootdistcens(f1, niter = niter, parallel = "snow", ncpus = 2))
proc.time() - ptm

# parallel version using multicore (not available on Windows)
ptm <- proc.time()
summary(bootdistcens(f1, niter = niter, parallel = "multicore", ncpus = 2))
proc.time() - ptm
```

CIcdfplot

*Empirical cumulative distribution function with pointwise confidence intervals on probabilities or on quantiles***Description**

cdfband plots the empirical cumulative distribution function with the bootstrapped pointwise confidence intervals on probabilities or on quantiles.

Usage

```
CIcdfplot(b, CI.output, CI.type = "two.sided", CI.level = 0.95, CI.col = "red",
  CI.lty = 2, CI.fill = NULL, CI.only = FALSE, xlim, ylim, xlogscale = FALSE,
  ylogscale = FALSE, main, xlab, ylab, datapch, datacol, fitlty, fitcol, fitlwd,
  horizontals = TRUE, verticals = FALSE, do.points = TRUE, use.ppoints = TRUE,
  a.ppoints = 0.5, name.points = NULL, lines01 = FALSE, plotstyle = "graphics", ...)
```

Arguments

b	One "bootdist" object.
CI.output	The quantity on which (bootstrapped) bootstrapped confidence intervals are computed: either "probability" or "quantile".
CI.type	Type of confidence intervals: either "two.sided" or one-sided intervals ("less" or "greater").
CI.level	The confidence level.
CI.col	the color of the confidence intervals.
CI.lty	the line type of the confidence intervals.
CI.fill	a color to fill the confidence area. Default is NULL corresponding to no filling.
CI.only	A logical whether to plot empirical and fitted distribution functions or only the confidence intervals. Default to FALSE.
xlim	The x -limits of the plot.
ylim	The y -limits of the plot.
xlogscale	If TRUE, uses a logarithmic scale for the x -axis.
ylogscale	If TRUE, uses a logarithmic scale for the y -axis.
main	A main title for the plot, see also title .
xlab	A label for the x -axis, defaults to a description of x .
ylab	A label for the y -axis, defaults to a description of y .
datapch	An integer specifying a symbol to be used in plotting data points, see also points (only for non censored data).
datacol	A specification of the color to be used in plotting data points.
fitcol	A (vector of) color(s) to plot fitted distributions. If there are fewer colors than fits they are recycled in the standard fashion.

<code>fitlty</code>	A (vector of) line type(s) to plot fitted distributions/densities. If there are fewer values than fits they are recycled in the standard fashion. See also <code>par</code> .
<code>fitlwd</code>	A (vector of) line size(s) to plot fitted distributions/densities. If there are fewer values than fits they are recycled in the standard fashion. See also <code>par</code> .
<code>horizontal</code>	If TRUE, draws horizontal lines for the step empirical cdf function (only for non censored data). See also <code>plot.stepfun</code> .
<code>vertical</code>	If TRUE, draws also vertical lines for the empirical cdf function. Only taken into account if <code>horizontal=TRUE</code> (only for non censored data).
<code>do.points</code>	logical; if TRUE, also draw points at the x-locations. Default is TRUE (only for non censored data).
<code>use.ppoints</code>	If TRUE, probability points of the empirical distribution are defined using function <code>ppoints</code> as $(1:n - a.ppoints)/(n - 2a.ppoints + 1)$ (only for non censored data). If FALSE, probability points are simply defined as $(1:n)/n$. This argument is ignored for discrete data.
<code>a.ppoints</code>	If <code>use.ppoints=TRUE</code> , this is passed to function <code>ppoints</code> (only for non censored data).
<code>name.points</code>	Label vector for points if they are drawn i.e. if <code>do.points = TRUE</code> (only for non censored data).
<code>lines01</code>	A logical to plot two horizontal lines at $h=0$ and $h=1$ for <code>cdfcomp</code> .
<code>plotstyle</code>	"graphics" or "ggplot". If "graphics", the display is built with <code>graphics</code> functions. If "ggplot", a graphic object output is created with <code>ggplot2</code> functions (the <code>ggplot2</code> package must be installed).
<code>...</code>	Further graphical arguments passed to <code>matline</code> or <code>polygon</code> , respectively when <code>CI.fill=FALSE</code> and <code>CI.fill=TRUE</code> .

Details

`CIcdfplot` provides a plot of the empirical distribution using `cdfcomp` or `cdfcompcons`, with bootstrapped pointwise confidence intervals on probabilities (y values) or on quantiles (x values). Each interval is computed by evaluating the quantity of interest (probability associated to an x value or quantile associated to an y value) using all the bootstrapped values of parameters to get a bootstrapped sample of the quantity of interest and then by calculating percentiles on this sample to get a confidence interval (classically 2.5 and 97.5 percentiles for a 95 percent confidence level). If `CI.fill != NULL`, then the whole confidence area is filled by the color `CI.fill` thanks to the function `polygon`, otherwise only borders are drawn thanks to the function `matline`. Further graphical arguments can be passed to these functions using the three dots arguments `...`

Author(s)

Christophe Dutang and Marie-Laure Delignette-Muller.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:10.18637/jss.v064.i04.

See Also

See also [cdfcomp](#), [cdfcompens](#), [bootdist](#) and [quantile](#).

Please visit the [Frequently Asked Questions](#).

Examples

```
# We choose a low number of bootstrap replicates in order to satisfy CRAN running times
# constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.

set.seed(123) # here just to make random sampling reproducible

if (requireNamespace ("ggplot2", quietly = TRUE)) {ggplotEx <- TRUE}

# (1) Fit of an exponential distribution
#

s1 <- rexp(50, 1)
f1 <- fitdist(s1, "exp")
b1 <- bootdist(f1, niter= 11) #voluntarily low to decrease computation time

# plot 95 percent bilateral confidence intervals on y values (probabilities)
CIcdfplot(b1, CI.level= 95/100, CI.output = "probability")
if (ggplotEx) CIcdfplot(b1, CI.level= 95/100, CI.output = "probability", plotstyle = "ggplot")

# plot of the previous intervals as a band
CIcdfplot(b1, CI.level= 95/100, CI.output = "probability",
  CI.fill = "pink", CI.col = "red")
if (ggplotEx) CIcdfplot(b1, CI.level= 95/100, CI.output = "probability",
  CI.fill = "pink", CI.col = "red", plotstyle = "ggplot")

# plot of the previous intervals as a band without empirical and fitted dist. functions
CIcdfplot(b1, CI.level= 95/100, CI.output = "probability", CI.only = TRUE,
  CI.fill = "pink", CI.col = "red")
if (ggplotEx) CIcdfplot(b1, CI.level= 95/100, CI.output = "probability", CI.only = TRUE,
  CI.fill = "pink", CI.col = "red", plotstyle = "ggplot")

# same plot without contours
CIcdfplot(b1, CI.level= 95/100, CI.output = "probability", CI.only = TRUE,
  CI.fill = "pink", CI.col = "pink")
if (ggplotEx) CIcdfplot(b1, CI.level= 95/100, CI.output = "probability", CI.only = TRUE,
  CI.fill = "pink", CI.col = "pink", plotstyle = "ggplot")

# plot 95 percent bilateral confidence intervals on x values (quantiles)
CIcdfplot(b1, CI.level= 95/100, CI.output = "quantile")
if (ggplotEx) CIcdfplot(b1, CI.level= 95/100, CI.output = "quantile", plotstyle = "ggplot")

# plot 95 percent unilateral confidence intervals on quantiles
CIcdfplot(b1, CI.level = 95/100, CI.output = "quant", CI.type = "less",
  CI.fill = "grey80", CI.col = "black", CI.lty = 1)
```

```

if (ggplotEx) CIcdfplot(b1, CI.level = 95/100, CI.output = "quant", CI.type = "less",
  CI.fill = "grey80", CI.col = "black", CI.lty = 1, plotstyle = "ggplot")

CIcdfplot(b1, CI.level= 95/100, CI.output = "quant", CI.type = "greater",
  CI.fill = "grey80", CI.col = "black", CI.lty = 1)
if (ggplotEx) CIcdfplot(b1, CI.level= 95/100, CI.output = "quant", CI.type = "greater",
  CI.fill = "grey80", CI.col = "black", CI.lty = 1, plotstyle = "ggplot")

# (2) Fit of a normal distribution on acute toxicity log-transformed values of
# endosulfan for nonarthropod invertebrates, using maximum likelihood estimation
# to estimate what is called a species sensitivity distribution
# (SSD) in ecotoxicology, followed by estimation of the 5, 10 and 20 percent quantile
# values of the fitted distribution, which are called the 5, 10, 20 percent hazardous
# concentrations (HC5, HC10, HC20) in ecotoxicology, with their
# confidence intervals, from a small number of bootstrap
# iterations to satisfy CRAN running times constraint and plot of the band
# representing pointwise confidence intervals on any quantiles (any HCx values)
# For practical applications, we recommend to use at least niter=501 or niter=1001.
#

data(endosulfan)
log10ATV <- log10(subset(endosulfan, group == "NonArthroInvert")$ATV)
namesATV <- subset(endosulfan, group == "NonArthroInvert")$taxa
fln <- fitdist(log10ATV, "norm")
bln <- bootdist(fln, bootmethod="param", niter=101)
quantile(bln, probs = c(0.05, 0.1, 0.2))
CIcdfplot(bln, CI.output = "quantile", CI.fill = "lightblue", CI.col = "blue",
  xlim = c(0,5), name.points=namesATV)
if (ggplotEx) CIcdfplot(bln, CI.output = "quantile", CI.fill = "lightblue", CI.col = "blue",
  xlim = c(0,5), name.points=namesATV, plotstyle = "ggplot")

# (3) Same type of example as example (2) from ecotoxicology
# with censored data
#
data(salinity)
log10LC50 <-log10(salinity)
fln <- fitdistcens(log10LC50,"norm")
bln <- bootdistcens(fln, niter=101)
(HC5ln <- quantile(bln,probs = 0.05))
CIcdfplot(bln, CI.output = "quantile", CI.fill = "lightblue", CI.col = "blue",
  xlab = "log10(LC50)",xlim=c(0.5,2),lines01 = TRUE)
if (ggplotEx) CIcdfplot(bln, CI.output = "quantile", CI.fill = "lightblue", CI.col = "blue",
  xlab = "log10(LC50)",xlim=c(0.5,2),lines01 = TRUE, plotstyle = "ggplot")
# zoom around the HC5
CIcdfplot(bln, CI.output = "quantile", CI.fill = "lightblue", CI.col = "blue",
  xlab = "log10(LC50)", lines01 = TRUE, xlim = c(0.8, 1.5), ylim = c(0, 0.1))
abline(h = 0.05, lty = 2) # line corresponding to a CDF of 5 percent

if (ggplotEx) CIcdfplot(bln, CI.output = "quantile", CI.fill = "lightblue", CI.col = "blue",
  xlab = "log10(LC50)", lines01 = TRUE, xlim = c(0.8, 1.5), ylim = c(0, 0.1),
  plotstyle = "ggplot") +
  ggplot2::geom_hline(yintercept = 0.05, lty = 2) # line corresponding to a CDF of 5 percent

```

danish

Danish reinsurance claim dataset

Description

The univariate dataset was collected at Copenhagen Reinsurance and comprise 2167 fire losses over the period 1980 to 1990. They have been adjusted for inflation to reflect 1985 values and are expressed in millions of Danish Krone.

The multivariate data set is the same data as above but the total claim has been divided into a building loss, a loss of contents and a loss of profits.

Usage

```
data(danishuni)
data(danishmulti)
```

Format

`danishuni` contains two columns:

`Date` The day of claim occurrence.

`Loss` The total loss amount in millions of Danish Krone (DKK).

`danishmulti` contains five columns:

`Date` The day of claim occurrence.

`Building` The loss amount (mDKK) of the building coverage.

`Contents` The loss amount (mDKK) of the contents coverage.

`Profits` The loss amount (mDKK) of the profit coverage.

`Total` The total loss amount (mDKK).

All columns are numeric except Date columns of class Date.

Source

Embrechts, P., Kluppelberg, C. and Mikosch, T. (1997) *Modelling Extremal Events for Insurance and Finance*. Berlin: Springer.

References

Dataset used in McNeil (1996), *Estimating the Tails of Loss Severity Distributions using Extreme Value Theory*, ASTIN Bull. Davison, A. C. (2003) *Statistical Models*. Cambridge University Press. Page 278.

Examples

```
# (1) load of data
#
data(danishuni)

# (2) plot and description of data
#
plotdist(danishuni$Loss)

# (3) load of data
#
data(danishmulti)

# (4) plot and description of data
#
idx <- sample(1:NROW(danishmulti), 10)
barplot(danishmulti$Building[idx], col = "grey25",
        ylim = c(0, max(danishmulti$Total[idx])), main = "Some claims of danish data set")
barplot(danishmulti$Content[idx], add = TRUE, col = "grey50", axes = FALSE)
barplot(danishmulti$Profits[idx], add = TRUE, col = "grey75", axes = FALSE)
legend("topleft", legend = c("Building", "Content", "Profits"),
       fill = c("grey25", "grey50", "grey75"))
```

dataFAQ

Datasets for the FAQ

Description

Datasets used in the FAQ vignette.

Usage

```
data(dataFAQlog1)
data(dataFAQscale1)
data(dataFAQscale2)
```

Format

dataFAQlog1 dataFAQscale1 dataFAQscale2 are vectors of numeric data.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

descdist

*Description of an empirical distribution for non-censored data***Description**

Computes descriptive parameters of an empirical distribution for non-censored data and provides a skewness-kurtosis plot.

Usage

```
descdist(data, discrete = FALSE, boot = NULL, method = "unbiased",
graph = TRUE, print = TRUE, obs.col = "red", obs.pch = 16, boot.col = "orange")
```

```
## S3 method for class 'descdist'
print(x, ...)
```

Arguments

data	A numeric vector.
discrete	If TRUE, the distribution is considered as discrete.
boot	If not NULL, boot values of skewness and kurtosis are plotted from bootstrap samples of data. boot must be fixed in this case to an integer above 10.
method	"unbiased" for unbiased estimated values of statistics or "sample" for sample values.
graph	If FALSE, the skewness-kurtosis graph is not plotted.
print	If FALSE, the descriptive parameters computed are not printed.
obs.col	Color used for the observed point on the skewness-kurtosis graph.
obs.pch	plotting character used for the observed point on the skewness-kurtosis graph.
boot.col	Color used for bootstrap sample of points on the skewness-kurtosis graph.
x	An object of class "descdist".
...	Further arguments to be passed to generic functions

Details

Minimum, maximum, median, mean, sample sd, and sample (if method=="sample") or by default unbiased estimations of skewness and Pearson's kurtosis values are printed (Sokal and Rohlf, 1995). A skewness-kurtosis plot such as the one proposed by Cullen and Frey (1999) is given for the empirical distribution. On this plot, values for common distributions are also displayed as a tools to help the choice of distributions to fit to data. For some distributions (normal, uniform, logistic, exponential for example), there is only one possible value for the skewness and the kurtosis (for a normal distribution for example, skewness = 0 and kurtosis = 3), and the distribution is thus represented by a point on the plot. For other distributions, areas of possible values are represented, consisting in lines (gamma and lognormal distributions for example), or larger areas (beta distribution for example). The Weibull distribution is not represented on the graph but it is indicated

on the legend that shapes close to lognormal and gamma distributions may be obtained with this distribution.

In order to take into account the uncertainty of the estimated values of kurtosis and skewness from data, the data set may be bootstrapped by fixing the argument `boot` to an integer above 10. `boot` values of skewness and kurtosis corresponding to the `boot` bootstrap samples are then computed and reported in blue color on the skewness-kurtosis plot.

If `discrete` is TRUE, the represented distributions are the Poisson, negative binomial distributions, and the normal distribution to which previous discrete distributions may converge. If `discrete` is FALSE, these are uniform, normal, logistic, lognormal, beta and gamma distributions.

Value

`descdist` returns a list with 7 components,

<code>min</code>	the minimum value
<code>max</code>	the maximum value
<code>median</code>	the median value
<code>mean</code>	the mean value
<code>sd</code>	the standard deviation sample or estimated value
<code>skewness</code>	the skewness sample or estimated value
<code>kurtosis</code>	the kurtosis sample or estimated value
<code>method</code>	the method specified in input ("unbiased" for unbiased estimated values of statistics or "sample" for sample values.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

- Cullen AC and Frey HC (1999), *Probabilistic techniques in exposure assessment*. Plenum Press, USA, pp. 81-159.
- Evans M, Hastings N and Peacock B (2000), *Statistical distributions*. John Wiley and Sons Inc, [doi:10.1002/9780470627242](https://doi.org/10.1002/9780470627242).
- Sokal RR and Rohlf FJ (1995), *Biometry*. W.H. Freeman and Company, USA, pp. 111-115.
- Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, [doi:10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [plotdist](#).

Please visit the [Frequently Asked Questions](#).

Examples

```

set.seed(123) # here just to make random sampling reproducible

# (1) Description of a sample from a normal distribution
# with and without uncertainty on skewness and kurtosis estimated by bootstrap
#
x1 <- rnorm(100)
descdist(x1)
descdist(x1,boot=11)

# (2) Description of a sample from a beta distribution
# with uncertainty on skewness and kurtosis estimated by bootstrap
# with changing of default colors and plotting character for observed point
#
descdist(rbeta(100,shape1=0.05,shape2=1),boot=11,
  obs.col="blue", obs.pch = 15, boot.col="darkgreen")

# (3) Description of a sample from a gamma distribution
# with uncertainty on skewness and kurtosis estimated by bootstrap
# without plotting
#
descdist(rgamma(100,shape=2,rate=1),boot=11,graph=FALSE)

# (4) Description of a sample from a Poisson distribution
# with uncertainty on skewness and kurtosis estimated by bootstrap
#
descdist(rpois(100,lambda=2),discrete=TRUE,boot=11)

# (5) Description of serving size data
# with uncertainty on skewness and kurtosis estimated by bootstrap
#
data(groundbeef)
serving <- groundbeef$serving
descdist(serving, boot=11)

```

detectbound

Detect bounds for density function

Description

Manual detection of bounds of parameter of a density function/

Usage

```
detectbound(distname, vstart, obs, fix.arg=NULL, echo=FALSE)
```

Arguments

distname A character string "name" naming a distribution for which the corresponding density function dname must be classically defined.

vstart	A named vector giving the initial values of parameters of the named distribution.
obs	A numeric vector for non censored data.
fix.arg	An optional named vector giving the values of fixed parameters of the named distribution. Default to NULL.
echo	A logical to show some traces.

Details

This function manually tests the following bounds : -1, 0, and 1.

Value

detectbound returns a 2-row matrix with the lower bounds in the first row and the upper bounds in the second row.

Author(s)

Christophe Dutang and Marie-Laure Delignette-Muller.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:[10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [fitdist](#).

Please visit the [Frequently Asked Questions](#).

Examples

```
# case where the density returns a Not-an-Numeric value.
detectbound("exp", c(rate=3), 1:10)
detectbound("binom", c(size=3, prob=1/2), 1:10)
detectbound("nbinom", c(size=3, prob=1/2), 1:10)
```

endosulfan

Species Sensitivity Distribution (SSD) for endosulfan

Description

Summary of 48- to 96-hour acute toxicity values (LC50 and EC50 values) for exposure of Australian an Non-Australian taxa to endosulfan.

Usage

```
data(endosulfan)
```

Format

endosulfan is a data frame with 4 columns, named ATV for Acute Toxicity Value (geometric mean of LC50 ou EC50 values in micrograms per liter), Australian (coding for Australian or another origin), group (arthropods, fish or non-arthropod invertebrates) and taxa.

Source

Hose, G.C., Van den Brink, P.J. 2004. Confirming the Species-Sensitivity Distribution Concept for Endosulfan Using Laboratory, Mesocosms, and Field Data. *Archives of Environmental Contamination and Toxicology*, **47**, 511-520.

Examples

```
# (1) load of data
#
data(endosulfan)

# (2) plot and description of data for non Australian fish in decimal logarithm
#
log10ATV <- log10(subset(endosulfan, (Australian == "no") & (group == "Fish"))$ATV)
plotdist(log10ATV)
descdist(log10ATV, boot=11)

# (3) fit of a normal and a logistic distribution to data in log10
# (classical distributions used for SSD)
# and visual comparison of the fits
#
f1n <- fitdist(log10ATV, "norm")
summary(f1n)

f1l <- fitdist(log10ATV, "logis")
summary(f1l)

cdfcomp(list(f1n, f1l), legendtext=c("normal", "logistic"),
        xlab="log10ATV")

denscomp(list(f1n, f1l), legendtext=c("normal", "logistic"),
        xlab="log10ATV")

qqcomp(list(f1n, f1l), legendtext=c("normal", "logistic"))
ppcomp(list(f1n, f1l), legendtext=c("normal", "logistic"))

gofstat(list(f1n, f1l), fitnames = c("lognormal", "loglogistic"))

# (4) estimation of the 5 percent quantile value of
# logistic fitted distribution (5 percent hazardous concentration : HC5)
# with its two-sided 95 percent confidence interval calculated by
# parametric bootstrap
# with a small number of iterations to satisfy CRAN running times constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.
#
# in log10(ATV)
```

```

bll <- bootdist(fll,niter=51)
HC511 <- quantile(bll,probs = 0.05)
# in ATV
10^(HC511$quantiles)
10^(HC511$quantCI)

# (5) estimation of the 5 percent quantile value of
# the fitted logistic distribution (5 percent hazardous concentration : HC5)
# with its one-sided 95 percent confidence interval (type "greater")
# calculated by
# nonparametric bootstrap
# with a small number of iterations to satisfy CRAN running times constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.
#
# in log10(ATV)
bllnonpar <- bootdist(fll,niter=51,bootmethod = "nonparam")
HC511greater <- quantile(bllnonpar,probs = 0.05, CI.type="greater")
# in ATV
10^(HC511greater$quantiles)
10^(HC511greater$quantCI)

# (6) fit of a logistic distribution
# by minimizing the modified Anderson-Darling AD2L distance
# cf. ?mgdist for definition of this distance
#

fllAD2L <- fitdist(log10ATV,"logis",method="mge",gof="AD2L")
summary(fllAD2L)
plot(fllAD2L)

```

fitdist

Fit of univariate distributions to non-censored data

Description

Fit of univariate distributions to non-censored data by maximum likelihood (mle), moment matching (mme), quantile matching (qme) or maximizing goodness-of-fit estimation (mge). The latter is also known as minimizing distance estimation. Generic methods are print, plot, summary, quantile, logLik, AIC, BIC, vcov and coef.

Usage

```

fitdist(data, distr, method = c("mle", "mme", "qme", "mge", "mse"),
        start=NULL, fix.arg=NULL, discrete, keepdata = TRUE, keepdata.nb=100,
        calcvcov=TRUE, ...)

## S3 method for class 'fitdist'
print(x, ...)

```

```

## S3 method for class 'fitdist'
plot(x, breaks="default", ...)

## S3 method for class 'fitdist'
summary(object, ...)

## S3 method for class 'fitdist'
logLik(object, ...)

## S3 method for class 'fitdist'
AIC(object, ..., k = 2)

## S3 method for class 'fitdist'
BIC(object, ...)

## S3 method for class 'fitdist'
vcov(object, ...)

## S3 method for class 'fitdist'
coef(object, ...)

```

Arguments

data	A numeric vector.
distr	A character string "name" naming a distribution for which the corresponding density function <code>dname</code> , the corresponding distribution function <code>prname</code> and the corresponding quantile function <code>qname</code> must be defined, or directly the density function.
method	A character string coding for the fitting method: "mle" for 'maximum likelihood estimation', "mme" for 'moment matching estimation', "qme" for 'quantile matching estimation', "mge" for 'maximum goodness-of-fit estimation' and "mse" for 'maximum spacing estimation'.
start	A named list giving the initial values of parameters of the named distribution or a function of data computing initial values and returning a named list. This argument may be omitted (default) for some distributions for which reasonable starting values are computed (see the 'details' section of mledist). It may not be into account for closed-form formulas.
fix.arg	An optional named list giving the values of fixed parameters of the named distribution or a function of data computing (fixed) parameter values and returning a named list. Parameters with fixed value are thus NOT estimated by this maximum likelihood procedure. The use of this argument is not possible if <code>method="mme"</code> and a closed-form formula is used.
keepdata	a logical. If TRUE, dataset is returned, otherwise only a sample subset is returned.
keepdata.nb	When <code>keepdata=FALSE</code> , the length (>1) of the subset returned.
calcvcov	A logical indicating if (asymptotic) covariance matrix is required.

discrete	If TRUE, the distribution is considered as discrete. If discrete is missing, discrete is automatically set to TRUE when <code>distr</code> belongs to "binom", "nbinom", "geom", "hyper" or "pois" and to FALSE in the other cases. It is thus recommended to enter this argument when using another discrete distribution. This argument will not directly affect the results of the fit but will be passed to functions <code>gofstat</code> , <code>plotdist</code> and <code>cdfcomp</code> .
x	An object of class "fitdist".
object	An object of class "fitdist".
breaks	If "default" the histogram is plotted with the function <code>hist</code> with its default breaks definition. Else breaks is passed to the function <code>hist</code> . This argument is not taken into account with discrete distributions: "binom", "nbinom", "geom", "hyper" and "pois".
k	penalty per parameter to be passed to the AIC generic function (2 by default).
...	Further arguments to be passed to generic functions, or to one of the functions "mledist", "mmedist", "qmedist" or "mgedist" depending of the chosen method. See <code>mledist</code> , <code>mmedist</code> , <code>qmedist</code> , <code>mgedist</code> for details on parameter estimation.

Details

It is assumed that the `distr` argument specifies the distribution by the probability density function, the cumulative distribution function and the quantile function (d, p, q).

The four possible fitting methods are described below:

When `method="mle"` Maximum likelihood estimation consists in maximizing the log-likelihood. A numerical optimization is carried out in `mledist` via `optim` to find the best values (see `mledist` for details).

When `method="mme"` Moment matching estimation consists in equalizing theoretical and empirical moments. Estimated values of the distribution parameters are computed by a closed-form formula for the following distributions : "norm", "lnorm", "pois", "exp", "gamma", "nbinom", "geom", "beta", "unif" and "logis". Otherwise the theoretical and the empirical moments are matched numerically, by minimization of the sum of squared differences between observed and theoretical moments. In this last case, further arguments are needed in the call to `fitdist`: `order` and `memp` (see `mmedist` for details).

Since Version 1.2-0, `mmedist` automatically computes the asymptotic covariance matrix, hence the theoretical moments `mdist` should be defined up to an order which equals to twice the maximal order given `order`.

When `method="qme"` Quantile matching estimation consists in equalizing theoretical and empirical quantile. A numerical optimization is carried out in `qmedist` via `optim` to minimize of the sum of squared differences between observed and theoretical quantiles. The use of this method requires an additional argument `probs`, defined as the numeric vector of the probabilities for which the quantile(s) is(are) to be matched (see `qmedist` for details).

When `method="mge"` Maximum goodness-of-fit estimation consists in maximizing a goodness-of-fit statistics. A numerical optimization is carried out in `mgedist` via `optim` to minimize the goodness-of-fit distance. The use of this method requires an additional argument `gof` coding for the goodness-of-fit distance chosen. One can use the classical Cramer-von Mises distance

("CvM"), the classical Kolmogorov-Smirnov distance ("KS"), the classical Anderson-Darling distance ("AD") which gives more weight to the tails of the distribution, or one of the variants of this last distance proposed by Luceno (2006) (see [mgedist](#) for more details). This method is not suitable for discrete distributions.

When `method = "mse"` Maximum goodness-of-fit estimation consists in maximizing the average log spacing. A numerical optimization is carried out in [msedist](#) via `optim`.

By default, direct optimization of the log-likelihood (or other criteria depending of the chosen method) is performed using `optim`, with the "Nelder-Mead" method for distributions characterized by more than one parameter and the "BFGS" method for distributions characterized by only one parameter. The optimization algorithm used in `optim` can be chosen or another optimization function can be specified using `...` argument (see [mledist](#) for details). `start` may be omitted (i.e. NULL) for some classic distributions (see the 'details' section of [mledist](#)). Note that when errors are raised by `optim`, it's a good idea to start by adding traces during the optimization process by adding `control=list(trace=1, REPORT=1)` in `...` argument.

Once the parameter(s) is(are) estimated, `fitdist` computes the log-likelihood for every estimation method and for maximum likelihood estimation the standard errors of the estimates calculated from the Hessian at the solution found by `optim` or by the user-supplied function passed to `mledist`.

By default (`keepdata = TRUE`), the object returned by `fitdist` contains the data vector given in input. When dealing with large datasets, we can remove the original dataset from the output by setting `keepdata = FALSE`. In such a case, only `keepdata.nb` points (at most) are kept by random subsampling `keepdata.nb-2` points from the dataset and adding the minimum and the maximum. If combined with [bootdist](#), and use with non-parametric bootstrap be aware that bootstrap is performed on the subset randomly selected in `fitdist`. Currently, the graphical comparisons of multiple fits is not available in this framework.

Weighted version of the estimation process is available for `method = "mle"`, `"mme"`, `"qme"` by using `weights=...`. See the corresponding man page for details. Weighted maximum GOF estimation (when `method = "mge"`) is not allowed. It is not yet possible to take into account weights in functions `plotdist`, `plot.fitdist`, `cdfcomp`, `denscomp`, `ppcomp`, `qqcomp`, `gofstat` and `descdist` (developments planned in the future).

Once the parameter(s) is(are) estimated, [gofstat](#) allows to compute goodness-of-fit statistics.

NB: if your data values are particularly small or large, a scaling may be needed before the optimization process. See example (14) in this man page and examples (14,15) in the test file of the package. Please also take a look at the [Rmpfr](#) package available on CRAN for numerical accuracy issues.

Value

`fitdist` returns an object of class `"fitdist"`, a list with the following components:

<code>estimate</code>	the parameter estimates.
<code>method</code>	the character string coding for the fitting method : <code>"mle"</code> for 'maximum likelihood estimation', <code>"mme"</code> for 'matching moment estimation', <code>"qme"</code> for 'matching quantile estimation' <code>"mge"</code> for 'maximum goodness-of-fit estimation' and <code>"mse"</code> for 'maximum spacing estimation'.
<code>sd</code>	the estimated standard errors, NA if numerically not computable or NULL if not available.

cor	the estimated correlation matrix, NA if numerically not computable or NULL if not available.
vcov	the estimated variance-covariance matrix, NULL if not available for the estimation method considered.
loglik	the log-likelihood.
aic	the Akaike information criterion.
bic	the the so-called BIC or SBC (Schwarz Bayesian criterion).
n	the length of the data set.
data	the data set.
distname	the name of the distribution.
fix.arg	the named list giving the values of parameters of the named distribution that must be kept fixed rather than estimated by maximum likelihood or NULL if there are no such parameters.
fix.arg.fun	the function used to set the value of fix.arg or NULL.
dots	the list of further arguments passed in ... to be used in <code>bootdist</code> in iterative calls to <code>mledist</code> , <code>mmedist</code> , <code>qmedist</code> , <code>mgedist</code> or NULL if no such arguments.
convergence	an integer code for the convergence of <code>optim/constrOptim</code> defined as below or defined by the user in the user-supplied optimization function. 0 indicates successful convergence. 1 indicates that the iteration limit of <code>optim</code> has been reached. 10 indicates degeneracy of the Nelder-Mead simplex. 100 indicates that <code>optim</code> encountered an internal error.
discrete	the input argument or the automatic definition by the function to be passed to functions <code>gofstat</code> , <code>plotdist</code> and <code>cdfcomp</code> .
weights	the vector of weights used in the estimation process or NULL.

Generic functions:

`print` The print of a "fitdist" object shows few traces about the fitting method and the fitted distribution.

`summary` The summary provides the parameter estimates of the fitted distribution, the log-likelihood, AIC and BIC statistics and when the maximum likelihood is used, the standard errors of the parameter estimates and the correlation matrix between parameter estimates.

`plot` The plot of an object of class "fitdist" returned by `fitdist` uses the function `plotdist`. An object of class "fitdist" or a list of objects of class "fitdist" corresponding to various fits using the same data set may also be plotted using a cdf plot (function `cdfcomp`), a density plot (function `denscomp`), a density Q-Q plot (function `qqcomp`), or a P-P plot (function `ppcomp`).

`logLik` Extracts the estimated log-likelihood from the "fitdist" object.

`AIC` Extracts the AIC from the "fitdist" object.

`BIC` Extracts the estimated BIC from the "fitdist" object.

`vcov` Extracts the estimated var-covariance matrix from the "fitdist" object (only available When `method = "mle"`).

`coef` Extracts the fitted coefficients from the "fitdist" object.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

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See Also

See [fitdistrplus](#) for an overview of the package. See [mledist](#), [mmedist](#), [qmedist](#), [mgedist](#), [msedist](#) for details on parameter estimation. See [gofstat](#) for goodness-of-fit statistics. See [plotdist](#), [graphcomp](#), [CIcdfplot](#) for graphs (with or without uncertainty and/or multiple fits). See [llplot](#) for (log-)likelihood plots in the neighborhood of the fitted value. See [bootdist](#) for bootstrap procedures and [fitdistcens](#) for censored-data fitting methods. See [optim](#) for base R optimization procedures. See [quantile.fitdist](#), another generic function, which calculates quantiles from the fitted distribution. See [quantile](#) for base R quantile computation.

Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) fit of a gamma distribution by maximum likelihood estimation
#
data(groundbeef)
serving <- groundbeef$serving
fitg <- fitdist(serving, "gamma")
summary(fitg)
plot(fitg)
plot(fitg, demp = TRUE)
plot(fitg, histo = FALSE, demp = TRUE)
cdfcomp(fitg, addlegend=FALSE)
denscomp(fitg, addlegend=FALSE)
ppcomp(fitg, addlegend=FALSE)
qqcomp(fitg, addlegend=FALSE)

# (2) use the moment matching estimation (using a closed formula)
#
```

```

fitgmme <- fitdist(serving, "gamma", method="mme")
summary(fitgmme)

# (3) Comparison of various fits
#

fitW <- fitdist(serving, "weibull")
fitg <- fitdist(serving, "gamma")
fitln <- fitdist(serving, "lnorm")
summary(fitW)
summary(fitg)
summary(fitln)
cdfcomp(list(fitW, fitg, fitln), legendtext=c("Weibull", "gamma", "lognormal"))
denscomp(list(fitW, fitg, fitln), legendtext=c("Weibull", "gamma", "lognormal"))
qqcomp(list(fitW, fitg, fitln), legendtext=c("Weibull", "gamma", "lognormal"))
ppcomp(list(fitW, fitg, fitln), legendtext=c("Weibull", "gamma", "lognormal"))
gofstat(list(fitW, fitg, fitln), fitnames=c("Weibull", "gamma", "lognormal"))

# (4) defining your own distribution functions, here for the Gumbel distribution
# for other distributions, see the CRAN task view
# dedicated to probability distributions
#

dgumbel <- function(x, a, b) 1/b*exp((a-x)/b)*exp(-exp((a-x)/b))
pgumbel <- function(q, a, b) exp(-exp((a-q)/b))
qgumbel <- function(p, a, b) a-b*log(-log(p))

fitgumbel <- fitdist(serving, "gumbel", start=list(a=10, b=10))
summary(fitgumbel)
plot(fitgumbel)

# (5) fit discrete distributions (Poisson and negative binomial)
#

data(toxocara)
number <- toxocara$number
fitp <- fitdist(number, "pois")
summary(fitp)
plot(fitp)
fitnb <- fitdist(number, "nbinom")
summary(fitnb)
plot(fitnb)

cdfcomp(list(fitp, fitnb))
gofstat(list(fitp, fitnb))

# (6) how to change the optimisation method?
#

data(groundbeef)
serving <- groundbeef$serving
fitdist(serving, "gamma", optim.method="Nelder-Mead")

```

```

fitdist(serving, "gamma", optim.method="BFGS")
fitdist(serving, "gamma", optim.method="SANN")

# (7) custom optimization function
#

#create the sample
mysample <- rexp(100, 5)
mystart <- list(rate=8)

res1 <- fitdist(mysample, dexp, start= mystart, optim.method="Nelder-Mead")

#show the result
summary(res1)

#the warning tell us to use optimise, because the Nelder-Mead is not adequate.

#to meet the standard 'fn' argument and specific name arguments, we wrap optimize,
myoptimize <- function(fn, par, ...)
{
  res <- optimize(f=fn, ..., maximum=FALSE)
  #assume the optimization function minimize

  standardres <- c(res, convergence=0, value=res$objective,
                  par=res$minimum, hessian=NA)

  return(standardres)
}

#call fitdist with a 'custom' optimization function
res2 <- fitdist(mysample, "exp", start=mystart, custom.optim=myoptimize,
               interval=c(0, 100))

#show the result
summary(res2)

# (8) custom optimization function - another example with the genetic algorithm
#

#set a sample
fit1 <- fitdist(serving, "gamma")
summary(fit1)

#wrap genoud function rgenoud package
mygenoud <- function(fn, par, ...)
{
  require("rgenoud")
  res <- genoud(fn, starting.values=par, ...)
  standardres <- c(res, convergence=0)

  return(standardres)
}

```

```

}

#call fitdist with a 'custom' optimization function
fit2 <- fitdist(serving, "gamma", custom.optim=mygenoud, nvars=2,
  Domains=cbind(c(0, 0), c(10, 10)), boundary.enforcement=1,
  print.level=1, hessian=TRUE)

summary(fit2)

# (9) estimation of the standard deviation of a gamma distribution
# by maximum likelihood with the shape fixed at 4 using the argument fix.arg
#
data(groundbeef)
serving <- groundbeef$serving
f1c <- fitdist(serving, "gamma", start=list(rate=0.1), fix.arg=list(shape=4))
summary(f1c)
plot(f1c)

# (10) fit of a Weibull distribution to serving size data
# by maximum likelihood estimation
# or by quantile matching estimation (in this example
# matching first and third quartiles)
#
data(groundbeef)
serving <- groundbeef$serving
fWmle <- fitdist(serving, "weibull")
summary(fWmle)
plot(fWmle)
gofstat(fWmle)

fWqme <- fitdist(serving, "weibull", method="qme", probs=c(0.25, 0.75))
summary(fWqme)
plot(fWqme)
gofstat(fWqme)

# (11) Fit of a Pareto distribution by numerical moment matching estimation
#

require("actuar")
#simulate a sample
x4 <- rpareto(1000, 6, 2)

#empirical raw moment
memp <- function(x, order) mean(x^order)

#fit
fP <- fitdist(x4, "pareto", method="mme", order=c(1, 2), memp="memp",
  start=list(shape=10, scale=10), lower=1, upper=Inf)

```

```

summary(fP)
plot(fP)

# (12) Fit of a Weibull distribution to serving size data by maximum
# goodness-of-fit estimation using all the distances available
#

data(groundbeef)
serving <- groundbeef$serving
(f1 <- fitdist(serving, "weibull", method="mge", gof="CvM"))
(f2 <- fitdist(serving, "weibull", method="mge", gof="KS"))
(f3 <- fitdist(serving, "weibull", method="mge", gof="AD"))
(f4 <- fitdist(serving, "weibull", method="mge", gof="ADR"))
(f5 <- fitdist(serving, "weibull", method="mge", gof="ADL"))
(f6 <- fitdist(serving, "weibull", method="mge", gof="AD2R"))
(f7 <- fitdist(serving, "weibull", method="mge", gof="AD2L"))
(f8 <- fitdist(serving, "weibull", method="mge", gof="AD2"))
cdfcomp(list(f1, f2, f3, f4, f5, f6, f7, f8))
cdfcomp(list(f1, f2, f3, f4, f5, f6, f7, f8),
  xlogscale=TRUE, xlim=c(8, 250), verticals=TRUE)
denscomp(list(f1, f2, f3, f4, f5, f6, f7, f8))

# (13) Fit of a uniform distribution using maximum likelihood
# (a closed formula is used in this special case where the loglikelihood is not defined),
# or maximum goodness-of-fit with Cramer-von Mises or Kolmogorov-Smirnov distance
#

u <- runif(50, min=5, max=10)

fumle <- fitdist(u, "unif", method="mle")
summary(fumle)
plot(fumle)
gofstat(fumle)

fuCvM <- fitdist(u, "unif", method="mge", gof="CvM")
summary(fuCvM)
plot(fuCvM)
gofstat(fuCvM)

fuKS <- fitdist(u, "unif", method="mge", gof="KS")
summary(fuKS)
plot(fuKS)
gofstat(fuKS)

# (14) scaling problem
# the simulated dataset (below) has particularly small values, hence without scaling (10^0),
# the optimization raises an error. The for loop shows how scaling by 10^i
# for i=1,...,6 makes the fitting procedure work correctly.

x2 <- rnorm(100, 1e-4, 2e-4)

```

```

for(i in 0:6)
  cat(i, try(fitdist(x2*10^i, "cauchy", method="mle")$estimate, silent=TRUE), "\n")

# (15) Fit of a normal distribution on acute toxicity values of endosulfan in log10 for
# nonarthropod invertebrates, using maximum likelihood estimation
# to estimate what is called a species sensitivity distribution
# (SSD) in ecotoxicology, followed by estimation of the 5 percent quantile value of
# the fitted distribution (which is called the 5 percent hazardous concentration, HC5,
# in ecotoxicology) and estimation of other quantiles.
#
data(endosulfan)
ATV <- subset(endosulfan, group == "NonArthroInvert")$ATV
log10ATV <- log10(subset(endosulfan, group == "NonArthroInvert")$ATV)
fln <- fitdist(log10ATV, "norm")

quantile(fln, probs = 0.05)
quantile(fln, probs = c(0.05, 0.1, 0.2))

# (16) Fit of a triangular distribution using Cramer-von Mises or
# Kolmogorov-Smirnov distance
#

require("mc2d")
t <- rtriang(100, min=5, mode=6, max=10)
fCvM <- fitdist(t, "triang", method="mge", start = list(min=4, mode=6,max=9), gof="CvM")
fKS <- fitdist(t, "triang", method="mge", start = list(min=4, mode=6,max=9), gof="KS")
cdfcomp(list(fCvM,fKS))

# (17) fit a non classical discrete distribution (the zero inflated Poisson distribution)
#

require("gamlss.dist")
# depending of the random sample, the fit may fail
# it is why the seed was redefined here
# for you to get a sample on which it works
set.seed(1234)
x <- rZIP(n = 30, mu = 5, sigma = 0.2)
plotdist(x, discrete = TRUE)

fitzip <- fitdist(x, "ZIP", start = list(mu = 4, sigma = 0.15), discrete = TRUE,
optim.method = "L-BFGS-B", lower = c(0, 0), upper = c(Inf, 1))
summary(fitzip)
plot(fitzip)
fitp <- fitdist(x, "pois")
cdfcomp(list(fitzip, fitp))
gofstat(list(fitzip, fitp))

```

```

# (18) examples with distributions in actuar (predefined starting values)
#

require("actuar")
x <- c(2.3,0.1,2.7,2.2,0.4,2.6,0.2,1.,7.3,3.2,0.8,1.2,33.7,14.,
      21.4,7.7,1.,1.9,0.7,12.6,3.2,7.3,4.9,4000.,2.5,6.7,3.,63.,
      6.,1.6,10.1,1.2,1.5,1.2,30.,3.2,3.5,1.2,0.2,1.9,0.7,17.,
      2.8,4.8,1.3,3.7,0.2,1.8,2.6,5.9,2.6,6.3,1.4,0.8)
#log logistic
ft_lllogis <- fitdist(x,'llogis')

x <- c(0.3837053, 0.8576858, 0.3552237, 0.6226119, 0.4783756, 0.3139799, 0.4051403,
      0.4537631, 0.4711057, 0.5647414, 0.6479617, 0.7134207, 0.5259464, 0.5949068,
      0.3509200, 0.3783077, 0.5226465, 1.0241043, 0.4384580, 1.3341520)
#inverse weibull
ft_iw <- fitdist(x,'invweibull')

```

fitdistcens

Fitting of univariate distributions to censored data

Description

Fits a univariate distribution to censored data by maximum likelihood.

Usage

```

fitdistcens(censdata, distr, start=NULL, fix.arg=NULL,
  keepdata = TRUE, keepdata.nb=100, calcvcov=TRUE, ...)

```

```

## S3 method for class 'fitdistcens'
print(x, ...)

```

```

## S3 method for class 'fitdistcens'
plot(x, ...)

```

```

## S3 method for class 'fitdistcens'
summary(object, ...)

```

```

## S3 method for class 'fitdistcens'
logLik(object, ...)

```

```

## S3 method for class 'fitdistcens'
AIC(object, ..., k = 2)

```

```

## S3 method for class 'fitdistcens'

```

```

BIC(object, ...)

## S3 method for class 'fitdistcens'
vcov(object, ...)

## S3 method for class 'fitdistcens'
coef(object, ...)

```

Arguments

<code>censdata</code>	A dataframe of two columns respectively named <code>left</code> and <code>right</code> , describing each observed value as an interval. The left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The right column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations.
<code>distr</code>	A character string "name" naming a distribution, for which the corresponding density function <code>dname</code> and the corresponding distribution function <code>pname</code> must be defined, or directly the density function.
<code>start</code>	A named list giving the initial values of parameters of the named distribution. This argument may be omitted for some distributions for which reasonable starting values are computed (see the 'details' section of <code>mledist</code>).
<code>fix.arg</code>	An optional named list giving the values of parameters of the named distribution that must be kept fixed rather than estimated by maximum likelihood.
<code>x</code>	an object of class "fitdistcens".
<code>object</code>	an object of class "fitdistcens".
<code>keepdata</code>	a logical. If TRUE, dataset is returned, otherwise only a sample subset is returned.
<code>keepdata.nb</code>	When <code>keepdata=FALSE</code> , the length of the subset returned.
<code>calvcov</code>	A logical indicating if (asymptotic) covariance matrix is required.
<code>k</code>	penalty per parameter to be passed to the AIC generic function (2 by default).
<code>...</code>	further arguments to be passed to generic functions, to the function <code>plotdistcens</code> in order to control the type of ecdf-plot used for censored data, or to the function <code>mledist</code> in order to control the optimization method.

Details

Maximum likelihood estimations of the distribution parameters are computed using the function `mledist`. By default direct optimization of the log-likelihood is performed using `optim`, with the "Nelder-Mead" method for distributions characterized by more than one parameter and the "BFGS" method for distributions characterized by only one parameter. The algorithm used in `optim` can be chosen or another optimization function can be specified using `...` argument (see `mledist` for details). `start` may be omitted (i.e. NULL) for some classic distributions (see the 'details' section

of `mledist`). Note that when errors are raised by `optim`, it's a good idea to start by adding traces during the optimization process by adding `control=list(trace=1, REPORT=1)` in `...` argument.

The function is not able to fit a uniform distribution. With the parameter estimates, the function returns the log-likelihood and the standard errors of the estimates calculated from the Hessian at the solution found by `optim` or by the user-supplied function passed to `mledist`.

By default (`keepdata = TRUE`), the object returned by `fitdist` contains the data vector given in input. When dealing with large datasets, we can remove the original dataset from the output by setting `keepdata = FALSE`. In such a case, only `keepdata.nb` points (at most) are kept by random subsampling `keepdata.nb-4` points from the dataset and adding the component-wise minimum and maximum. If combined with `bootdistcens`, be aware that bootstrap is performed on the subset randomly selected in `fitdistcens`. Currently, the graphical comparisons of multiple fits is not available in this framework.

Weighted version of the estimation process is available for `method = "mle"` by using `weights=...`. See the corresponding man page for details. It is not yet possible to take into account weights in functions `plotdistcens`, `plot.fitdistcens` and `cdfcompdens` (developments planned in the future).

Once the parameter(s) is(are) estimated, `gofstat` allows to compute goodness-of-fit statistics.

Value

`fitdistcens` returns an object of class `"fitdistcens"`, a list with the following components:

<code>estimate</code>	the parameter estimates.
<code>method</code>	the character string coding for the fitting method : only <code>"mle"</code> for 'maximum likelihood estimation'.
<code>sd</code>	the estimated standard errors.
<code>cor</code>	the estimated correlation matrix, NA if numerically not computable or NULL if not available.
<code>vcov</code>	the estimated variance-covariance matrix, NULL if not available.
<code>loglik</code>	the log-likelihood.
<code>aic</code>	the Akaike information criterion.
<code>bic</code>	the the so-called BIC or SBC (Schwarz Bayesian criterion).
<code>censdata</code>	the censored data set.
<code>distname</code>	the name of the distribution.
<code>fix.arg</code>	the named list giving the values of parameters of the named distribution that must be kept fixed rather than estimated by maximum likelihood or NULL if there are no such parameters.
<code>fix.arg.fun</code>	the function used to set the value of <code>fix.arg</code> or NULL.
<code>dots</code>	the list of further arguments passed in <code>...</code> to be used in <code>bootdistcens</code> to control the optimization method used in iterative calls to <code>mledist</code> or NULL if no such arguments.
<code>convergence</code>	an integer code for the convergence of <code>optim/constrOptim</code> defined as below or defined by the user in the user-supplied optimization function. 0 indicates successful convergence. 1 indicates that the iteration limit of <code>optim</code> has been reached. 10 indicates degeneracy of the Nealder-Mead simplex. 100 indicates that <code>optim</code> encountered an internal error.

discrete always FALSE.
weights the vector of weights used in the estimation process or NULL.

Generic functions:

print The print of a "fitdist" object shows few traces about the fitting method and the fitted distribution.

summary The summary provides the parameter estimates of the fitted distribution, the log-likelihood, AIC and BIC statistics, the standard errors of the parameter estimates and the correlation matrix between parameter estimates.

plot The plot of an object of class "fitdistcens" returned by fitdistcens uses the function [plotdistcens](#).

logLik Extracts the estimated log-likelihood from the "fitdistcens" object.

AIC Extracts the AIC from the "fitdistcens" object.

BIC Extracts the BIC from the "fitdistcens" object.

vcov Extracts the estimated var-covariance matrix from the "fitdistcens" object (only available When method = "mle").

coef Extracts the fitted coefficients from the "fitdistcens" object.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

Venables WN and Ripley BD (2002), *Modern applied statistics with S*. Springer, New York, pp. 435-446, doi:[10.1007/9780387217062](https://doi.org/10.1007/9780387217062).

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:[10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [Surv2fitdistcens](#) to convert Surv outputs to a data frame appropriate for fitdistcens. See [plotdistcens](#), [optim](#) and [quantile.fitdistcens](#) for generic functions. See [gofstat](#) for goodness-of-fit statistics. See [fitdistrplus](#) for an overview of the package.

Please visit the [Frequently Asked Questions](#).

Examples

```
# (1) Fit of a lognormal distribution to bacterial contamination data
#
data(smokedfish)
fitsf <- fitdistcens(smokedfish,"lnorm")
summary(fitsf)
# default plot using the Wang technique (see ?plotdiscens for details)
plot(fitsf)
# plot using the Turnbull algorithm (see ?plotdiscens for details)
# with confidence intervals for the empirical distribution
```

```

plot(fitsf, NPMLE = TRUE, NPMLE.method = "Turnbull", Turnbull.confint = TRUE)
# basic plot using intervals and points (see ?plotdiscens for details)
plot(fitsf, NPMLE = FALSE)
# plot of the same fit using the Turnbull algorithm in logscale
cdfcompdens(fitsf,main="bacterial contamination fits",
  xlab="bacterial concentration (CFU/g)",ylab="F",
  addlegend = FALSE,lines01 = TRUE, xlogscale = TRUE, xlim = c(1e-2,1e2))
# zoom on large values of F
cdfcompdens(fitsf,main="bacterial contamination fits",
  xlab="bacterial concentration (CFU/g)",ylab="F",
  addlegend = FALSE,lines01 = TRUE, xlogscale = TRUE,
  xlim = c(1e-2,1e2),ylim=c(0.4,1))

# (2) Fit of a normal distribution on acute toxicity values
# of fluazinam (in decimal logarithm) for
# macroinvertebrates and zooplankton, using maximum likelihood estimation
# to estimate what is called a species sensitivity distribution
# (SSD) in ecotoxicology
#
data(fluazinam)
log10EC50 <-log10(fluazinam)
fln <- fitdistcens(log10EC50,"norm")
fln
summary(fln)
plot(fln)

# (3) defining your own distribution functions, here for the Gumbel distribution
# for other distributions, see the CRAN task view dedicated to
# probability distributions
#
dgumbel <- function(x,a,b) 1/b*exp((a-x)/b)*exp(-exp((a-x)/b))
pgumbel <- function(q,a,b) exp(-exp((a-q)/b))
qgumbel <- function(p,a,b) a-b*log(-log(p))
fg <- fitdistcens(log10EC50,"gumbel",start=list(a=1,b=1))
summary(fg)
plot(fg)

# (4) comparison of fits of various distributions
#
fll <- fitdistcens(log10EC50,"logis")
summary(fll)

cdfcompdens(list(fln,fll,fg),legendtext=c("normal","logistic","gumbel"),
  xlab = "log10(EC50)")

# (5) how to change the optimisation method?
#
fitdistcens(log10EC50,"logis",optim.method="Nelder-Mead")
fitdistcens(log10EC50,"logis",optim.method="BFGS")

```

```

fitdistcens(log10EC50,"logis",optim.method="SANN")

# (6) custom optimisation function - example with the genetic algorithm
#

#wrap genoud function rgenoud package
mygenoud <- function(fn, par, ...)
{
  require("rgenoud")
  res <- genoud(fn, starting.values=par, ...)
  standardres <- c(res, convergence=0)

  return(standardres)
}

# call fitdistcens with a 'custom' optimization function
fit.with.genoud <- fitdistcens(log10EC50,"logis", custom.optim=mygenoud, nvars=2,
  Domains=cbind(c(0,0), c(5, 5)), boundary.enforcement=1,
  print.level=1, hessian=TRUE)

summary(fit.with.genoud)

# (7) estimation of the mean of a normal distribution
# by maximum likelihood with the standard deviation fixed at 1 using the argument fix.arg
#
flnb <- fitdistcens(log10EC50, "norm", start = list(mean = 1),fix.arg = list(sd = 1))

# (8) Fit of a lognormal distribution on acute toxicity values of fluazinam for
# macroinvertebrates and zooplankton, using maximum likelihood estimation
# to estimate what is called a species sensitivity distribution
# (SSD) in ecotoxicology, followed by estimation of the 5 percent quantile value of
# the fitted distribution (which is called the 5 percent hazardous concentration, HC5,
# in ecotoxicology) and estimation of other quantiles.

data(fluazinam)
log10EC50 <-log10(fluazinam)
fln <- fitdistcens(log10EC50,"norm")

quantile(fln, probs = 0.05)
quantile(fln, probs = c(0.05, 0.1, 0.2))

# (9) Fit of a lognormal distribution on 72-hour acute salinity tolerance (LC50 values)
# of riverine macro-invertebrates using maximum likelihood estimation

data(salinity)
log10LC50 <-log10(salinity)
fln <- fitdistcens(log10LC50,"norm")
plot(fln)

```

fluazinam

Species-Sensitivity Distribution (SSD) for Fluazinam

Description

48-hour acute toxicity values (EC50 values) for exposure of macroinvertebrates and zooplankton to fluazinam.

Usage

```
data(fluazinam)
```

Format

fluazinam is a data frame with 2 columns named left and right, describing each observed EC50 value (in micrograms per liter) as an interval. The left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The right column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for noncensored observations.

Source

Hose, G.C., Van den Brink, P.J. 2004. The species sensitivity distribution approach compared to a microcosm study: A case study with the fungicide fluazinam. *Ecotoxicology and Environmental Safety*, **73**, 109-122.

Examples

```
# (1) load of data
#
data(fluazinam)

# (2) plot of data using Turnbull cdf plot
#
log10EC50 <- log10(fluazinam)
plotdistcens(log10EC50)

# (3) fit of a lognormal and a logistic distribution to data
# (classical distributions used for species sensitivity
# distributions, SSD, in ecotoxicology)
# and visual comparison of the fits using Turnbull cdf plot
#
f1n <- fitdistcens(log10EC50, "norm")
summary(f1n)

f1l <- fitdistcens(log10EC50, "logis")
summary(f1l)
```

```

cdfcompdens(list(fln,fll), legendtext = c("normal", "logistic"),
  xlab = "log10(EC50)")

# (4) estimation of the 5 percent quantile value of
# the normal fitted distribution (5 percent hazardous concentration : HC5)
# with its two-sided 95 percent confidence interval calculated by
# non parametric bootstrap
# with a small number of iterations to satisfy CRAN running times constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.
#
# in log10(EC50)
bln <- bootdistdens(fln, niter = 101)
HC5ln <- quantile(bln, probs = 0.05)
# in EC50
10^(HC5ln$quantiles)
10^(HC5ln$quantCI)

# (5) estimation of the HC5 value
# with its one-sided 95 percent confidence interval (type "greater")
#
# in log10(EC50)
HC5lnb <- quantile(bln, probs = 0.05, CI.type = "greater")

# in LC50
10^(HC5lnb$quantiles)
10^(HC5lnb$quantCI)

```

fremale

Fictive survival dataset of a french Male population

Description

100 male individuals randomly taken from `frefictivetable` in `CASdatasets` package

Usage

```
data(fremale)
```

Format

`fremale` is a data frame with 3 columns names `AgeIn`, `AgeOut` respectively for entry age and exit age; `Death` a binary dummy: 1 indicating the death of the individual; 0 a censored observation.

References

See full dataset `frefictivetable` of `CASdatasets` at <http://dutangc.perso.math.cnrs.fr/RRepository/>

Examples

```
# (1) load of data
#
data(fremale)
summary(fremale)
```

gofstat

Goodness-of-fit statistics

Description

Computes goodness-of-fit statistics for parametric distributions fitted to a same censored or non-censored data set.

Usage

```
gofstat(f, chisqbreaks, meancount, discrete, fitnames=NULL)

## S3 method for class 'gofstat.fitdist'
print(x, ...)
## S3 method for class 'gofstat.fitdistcens'
print(x, ...)
```

Arguments

f	An object of class "fitdist" (or "fitdistcens"), output of the function fitdist() (resp. "fitdistcens()"), or a list of "fitdist" objects, or a list of "fitdistcens" objects.
chisqbreaks	Only usable for non censored data, a numeric vector defining the breaks of the cells used to compute the chi-squared statistic. If omitted, these breaks are automatically computed from the data in order to reach roughly the same number of observations per cell, roughly equal to the argument meancount, or slightly more if there are some ties.
meancount	Only usable for non censored data, the mean number of observations per cell expected for the definition of the breaks of the cells used to compute the chi-squared statistic. This argument will not be taken into account if the breaks are directly defined in the argument chisqbreaks. If chisqbreaks and meancount are both omitted, meancount is fixed in order to obtain roughly $(4n)^{2/5}$ cells with n the length of the dataset.
discrete	If TRUE, only the Chi-squared statistic and information criteria are computed. If missing, discrete is passed from the first object of class "fitdist" of the list f. For censored data this argument is ignored, as censored data are considered continuous.
fitnames	A vector defining the names of the fits.
x	An object of class "gofstat.fitdist" or "gofstat.fitdistcens".
...	Further arguments to be passed to generic functions.

Details

For any type of data (censored or not), information criteria are calculated. For non censored data, added Goodness-of-fit statistics are computed as described below.

The Chi-squared statistic is computed using cells defined by the argument `chisqbreaks` or cells automatically defined from data, in order to reach roughly the same number of observations per cell, roughly equal to the argument `meancount`, or slightly more if there are some ties. The choice to define cells from the empirical distribution (data), and not from the theoretical distribution, was done to enable the comparison of Chi-squared values obtained with different distributions fitted on a same data set. If `chisqbreaks` and `meancount` are both omitted, `meancount` is fixed in order to obtain roughly $(4n)^{2/5}$ cells, with n the length of the data set (Vose, 2000). The Chi-squared statistic is not computed if the program fails to define enough cells due to a too small dataset. When the Chi-squared statistic is computed, and if the degree of freedom (nb of cells - nb of parameters - 1) of the corresponding distribution is strictly positive, the p-value of the Chi-squared test is returned.

For continuous distributions, Kolmogorov-Smirnov, Cramer-von Mises and Anderson-Darling and statistics are also computed, as defined by Stephens (1986).

An approximate Kolmogorov-Smirnov test is performed by assuming the distribution parameters known. The critical value defined by Stephens (1986) for a completely specified distribution is used to reject or not the distribution at the significance level 0.05. Because of this approximation, the result of the test (decision of rejection of the distribution or not) is returned only for data sets with more than 30 observations. Note that this approximate test may be too conservative.

For data sets with more than 5 observations and for distributions for which the test is described by Stephens (1986) for maximum likelihood estimations ("`exp`", "`cauchy`", "`gamma`" and "`weibull`"), the Cramer-von Mises and Anderson-darling tests are performed as described by Stephens (1986). Those tests take into account the fact that the parameters are not known but estimated from the data by maximum likelihood. The result is the decision to reject or not the distribution at the significance level 0.05. Those tests are available only for maximum likelihood estimations.

Only recommended statistics are automatically printed, i.e. Cramer-von Mises, Anderson-Darling and Kolmogorov statistics for continuous distributions and Chi-squared statistics for discrete ones ("`binom`", "`nbinom`", "`geom`", "`hyper`" and "`pois`").

Results of the tests are not printed but stored in the output of the function.

Value

`gofstat()` returns an object of class "`gofstat.fitdist`" or "`gofstat.fitdistcens`" with following components or a sublist of them (only `aic`, `bic` and `nbfit` for censored data) ,

<code>chisq</code>	a named vector with the Chi-squared statistics or NULL if not computed
<code>chisqbreaks</code>	common breaks used to define cells in the Chi-squared statistic
<code>chisqpvalue</code>	a named vector with the p-values of the Chi-squared statistic or NULL if not computed
<code>chisqdf</code>	a named vector with the degrees of freedom of the Chi-squared distribution or NULL if not computed
<code>chisqtable</code>	a table with observed and theoretical counts used for the Chi-squared calculations
<code>cvm</code>	a named vector of the Cramer-von Mises statistics or "not computed" if not computed

cvmtest	a named vector of the decisions of the Cramer-von Mises test or "not computed" if not computed
ad	a named vector with the Anderson-Darling statistics or "not computed" if not computed
adtest	a named vector with the decisions of the Anderson-Darling test or "not computed" if not computed
ks	a named vector with the Kolmogorov-Smirnov statistic or "not computed" if not computed
kstest	a named vector with the decisions of the Kolmogorov-Smirnov test or "not computed" if not computed
aic	a named vector with the values of the Akaike's Information Criterion.
bic	a named vector with the values of the Bayesian Information Criterion.
discrete	the input argument or the automatic definition by the function from the first object of class "fitdist" of the list in input.
nbfit	Number of fits in argument.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

- Cullen AC and Frey HC (1999), *Probabilistic techniques in exposure assessment*. Plenum Press, USA, pp. 81-155.
- Stephens MA (1986), *Tests based on edf statistics*. In Goodness-of-fit techniques (D'Agostino RB and Stephens MA, eds), Marcel Dekker, New York, pp. 97-194.
- Venables WN and Ripley BD (2002), *Modern applied statistics with S*. Springer, New York, pp. 435-446, doi:[10.1007/9780387217062](https://doi.org/10.1007/9780387217062).
- Vose D (2000), *Risk analysis, a quantitative guide*. John Wiley & Sons Ltd, Chichester, England, pp. 99-143.
- Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:[10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [fitdist](#).

Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) fit of two distributions to the serving size data
# by maximum likelihood estimation
# and comparison of goodness-of-fit statistics
#
```

```

data(groundbeef)
serving <- groundbeef$serving
(fitg <- fitdist(serving, "gamma"))
gofstat(fitg)
(fitln <- fitdist(serving, "lnorm"))
gofstat(fitln)

gofstat(list(fitg, fitln))

# (2) fit of two discrete distributions to toxocara data
# and comparison of goodness-of-fit statistics
#

data(toxocara)
number <- toxocara$number

fitp <- fitdist(number,"pois")
summary(fitp)
plot(fitp)

fitnb <- fitdist(number,"nbinom")
summary(fitnb)
plot(fitnb)

gofstat(list(fitp, fitnb),fitnames = c("Poisson","negbin"))

# (3) Get Chi-squared results in addition to
#     recommended statistics for continuous distributions
#

x4 <- rweibull(n=1000,shape=2,scale=1)
# fit of the good distribution
f4 <- fitdist(x4,"weibull")
plot(f4)

# fit of a bad distribution
f4b <- fitdist(x4,"cauchy")
plot(f4b)

(g <- gofstat(list(f4,f4b),fitnames=c("Weibull", "Cauchy")))
g$chisq
g$chisqdf
g$chisqpvalue
g$chisqtable

# and by defining the breaks
(g <- gofstat(list(f4,f4b),
chisqbreaks = seq(from = min(x4), to = max(x4), length.out = 10), fitnames=c("Weibull", "Cauchy")))
g$chisq
g$chisqdf
g$chisqpvalue

```

```

g$chisqtable

# (4) fit of two distributions on acute toxicity values
# of fluazinam (in decimal logarithm) for
# macroinvertebrates and zooplankton
# and comparison of goodness-of-fit statistics
#

data(fluazinam)
log10EC50 <-log10(fluazinam)
(fln <- fitdistcens(log10EC50,"norm"))
plot(fln)
gofstat(fln)
(fll <- fitdistcens(log10EC50,"logis"))
plot(fll)
gofstat(fll)

gofstat(list(fll, fln), fitnames = c("loglogistic", "lognormal"))

```

graphcomp

Graphical comparison of multiple fitted distributions (for non-censored data)

Description

cdfcomp plots the empirical cumulative distribution against fitted distribution functions, denscomp plots the histogram against fitted density functions, qqcomp plots theoretical quantiles against empirical ones, ppcomp plots theoretical probabilities against empirical ones. Only cdfcomp is able to plot fits of a discrete distribution.

Usage

```

cdfcomp(ft, xlim, ylim, xlogscale = FALSE, ylogscale = FALSE, main, xlab, ylab,
  datapch, datacol, fitlty, fitcol, fitlwd, addlegend = TRUE, legendtext,
  xlegend = "bottomright", ylegend = NULL, horizontals = TRUE, verticals = FALSE,
  do.points = TRUE, use.ppoints = TRUE, a.ppoints = 0.5, name.points = NULL,
  lines01 = FALSE, discrete, add = FALSE, plotstyle = "graphics",
  fitnbpts = 101, ...)

```

```

denscomp(ft, xlim, ylim, probability = TRUE, main, xlab, ylab, datacol, fitlty,
  fitcol, fitlwd, addlegend = TRUE, legendtext, xlegend = "topright", ylegend = NULL,
  demp = FALSE, dempcol = "black", plotstyle = "graphics",
  discrete, fitnbpts = 101, fittype="l", ...)

```

```

qqcomp(ft, xlim, ylim, xlogscale = FALSE, ylogscale = FALSE, main, xlab, ylab,
  fitpch, fitcol, fitlwd, addlegend = TRUE, legendtext, xlegend = "bottomright",
  ylegend = NULL, use.ppoints = TRUE, a.ppoints = 0.5, line01 = TRUE,

```

```

line01col = "black", line01lty = 1, ynoise = TRUE, plotstyle = "graphics", ...)

ppcomp(ft, xlim, ylim, xlogscale = FALSE, ylogscale = FALSE, main, xlab, ylab,
  fitpch, fitcol, fitlwd, addlegend = TRUE, legendtext, xlegend = "bottomright",
  ylegend = NULL, use.ppoints = TRUE, a.ppoints = 0.5, line01 = TRUE,
  line01col = "black", line01lty = 1, ynoise = TRUE, plotstyle = "graphics", ...)

```

Arguments

ft	One "fitdist" object or a list of objects of class "fitdist".
xlim	The x -limits of the plot.
ylim	The y -limits of the plot.
xlogscale	If TRUE, uses a logarithmic scale for the x -axis.
ylogscale	If TRUE, uses a logarithmic scale for the y -axis.
main	A main title for the plot. See also title .
xlab	A label for the x -axis, defaults to a description of x .
ylab	A label for the y -axis, defaults to a description of y .
datapch	An integer specifying a symbol to be used in plotting data points. See also par .
datacol	A specification of the color to be used in plotting data points. See also par .
fitcol	A (vector of) color(s) to plot fitted distributions. If there are fewer colors than fits they are recycled in the standard fashion. See also par .
fitlty	A (vector of) line type(s) to plot fitted distributions/densities. If there are fewer values than fits they are recycled in the standard fashion. See also par .
fitlwd	A (vector of) line size(s) to plot fitted distributions/densities. If there are fewer values than fits they are recycled in the standard fashion. See also par .
fitpch	A (vector of) line type(s) to plot fitted quantiles/probabilities. If there are fewer values than fits they are recycled in the standard fashion. See also par .
fittype	The type of plot for fitted probabilities in the case of discrete distributions: possible types are "p" for points, "l" for lines and "o" for both overplotted (as in plot.default). fittype is not used for non-discrete distributions.
fitnbpts	A numeric for the number of points to compute fitted probabilities or cumulative probabilities. Default to 101.
addlegend	If TRUE, a legend is added to the plot.
legendtext	A character or expression vector of length ≥ 1 to appear in the legend. See also legend .
xlegend, ylegend	The x and y coordinates to be used to position the legend. They can be specified by keyword. If plotstyle = "graphics", see xy.coords and legend . If plotstyle = "ggplot", the xlegend keyword must be one of top, bottom, left, or right. See also guide_legend in ggplot2
horizontal	If TRUE, draws horizontal lines for the step empirical cumulative distribution function (ecdf). See also plot.stepfun .

<code>verticals</code>	If TRUE, draws vertical lines for the empirical cumulative distribution function (ecdf). Only taken into account if <code>horizontal=TRUE</code> .
<code>do.points</code>	If TRUE (by default), draws points at the x-locations. For large dataset ($n > 1e4$), <code>do.points</code> is ignored and no point is drawn.
<code>use.ppoints</code>	If TRUE, probability points of the empirical distribution are defined using function <code>ppoints</code> as $(1:n - a.ppoints)/(n - 2a.ppoints + 1)$. If FALSE, probability points are simply defined as $(1:n)/n$. This argument is ignored for discrete data.
<code>a.ppoints</code>	If <code>use.ppoints=TRUE</code> , this is passed to the <code>ppoints</code> function.
<code>name.points</code>	Label vector for points if they are drawn i.e. if <code>do.points = TRUE</code> (only for non censored data).
<code>lines01</code>	A logical to plot two horizontal lines at $h=0$ and $h=1$ for <code>cdfcomp</code> .
<code>line01</code>	A logical to plot an horizontal line $y = x$ for <code>qqcomp</code> and <code>ppcomp</code> .
<code>line01col, line01lty</code>	Color and line type for <code>line01</code> . See also <code>par</code> .
<code>demp</code>	A logical to add the empirical density on the plot, using the <code>density</code> function.
<code>dempcol</code>	A color for the empirical density in case it is added on the plot (<code>demp=TRUE</code>).
<code>ynoise</code>	A logical to add a small noise when plotting empirical quantiles/probabilities for <code>qqcomp</code> and <code>ppcomp</code> .
<code>probability</code>	A logical to use the probability scale for <code>denscomp</code> . See also <code>hist</code> .
<code>discrete</code>	If TRUE, the distributions are considered discrete. When missing, <code>discrete</code> is set to TRUE if at least one object of the list <code>ft</code> is discrete.
<code>add</code>	If TRUE, adds to an already existing plot. If FALSE, starts a new plot. This parameter is not available when <code>plotstyle = "ggplot"</code> .
<code>plotstyle</code>	"graphics" or "ggplot". If "graphics", the display is built with <code>graphics</code> functions. If "ggplot", a graphic object output is created with <code>ggplot2</code> functions (the <code>ggplot2</code> package must be installed).
...	Further graphical arguments passed to graphical functions used in <code>cdfcomp</code> , <code>denscomp</code> , <code>ppcomp</code> and <code>qqcomp</code> when <code>plotstyle = "graphics"</code> . When <code>plotstyle = "ggplot"</code> , these arguments are only used by the histogram plot (<code>hist</code>) in the <code>denscomp</code> function. When <code>plotstyle = "ggplot"</code> , the graphical output can be customized with relevant <code>ggplot2</code> functions after you store your output.

Details

`cdfcomp` provides a plot of the empirical distribution and each fitted distribution in cdf, by default using the Hazen's rule for the empirical distribution, with probability points defined as $(1:n - 0.5)/n$. If `discrete` is TRUE, probability points are always defined as $(1:n)/n$. For large dataset ($n > 1e4$), no point is drawn but the line for ecdf is drawn instead. Note that when `horizontal`, `verticals` and `do.points` are FALSE, no empirical point is drawn, only the fitted cdf is shown.

`denscomp` provides a density plot of each fitted distribution with the histogram of the data for conyuous data. When `discrete=TRUE`, distributions are considered as discrete, no histogram is plotted but `demp` is forced to TRUE and fitted and empirical probabilities are plotted either with vertical lines `fitype="l"`, with single points `fitype="p"` or both lines and points `fitype="o"`.

ppcomp provides a plot of the probabilities of each fitted distribution (x -axis) against the empirical probabilities (y -axis) by default defined as $(1:n - 0.5)/n$ (data are assumed continuous). For large dataset ($n > 1e4$), lines are drawn instead of points and customized with the `fitpch` parameter.

qqcomp provides a plot of the quantiles of each theoretical distribution (x -axis) against the empirical quantiles of the data (y -axis), by default defining probability points as $(1:n - 0.5)/n$ for theoretical quantile calculation (data are assumed continuous). For large dataset ($n > 1e4$), lines are drawn instead of points and customized with the `fitpch` parameter.

By default a legend is added to these plots. Many graphical arguments are optional, dedicated to personalize the plots, and fixed to default values if omitted.

Value

*comp returns a list of drawn points and/or lines when `plotstyle == "graphics"` or an object of class `"ggplot"` when `plotstyle == "ggplot"`.

Author(s)

Christophe Dutang, Marie-Laure Delignette-Muller and Aurelie Siberchicot.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:[10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [plot](#), [legend](#), [ppoints](#), [plot.stepfun](#) for classic plotting functions. See [CIcdfplot](#) and [plotdist](#) for other plot functions of `fitdistrplus`.

Please visit the [Frequently Asked Questions](#).

Examples

```
# (1) Plot various distributions fitted to serving size data
#
data(groundbeef)
serving <- groundbeef$serving
fitW <- fitdist(serving, "weibull")
fitln <- fitdist(serving, "lnorm")
fitg <- fitdist(serving, "gamma")

cdfcomp(list(fitW, fitln, fitg), horizontal = FALSE)
cdfcomp(list(fitW, fitln, fitg), horizontal = TRUE)
cdfcomp(list(fitW, fitln, fitg), horizontal = TRUE, vertical = TRUE, datacol = "purple")
cdfcomp(list(fitW, fitln, fitg), legendtext = c("Weibull", "lognormal", "gamma"),
  main = "ground beef fits", xlab = "serving sizes (g)",
  ylab = "F", xlim = c(0, 250), xlegend = "center", lines01 = TRUE)
denscomp(list(fitW, fitln, fitg), legendtext = c("Weibull", "lognormal", "gamma"),
  main = "ground beef fits", xlab = "serving sizes (g)", xlim = c(0, 250), xlegend = "topright")
ppcomp(list(fitW, fitln, fitg), legendtext = c("Weibull", "lognormal", "gamma"),
  main = "ground beef fits", xlegend = "bottomright", line01 = TRUE)
```

```

qqcomp(list(fitW, fitln, fitg), legendtext = c("Weibull", "lognormal", "gamma"),
  main = "ground beef fits", xlegend = "bottomright", line01 = TRUE,
  xlim = c(0, 300), ylim = c(0, 300), fitpch = 16)

# (2) Plot lognormal distributions fitted by
# maximum goodness-of-fit estimation
# using various distances (data plotted in log scale)
#
data(endosulfan)
ATV <- subset(endosulfan, group == "NonArthroInvert")$ATV
taxaATV <- subset(endosulfan, group == "NonArthroInvert")$taxa
flnMGEKS <- fitdist(ATV, "lnorm", method = "mge", gof = "KS")
flnMGEAD <- fitdist(ATV, "lnorm", method = "mge", gof = "AD")
flnMGEADL <- fitdist(ATV, "lnorm", method = "mge", gof = "ADL")
flnMGEAD2L <- fitdist(ATV, "lnorm", method = "mge", gof = "AD2L")

cdfcomp(list(flnMGEKS, flnMGEAD, flnMGEADL, flnMGEAD2L),
  xlogscale = TRUE, main = "fits of a lognormal dist. using various GOF dist.",
  legendtext = c("MGE KS", "MGE AD", "MGE ADL", "MGE AD2L"),
  verticals = TRUE, xlim = c(1, 100000), name.points=taxaATV)
qqcomp(list(flnMGEKS, flnMGEAD, flnMGEADL, flnMGEAD2L),
  main = "fits of a lognormal dist. using various GOF dist.",
  legendtext = c("MGE KS", "MGE AD", "MGE ADL", "MGE AD2L"),
  xlogscale = TRUE, ylogscale = TRUE)
ppcomp(list(flnMGEKS, flnMGEAD, flnMGEADL, flnMGEAD2L),
  main = "fits of a lognormal dist. using various GOF dist.",
  legendtext = c("MGE KS", "MGE AD", "MGE ADL", "MGE AD2L"))

# (3) Plot normal and logistic distributions fitted by
# maximum likelihood estimation
# using various plotting positions in cdf plots
#
data(endosulfan)
log10ATV <- log10(subset(endosulfan, group == "NonArthroInvert")$ATV)
fln <- fitdist(log10ATV, "norm")
fll <- fitdist(log10ATV, "logis")

# default plot using Hazen plotting position: (1:n - 0.5)/n
cdfcomp(list(fln, fll), legendtext = c("normal", "logistic"), xlab = "log10ATV")

# plot using mean plotting position (named also Gumbel plotting position)
# (1:n)/(n + 1)
cdfcomp(list(fln, fll), legendtext = c("normal", "logistic"), xlab = "log10ATV",
  use.ppoints = TRUE, a.ppoints = 0)

# plot using basic plotting position: (1:n)/n
cdfcomp(list(fln, fll), legendtext = c("normal", "logistic"), xlab = "log10ATV",
  use.ppoints = FALSE)

# (4) Comparison of fits of two distributions fitted to discrete data

```

```

#
data(toxocara)
number <- toxocara$number
fitp <- fitdist(number, "pois")
fitnb <- fitdist(number, "nbinom")
cdfcomp(list(fitp, fitnb), legendtext = c("Poisson", "negative binomial"))
denscomp(list(fitp, fitnb),demp = TRUE, legendtext = c("Poisson", "negative binomial"))
denscomp(list(fitp, fitnb),demp = TRUE, fittype = "l", dempcol = "black",
  legendtext = c("Poisson", "negative binomial"))
denscomp(list(fitp, fitnb),demp = TRUE, fittype = "p", dempcol = "black",
  legendtext = c("Poisson", "negative binomial"))
denscomp(list(fitp, fitnb),demp = TRUE, fittype = "o", dempcol = "black",
  legendtext = c("Poisson", "negative binomial"))

# (5) Customizing of graphical output and use of ggplot2
#
data(groundbeef)
serving <- groundbeef$serving
fitW <- fitdist(serving, "weibull")
fitln <- fitdist(serving, "lnorm")
fitg <- fitdist(serving, "gamma")
if (requireNamespace ("ggplot2", quietly = TRUE)) {
  denscomp(list(fitW, fitln, fitg), plotstyle = "ggplot")
  cdfcomp(list(fitW, fitln, fitg), plotstyle = "ggplot")
  qqcomp(list(fitW, fitln, fitg), plotstyle = "ggplot")
  ppcomp(list(fitW, fitln, fitg), plotstyle = "ggplot")
}

# customizing graphical output with graphics
denscomp(list(fitW, fitln, fitg), legendtext = c("Weibull", "lognormal", "gamma"),
  main = "ground beef fits", xlab = "serving sizes (g)", xlim = c(0, 250),
  xlegend = "topright", addlegend = FALSE)

# customizing graphical output with ggplot2
if (requireNamespace ("ggplot2", quietly = TRUE)) {
  dcomp <- denscomp(list(fitW, fitln, fitg), legendtext = c("Weibull", "lognormal", "gamma"),
    xlab = "serving sizes (g)", xlim = c(0, 250),
    xlegend = "topright", plotstyle = "ggplot", breaks = 20, addlegend = FALSE)
  dcomp + ggplot2::theme_minimal() + ggplot2::ggtitle("Ground beef fits")
}

```

graphcompdens

Graphical comparison of multiple fitted distributions for censored data

Description

`cdfcompdens` plots the empirical cumulative distribution against fitted distribution functions, `qqcompdens`

plots theoretical quantiles against empirical ones, `ppcompens` plots theoretical probabilities against empirical ones.

Usage

```

cdfcompens(ft, xlim, ylim, xlogscale = FALSE, ylogscale = FALSE, main, xlab, ylab,
  datacol, fillrect, fitlty, fitcol, fitlwd, addlegend = TRUE, legendtext,
  xlegend = "bottomright", ylegend = NULL, lines01 = FALSE,
  Turnbull.confint = FALSE,
  NPML.method = "Wang",
  add = FALSE, plotstyle = "graphics", ...)
qqcompens(ft, xlim, ylim, xlogscale = FALSE, ylogscale = FALSE, main, xlab, ylab,
  fillrect, fitcol, fitlwd, addlegend = TRUE, legendtext, xlegend = "bottomright",
  ylegend = NULL, line01 = TRUE, line01col = "black", line01lty = 1,
  ynoise = TRUE, NPML.method = "Wang", plotstyle = "graphics", ...)
ppcompens(ft, xlim, ylim, xlogscale = FALSE, ylogscale = FALSE, main, xlab, ylab,
  fillrect, fitcol, fitlwd, addlegend = TRUE, legendtext, xlegend = "bottomright",
  ylegend = NULL, line01 = TRUE, line01col = "black", line01lty = 1,
  ynoise = TRUE, NPML.method = "Wang", plotstyle = "graphics", ...)

```

Arguments

<code>ft</code>	One "fitdistcens" object or a list of objects of class "fitdistcens".
<code>xlim</code>	The x -limits of the plot.
<code>ylim</code>	The y -limits of the plot.
<code>xlogscale</code>	If TRUE, uses a logarithmic scale for the x -axis.
<code>ylogscale</code>	If TRUE, uses a logarithmic scale for the y -axis.
<code>main</code>	A main title for the plot, see also title .
<code>xlab</code>	A label for the x -axis, defaults to a description of x .
<code>ylab</code>	A label for the y -axis, defaults to a description of y .
<code>datacol</code>	A specification of the color to be used in plotting data points.
<code>fillrect</code>	A specification of the color to be used for filling rectangles of non uniqueness of the empirical cumulative distribution (only used if <code>NPML.method</code> is equal to "Wang" in <code>cdfcompens</code>). Fix it to NA if you do not want to fill the rectangles.
<code>fitcol</code>	A (vector of) color(s) to plot fitted distributions. If there are fewer colors than fits they are recycled in the standard fashion.
<code>fitlty</code>	A (vector of) line type(s) to plot fitted distributions. If there are fewer values than fits they are recycled in the standard fashion. See also par .
<code>fitlwd</code>	A (vector of) line size(s) to plot fitted distributions. If there are fewer values than fits they are recycled in the standard fashion. See also par .
<code>addlegend</code>	If TRUE, a legend is added to the plot.
<code>legendtext</code>	A character or expression vector of length ≥ 1 to appear in the legend, see also legend .

xlegend, ylegend	The x and y coordinates to be used to position the legend. They can be specified by keyword. If <code>plotstyle = "graphics"</code> , see xy.coords and legend . If <code>plotstyle = "ggplot"</code> , the <code>xlegend</code> keyword must be one of <code>top</code> , <code>bottom</code> , <code>left</code> , or <code>right</code> . See also <code>guide_legend</code> in <code>ggplot2</code>
lines01	A logical to plot two horizontal lines at $h=0$ and $h=1$ for <code>cdfcompdens</code> .
Turnbull.confint	if TRUE confidence intervals will be added to the Turnbull plot. In that case <code>NPMLE.method</code> is forced to "Turnbull"
NPMLE.method	Three NPML techniques are provided, "Wang", the default one, rewritten from the package <code>npsurv</code> using function <code>constrOptim</code> from the package <code>stats</code> for optimisation, "Turnbull.middlepoints", an older one which is implemented in the package <code>survival</code> and "Turnbull.intervals" that uses the same Turnbull algorithm from the package <code>survival</code> but associates an interval to each equivalence class instead of the midpoint of this interval (see details). Only "Wang" and "Turnbull.intervals" enable the derivation of a Q-Q plot and a P-P plot.
add	If TRUE, adds to an already existing plot. If FALSE, starts a new plot. This parameter is not available when <code>plotstyle = "ggplot"</code> .
line01	A logical to plot an horizontal line $y = x$ for <code>qqcompdens</code> and <code>ppcompdens</code> .
line01col, line01lty	Color and line type for <code>line01</code> . See also par .
ynoise	A logical to add a small noise when plotting empirical quantiles/probabilities for <code>qqcompdens</code> and <code>ppcompdens</code> . <code>ynoise</code> is only used when various fits are plotted with the "graphics" <code>plotstyle</code> . Facets are used instead with the "ggplot" <code>plotstyle</code> .
plotstyle	"graphics" or "ggplot". If "graphics", the display is built with graphics functions. If "ggplot", a graphic object output is created with <code>ggplot2</code> functions (the <code>ggplot2</code> package must be installed). In "cdfcompdens", "ggplot" graphics are only available with "Wang" NPML technique.
...	Further graphical arguments passed to graphical functions used in <code>cdfcompdens</code> , <code>ppcompdens</code> and <code>qqcompdens</code> .

Details

See details of [plotdistdens](#) for a detailed description of provided goodness-of-fit plots.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

- Turnbull BW (1974), *Nonparametric estimation of a survivorship function with doubly censored data*. Journal of American Statistical Association, 69, 169-173.
- Wang Y (2008), *Dimension-reduced nonparametric maximum likelihood computation for interval-censored data*. Computational Statistics & Data Analysis, 52, 2388-2402.

Wang Y and Taylor SM (2013), *Efficient computation of nonparametric survival functions via a hierarchical mixture formulation*. *Statistics and Computing*, 23, 713-725.

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. *Journal of Statistical Software*, 64(4), 1-34.

See Also

See [plotdistcens](#), [survfit.formula](#), [legend](#) and [par](#).

Please visit the [Frequently Asked Questions](#).

Examples

```
# (1) Plot various distributions fitted to bacterial contamination data
#
data(smokedfish)
Clog10 <- log10(smokedfish)

fitsfn <- fitdistcens(Clog10,"norm")
summary(fitsfn)

fitsfl <- fitdistcens(Clog10,"logis")
summary(fitsfl)

dgumbel <- function(x,a,b) 1/b*exp((a-x)/b)*exp(-exp((a-x)/b))
pgumbel <- function(q,a,b) exp(-exp((a-q)/b))
qgumbel <- function(p,a,b) a-b*log(-log(p))
fitsfg<-fitdistcens(Clog10,"gumbel",start=list(a=-3,b=3))
summary(fitsfg)

# CDF plot
cdfcompens(list(fitsfn,fitsfl,fitsfg))
cdfcompens(list(fitsfn,fitsfl,fitsfg),datacol="orange",fillrect = NA,
  legendtext=c("normal","logistic","Gumbel"),
  main="bacterial contamination fits",
  xlab="bacterial concentration (CFU/g)",ylab="F",
  xlegend = "bottom",lines01 = TRUE)
# alternative Turnbull plot for the empirical cumulative distribution
# (default plot of the previous versions of the package)
cdfcompens(list(fitsfn,fitsfl,fitsfg), NPMLmethod = "Turnbull.middlepoints")

# customizing graphical output with ggplot2
if (requireNamespace ("ggplot2", quietly = TRUE)) {
  cdfcompens <- cdfcompens(list(fitsfn,fitsfl,fitsfg),datacol="orange",fillrect = NA,
    legendtext=c("normal","logistic","Gumbel"),
    xlab="bacterial concentration (CFU/g)",ylab="F",
    xlegend = "bottom",lines01 = TRUE, plotstyle = "ggplot")
  cdfcompens + ggplot2::theme_minimal() + ggplot2::ggtitle("Bacterial contamination fits")
}

# PP plot
ppcompens(list(fitsfn,fitsfl,fitsfg))
ppcompens(list(fitsfn,fitsfl,fitsfg), ynoise = FALSE)
```

```

par(mfrow = c(2,2))
ppcompcons(fitsfn)
ppcompcons(fitsfl)
ppcompcons(fitsfg)
par(mfrow = c(1,1))
if (requireNamespace ("ggplot2", quietly = TRUE)) {
  ppcompcons(list(fitsfn,fitsfl,fitsfg), plotstyle = "ggplot")
  ppcompcons(list(fitsfn,fitsfl,fitsfg), plotstyle = "ggplot",
    fillrect = c("lightpink", "lightblue", "lightgreen"),
    fitcol = c("red", "blue", "green"))
}

# QQ plot
qqcompcons(list(fitsfn,fitsfl,fitsfg))
qqcompcons(list(fitsfn,fitsfl,fitsfg), ynoise = FALSE)
par(mfrow = c(2,2))
qqcompcons(fitsfn)
qqcompcons(fitsfl)
qqcompcons(fitsfg)
par(mfrow = c(1,1))

if (requireNamespace ("ggplot2", quietly = TRUE)) {
  qqcompcons(list(fitsfn,fitsfl,fitsfg), ynoise = FALSE, plotstyle = "ggplot")
  qqcompcons(list(fitsfn,fitsfl,fitsfg), ynoise = FALSE, plotstyle = "ggplot",
    fillrect = c("lightpink", "lightblue", "lightgreen"),
    fitcol = c("red", "blue", "green"))
}

```

groundbeef

Ground beef serving size data set

Description

Serving sizes collected in a French survey, for ground beef patties consumed by children under 5 years old.

Usage

```
data(groundbeef)
```

Format

groundbeef is a data frame with 1 column (serving: serving sizes in grams)

Source

Delignette-Muller, M.L., Cornu, M. 2008. Quantitative risk assessment for *Escherichia coli* O157:H7 in frozen ground beef patties consumed by young children in French households. *International Journal of Food Microbiology*, **128**, 158-164.

Examples

```

# (1) load of data
#
data(groundbeef)

# (2) description and plot of data
#
serving <- groundbeef$serving
descdist(serving)
plotdist(serving)

# (3) fit of a Weibull distribution to data
#
fitW <- fitdist(serving, "weibull")
summary(fitW)
plot(fitW)
gofstat(fitW)

```

logLikplot

(Log)likelihood plot for a fit using maximum likelihood

Description

llplot plots the (log)likelihood around the estimation for distributions fitted by maximum likelihood.

Usage

```

llplot(mlefit, loglik = TRUE, expansion = 1, lseq = 50,
       back.col = TRUE, nlev = 10, pal.col = terrain.colors(100),
       fit.show = FALSE, fit.pch = 4, ...)

```

Arguments

mlefit	An object of class "fitdist" of "fitdistcens" obtained by maximum likelihood (with method = "mle")
loglik	a logical to plot log-likelihood or likelihood function.
expansion	a expansion factor to enlarge the default range of values explored for each parameter.
lseq	length of sequences of parameters.
back.col	logical (for llsurface only). Contours are plotted with a background gradient of colors if TRUE.
nlev	number of contour levels to plot.
pal.col	Palette of colors. Colors to be used as back (for llsurface only).
fit.show	a logical to plot the mle estimate.
fit.pch	the type of point used to plot the mle estimate.
...	Further graphical arguments passed to graphical functions.

Details

llplot plots the (log)likelihood surface(s) (or curve if there is only one estimated parameter) around the maximum likelihood estimation. It internally calls function [llsurface](#) and [llcurve](#). When there is more than two estimated parameters, the (log)likelihood surface is plotted for each combination of two parameters, fixing the other ones to their estimated value. For each (log)likelihood surface, when `back.col` [image](#) (2D-plot) is used and when `nlev > 0` [contour](#) (2D-plot) is used to add `nlev` contours. By default the range of values explored for each estimated parameter is of 2 standard error around the mle estimate but this range can be expanded (or contracted) using the argument `expansion`.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:[10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [llsurface](#) and [llcurve](#) for manual (log)likelihood plots (surface or curve) and [plot](#), [contour](#), [image](#) for classic plotting functions.

Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) a distribution with one parameter
#

x <- rexp(50)
fite <- fitdist(x, "exp")
llplot(fite)
llplot(fite, col = "red", fit.show = TRUE)
llplot(fite, col = "red", fit.show = TRUE, loglik = FALSE)

# (2) a distribution with two parameters
#

data(groundbeef)
serving <- groundbeef$serving
fitg <- fitdist(serving, "gamma")
llplot(fitg)

llplot(fitg, expansion = 2)
llplot(fitg, pal.col = heat.colors(100), fit.show = TRUE)
llplot(fitg, back.col = FALSE, nlev = 25, fit.show = TRUE)
```

```

# (3) a distribution with two parameters with one fixed
#
fitg2 <- fitdist(serving, "gamma", fix.arg = list(rate = 0.5))
llplot(fitg2, fit.show = TRUE)

# (4) a distribution with three parameters
#

data(endosulfan)
ATV <- endosulfan$ATV
require("actuar")
fBurr <- fitdist(ATV, "burr", start = list(shape1 = 0.3, shape2 = 1, rate = 1))
llplot(fBurr)
llplot(fBurr, back.col = FALSE, fit.show = TRUE, fit.pch = 16)
llplot(fBurr, nlev = 0, pal.col = rainbow(100), lseq = 100)

# (5) a distribution with two parameters fitted on censored data
#
data(salinity)
fsal <- fitdistcens(salinity, "lnorm")
llplot(fsal, fit.show = TRUE)
llplot(fsal, fit.show = TRUE, loglik = FALSE)

```

logLiksurface

(Log)likelihood surfaces or (log)likelihood curves

Description

llsurface plots the likelihood surface for distributions with two or more parameters, llcurve plots the likelihood curve for distributions with one or more parameters.

Usage

```

llsurface(data, distr, plot.arg, min.arg, max.arg, lseq = 50, fix.arg = NULL,
  loglik = TRUE, back.col = TRUE, nlev = 10, pal.col = terrain.colors(100),
  weights = NULL, ...)

llcurve(data, distr, plot.arg, min.arg, max.arg, lseq = 50, fix.arg = NULL,
  loglik = TRUE, weights = NULL, ...)

```

Arguments

<code>data</code>	A numeric vector for non censored data or a dataframe of two columns respectively named <code>left</code> and <code>right</code> , describing each observed value as an interval for censored data. In that case the left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The right column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations.
<code>distr</code>	A character string "name" naming a distribution for which the corresponding density function <code>dname</code> and the corresponding distribution function <code>pname</code> must be classically defined.
<code>plot.arg</code>	a two-element vector with the names of the two parameters that will vary for <code>l1surface</code> , only one element for <code>l1curve</code> .
<code>min.arg</code>	a two-element vector with lower plotting bounds for <code>l1surface</code> , only one element for <code>l1curve</code> .
<code>max.arg</code>	a two-element vector with upper plotting bounds for <code>l1surface</code> , only one element for <code>l1curve</code> .
<code>lseq</code>	length of sequences of parameters.
<code>fix.arg</code>	a named list with fixed value of other parameters.
<code>loglik</code>	a logical to plot log-likelihood or likelihood function.
<code>back.col</code>	logical (for <code>l1surface</code> only). Contours are plotted with a background gradient of colors if TRUE.
<code>nlev</code>	number of contour levels to plot (for <code>l1surface</code> only).
<code>pal.col</code>	Palette of colors. Colors to be used as back (for <code>l1surface</code> only).
<code>weights</code>	an optional vector of weights to be used in the fitting process. Should be NULL or a numeric vector with strictly positive values (classically the number of occurrences of each observation).
<code>...</code>	Further graphical arguments passed to graphical functions.

Details

These two function are not intended to be called directly but is internally called in `llplot`.

`l1surface` plots the likelihood surface for distributions with two varying parameters and other parameters fixed. When `back.col`, `image` (2D-plot) is used. When `nlev > 0`, `contour` (2D-plot) is used to add `nlev` contours.

`l1curve` plots the likelihood curve for distributions with one varying parameter and other parameters fixed.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:10.18637/jss.v064.i04.

See Also

See [llplot](#) for an automatic (log)likelihood plots (surface ou curve) of an object of class "fitdistr" or "fitdistrib" and [plot](#), [contour](#), [image](#) for classic plotting functions.

Please visit the [Frequently Asked Questions](#).

Examples

```
# (1) loglikelihood or likelihood curve
#

n <- 100
set.seed(123) # here just to make random sampling reproducible
x <- rexp(n)

llcurve(data = x, distr = "exp", plot.arg = "rate", min.arg = 0, max.arg = 4)

llcurve(data = x, distr = "exp", plot.arg = "rate", min.arg = 0, max.arg = 4,
loglik = FALSE)

llcurve(data = x, distr = "exp", plot.arg = "rate", min.arg = 0, max.arg = 4,
  main = "log-likelihood for exponential distribution", col = "red")
abline(v = 1, lty = 2)

# (2) loglikelihood surface
#

x <- rnorm(n, 0, 1)

llsurface(data =x, distr="norm", plot.arg=c("mean", "sd"),
  min.arg=c(-1, 0.5), max.arg=c(1, 3/2), back.col = FALSE,
  main="log-likelihood for normal distribution")
llsurface(data =x, distr="norm", plot.arg=c("mean", "sd"),
  min.arg=c(-1, 0.5), max.arg=c(1, 3/2),
  main="log-likelihood for normal distribution",
  nlev = 20, pal.col = heat.colors(100),)
points(0, 1, pch="+", col="red")
llsurface(data =x, distr="norm", plot.arg=c("mean", "sd"),
  min.arg=c(-1, 0.5), max.arg=c(1, 3/2),
  main="log-likelihood for normal distribution",
  nlev = 0, back.col = TRUE, pal.col = rainbow(100, s = 0.5, end = 0.8))
points(0, 1, pch="+", col="black")
```

mgedist	<i>Maximum goodness-of-fit fit of univariate continuous distributions</i>
---------	---

Description

Fit of univariate continuous distribution by maximizing goodness-of-fit (or minimizing distance) for non censored data.

Usage

```
mgedist(data, distr, gof = "CvM", start = NULL, fix.arg = NULL, optim.method = "default",
        lower = -Inf, upper = Inf, custom.optim = NULL, silent = TRUE, gradient = NULL,
        checkstartfix=FALSE, calcvcov=FALSE, ...)
```

Arguments

data	A numeric vector for non censored data.
distr	A character string "name" naming a distribution for which the corresponding quantile function qname and the corresponding density distribution dname must be classically defined.
gof	A character string coding for the name of the goodness-of-fit distance used : "CvM" for Cramer-von Mises distance, "KS" for Kolmogorov-Smirnov distance, "AD" for Anderson-Darling distance, "ADR", "ADL", "AD2R", "AD2L" and "AD2" for variants of Anderson-Darling distance described by Luceno (2006).
start	A named list giving the initial values of parameters of the named distribution or a function of data computing initial values and returning a named list. This argument may be omitted (default) for some distributions for which reasonable starting values are computed (see the 'details' section of mledist).
fix.arg	An optional named list giving the values of fixed parameters of the named distribution or a function of data computing (fixed) parameter values and returning a named list. Parameters with fixed value are thus NOT estimated.
optim.method	"default" or optimization method to pass to optim .
lower	Left bounds on the parameters for the "L-BFGS-B" method (see optim).
upper	Right bounds on the parameters for the "L-BFGS-B" method (see optim).
custom.optim	a function carrying the optimization.
silent	A logical to remove or show warnings when bootstrapping.
gradient	A function to return the gradient of the gof distance for the "BFGS", "CG" and "L-BFGS-B" methods. If it is NULL, a finite-difference approximation will be used.
checkstartfix	A logical to test starting and fixed values. Do not change it.
calcvcov	A logical indicating if (asymptotic) covariance matrix is required. (currently ignored)
...	further arguments passed to the optim , constrOptim or <code>custom.optim</code> function.

Details

The `mgedist` function numerically maximizes goodness-of-fit, or minimizes a goodness-of-fit distance coded by the argument `gof`. One may use one of the classical distances defined in Stephens (1986), the Cramer-von Mises distance ("`CvM`"), the Kolmogorov-Smirnov distance ("`KS`") or the Anderson-Darling distance ("`AD`") which gives more weight to the tails of the distribution, or one of the variants of this last distance proposed by Luceno (2006). The right-tail AD ("`ADR`") gives more weight only to the right tail, the left-tail AD ("`ADL`") gives more weight only to the left tail. Either of the tails, or both of them, can receive even larger weights by using second order Anderson-Darling Statistics (using "`AD2R`", "`AD2L`" or "`AD2`").

The optimization process is the same as `mledist`, see the 'details' section of that function.

This function is not intended to be called directly but is internally called in `fitdist` and `bootdist`.

This function is intended to be used only with continuous distributions and weighted maximum goodness-of-fit estimation is not allowed.

NB: if your data values are particularly small or large, a scaling may be needed before the optimization process. See example (4).

Value

`mgedist` returns a list with following components,

<code>estimate</code>	the parameter estimates.
<code>convergence</code>	an integer code for the convergence of <code>optim</code> defined as below or defined by the user in the user-supplied optimization function. 0 indicates successful convergence. 1 indicates that the iteration limit of <code>optim</code> has been reached. 10 indicates degeneracy of the Nelder-Mead simplex. 100 indicates that <code>optim</code> encountered an internal error.
<code>value</code>	the minimal value reached for the criterion to minimize.
<code>hessian</code>	a symmetric matrix computed by <code>optim</code> as an estimate of the Hessian at the solution found or computed in the user-supplied optimization function.
<code>optim.function</code>	the name of the optimization function used for maximum likelihood.
<code>optim.method</code>	when <code>optim</code> is used, the name of the algorithm used, the field <code>method</code> of the <code>custom.optim</code> function otherwise.
<code>fix.arg</code>	the named list giving the values of parameters of the named distribution that must kept fixed rather than estimated by maximum likelihood or NULL if there are no such parameters.
<code>fix.arg.fun</code>	the function used to set the value of <code>fix.arg</code> or NULL.
<code>weights</code>	the vector of weights used in the estimation process or NULL.
<code>counts</code>	A two-element integer vector giving the number of calls to the log-likelihood function and its gradient respectively. This excludes those calls needed to compute the Hessian, if requested, and any calls to log-likelihood function to compute a finite-difference approximation to the gradient. <code>counts</code> is returned by <code>optim</code> or the user-supplied function or set to NULL.
<code>optim.message</code>	A character string giving any additional information returned by the optimizer, or NULL. To understand exactly the message, see the source code.
<code>loglik</code>	the log-likelihood value.
<code>gof</code>	the code of the goodness-of-fit distance maximized.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

- Luceno A (2006), *Fitting the generalized Pareto distribution to data using maximum goodness-of-fit estimators*. Computational Statistics and Data Analysis, 51, 904-917, [doi:10.1016/j.csda.2005.09.011](https://doi.org/10.1016/j.csda.2005.09.011).
- Stephens MA (1986), *Tests based on edf statistics*. In Goodness-of-fit techniques (D'Agostino RB and Stephens MA, eds), Marcel Dekker, New York, pp. 97-194.
- Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, [doi:10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [mmedist](#), [mledist](#), [qmedist](#), [fitdistr](#) for other estimation methods.
Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) Fit of a Weibull distribution to serving size data by maximum
# goodness-of-fit estimation using all the distances available
#

data(groundbeef)
serving <- groundbeef$serving
mgedist(serving, "weibull", gof="CvM")
mgedist(serving, "weibull", gof="KS")
mgedist(serving, "weibull", gof="AD")
mgedist(serving, "weibull", gof="ADR")
mgedist(serving, "weibull", gof="ADL")
mgedist(serving, "weibull", gof="AD2R")
mgedist(serving, "weibull", gof="AD2L")
mgedist(serving, "weibull", gof="AD2")

# (2) Fit of a uniform distribution using Cramer-von Mises or
# Kolmogorov-Smirnov distance
#

u <- runif(100,min=5,max=10)
mgedist(u,"unif",gof="CvM")
mgedist(u,"unif",gof="KS")

# (3) Fit of a triangular distribution using Cramer-von Mises or
# Kolmogorov-Smirnov distance
#
```

```

require("mc2d")
t <- rtriang(100,min=5,mode=6,max=10)
mgedist(t,"triang",start = list(min=4, mode=6,max=9),gof="CVM")
mgedist(t,"triang",start = list(min=4, mode=6,max=9),gof="KS")

# (4) scaling problem
# the simulated dataset (below) has particularly small values, hence without scaling (10^0),
# the optimization raises an error. The for loop shows how scaling by 10^i
# for i=1,...,6 makes the fitting procedure work correctly.

x2 <- rnorm(100, 1e-4, 2e-4)
for(i in 6:0)
  cat(i, try(mgedist(x*10^i,"cauchy")$estimate, silent=TRUE), "\n")

```

mledist

Maximum likelihood fit of univariate distributions

Description

Fit of univariate distributions using maximum likelihood for censored or non censored data.

Usage

```

mledist(data, distr, start = NULL, fix.arg = NULL, optim.method = "default",
  lower = -Inf, upper = Inf, custom.optim = NULL, weights = NULL, silent = TRUE,
  gradient = NULL, checkstartfix=FALSE, calcvcov=FALSE, ...)

```

Arguments

data	A numeric vector for non censored data or a dataframe of two columns respectively named <code>left</code> and <code>right</code> , describing each observed value as an interval for censored data. In that case the <code>left</code> column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The <code>right</code> column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations.
distr	A character string "name" naming a distribution for which the corresponding density function <code>dname</code> and the corresponding distribution function <code>pname</code> must be classically defined.
start	A named list giving the initial values of parameters of the named distribution or a function of data computing initial values and returning a named list. This argument may be omitted (default) for some distributions for which reasonable starting values are computed (see details).

<code>fix.arg</code>	An optional named list giving the values of fixed parameters of the named distribution or a function of data computing (fixed) parameter values and returning a named list. Parameters with fixed value are thus NOT estimated by this maximum likelihood procedure.
<code>optim.method</code>	"default" (see details) or an optimization method to pass to <code>optim</code> .
<code>lower</code>	Left bounds on the parameters for the "L-BFGS-B" method (see <code>optim</code>).
<code>upper</code>	Right bounds on the parameters for the "L-BFGS-B" method (see <code>optim</code>).
<code>custom.optim</code>	a function carrying the MLE optimisation (see details).
<code>weights</code>	an optional vector of weights to be used in the fitting process. Should be NULL or a numeric vector with strictly positive integers (typically the number of occurrences of each observation). If non-NULL, weighted MLE is used, otherwise ordinary MLE.
<code>silent</code>	A logical to remove or show warnings when bootstrapping.
<code>gradient</code>	A function to return the gradient of the log-likelihood for the "BFGS", "CG" and "L-BFGS-B" methods. If it is NULL, a finite-difference approximation will be used, see details.
<code>checkstartfix</code>	A logical to test starting and fixed values. Do not change it.
<code>calvcov</code>	A logical indicating if (asymptotic) covariance matrix is required.
<code>...</code>	further arguments passed to the <code>optim</code> , <code>constrOptim</code> or <code>custom.optim</code> function.

Details

This function is not intended to be called directly but is internally called in `fitdist` and `bootdist` when used with the maximum likelihood method and `fitdistcens` and `bootdistcens`.

It is assumed that the `distr` argument specifies the distribution by the probability density function and the cumulative distribution function (d, p). The quantile function and the random generator function (q, r) may be needed by other function such as `mmedist`, `qmedist`, `mgedist`, `fitdist`, `fitdistcens`, `bootdistcens` and `bootdist`.

For the following named distributions, reasonable starting values will be computed if `start` is omitted (i.e. NULL): "norm", "lnorm", "exp" and "pois", "cauchy", "gamma", "logis", "nbinom" (parametrized by mu and size), "geom", "beta", "weibull" from the stats package; all distributions (except phase-type distributions) from the actuar package. Note that these starting values may not be good enough if the fit is poor. The function uses a closed-form formula to fit the uniform distribution. If `start` is a list, then it should be a named list with the same names as in the d,p,q,r functions of the chosen distribution. If `start` is a function of data, then the function should return a named list with the same names as in the d,p,q,r functions of the chosen distribution.

The `mledist` function allows user to set a fixed values for some parameters. As for `start`, if `fix.arg` is a list, then it should be a named list with the same names as in the d,p,q,r functions of the chosen distribution. If `fix.arg` is a function of data, then the function should return a named list with the same names as in the d,p,q,r functions of the chosen distribution.

When `custom.optim=NULL` (the default), maximum likelihood estimations of the distribution parameters are computed with the R base `optim` or `constrOptim`. If no finite bounds (`lower=-Inf` and `upper=Inf`) are supplied, `optim` is used with the method specified by `optim.method`. Note

that `optim.method="default"` means `optim.method="Nelder-Mead"` for distributions with at least two parameters and `optim.method="BFGS"` for distributions with only one parameter. If finite bounds are supplied (among lower and upper) and `gradient != NULL`, `constrOptim` is used. If finite bounds are supplied (among lower and upper) and `gradient == NULL`, `constrOptim` is used when `optim.method="Nelder-Mead"`; `optim` is used when `optim.method="L-BFGS-B"` or `"Brent"`; in other case, an error is raised (same behavior as `constrOptim`).

When errors are raised by `optim`, it's a good idea to start by adding traces during the optimization process by adding `control=list(trace=1, REPORT=1)`.

If `custom.optim` is not `NULL`, then the user-supplied function is used instead of the R base `optim`. The `custom.optim` must have (at least) the following arguments `fn` for the function to be optimized, `par` for the initialized parameters. Internally the function to be optimized will also have other arguments, such as `obs` with observations and `ddistname` with distribution name for non censored data (Beware of potential conflicts with optional arguments of `custom.optim`). It is assumed that `custom.optim` should carry out a MINIMIZATION. Finally, it should return at least the following components `par` for the estimate, `convergence` for the convergence code, `value` for `fn(par)`, `hessian`, `counts` for the number of calls (function and gradient) and `message` (default to `NULL`) for the error message when `custom.optim` raises an error, see the returned value of `optim`. See examples in `fitdist` and `fitdistcens`.

Optionally, a vector of weights can be used in the fitting process. By default (when `weights=NULL`), ordinary MLE is carried out, otherwise the specified weights are used to balance the log-likelihood contributions. It is not yet possible to take into account weights in functions `plotdist`, `plotdistcens`, `plot.fitdist`, `plot.fitdistcens`, `cdfcomp`, `cdfcompdens`, `denscomp`, `ppcomp`, `qqcomp`, `gofstat`, `descdist`, `bootdist`, `bootdistcens` and `mgedist`. (developments planned in the future).

NB: if your data values are particularly small or large, a scaling may be needed before the optimization process. See Example (7).

Value

`mledist` returns a list with following components,

<code>estimate</code>	the parameter estimates.
<code>convergence</code>	an integer code for the convergence of <code>optim/constrOptim</code> defined as below or defined by the user in the user-supplied optimization function. 0 indicates successful convergence. 1 indicates that the iteration limit of <code>optim</code> has been reached. 10 indicates degeneracy of the Nealder-Mead simplex. 100 indicates that <code>optim</code> encountered an internal error.
<code>value</code>	the minimal value reached for the criterion to minimize.
<code>hessian</code>	a symmetric matrix computed by <code>optim</code> as an estimate of the Hessian at the solution found or computed in the user-supplied optimization function. It is used in <code>fitdist</code> to estimate standard errors.
<code>optim.function</code>	the name of the optimization function used for maximum likelihood.
<code>optim.method</code>	when <code>optim</code> is used, the name of the algorithm used, the field <code>method</code> of the <code>custom.optim</code> function otherwise.
<code>fix.arg</code>	the named list giving the values of parameters of the named distribution that must kept fixed rather than estimated by maximum likelihood or <code>NULL</code> if there are no such parameters.

fix.arg.fun	the function used to set the value of fix.arg or NULL.
weights	the vector of weights used in the estimation process or NULL.
counts	A two-element integer vector giving the number of calls to the log-likelihood function and its gradient respectively. This excludes those calls needed to compute the Hessian, if requested, and any calls to log-likelihood function to compute a finite-difference approximation to the gradient. counts is returned by optim or the user-supplied function or set to NULL.
optim.message	A character string giving any additional information returned by the optimizer, or NULL. To understand exactly the message, see the source code.
loglik	the log-likelihood value.
method	"closed formula" if appropriate otherwise NULL.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

- Venables WN and Ripley BD (2002), *Modern applied statistics with S*. Springer, New York, pp. 435-446, [doi:10.1007/9780387217062](https://doi.org/10.1007/9780387217062).
- Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, [doi:10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [mmedist](#), [qmedist](#), [mgedist](#), [fitdistr](#), [fitdistrib](#), [fitdistrib](#) for other estimation methods, [optim](#), [constrOptim](#) for optimization routines, [bootdistrib](#) and [bootdistrib](#) for bootstrap, and [llplot](#) for plotting the (log)likelihood.

Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) basic fit of a normal distribution with maximum likelihood estimation
#

x1 <- rnorm(n=100)
mledist(x1,"norm")

# (2) defining your own distribution functions, here for the Gumbel distribution
# for other distributions, see the CRAN task view dedicated to probability distributions

dgumbel <- function(x,a,b) 1/b*exp((a-x)/b)*exp(-exp((a-x)/b))
mledist(x1,"gumbel",start=list(a=10,b=5))

# (3) fit of a discrete distribution (Poisson)
#
```

```

x2 <- rpois(n=30,lambda = 2)
mledist(x2,"pois")

# (4) fit a finite-support distribution (beta)
#

x3 <- rbeta(n=100,shape1=5, shape2=10)
mledist(x3,"beta")

# (5) fit frequency distributions on USArrests dataset.
#

x4 <- USArrests$Assault
mledist(x4, "pois")
mledist(x4, "nbinom")

# (6) fit a continuous distribution (Gumbel) to censored data.
#

data(fluazinam)
log10EC50 <-log10(fluazinam)
# definition of the Gumbel distribution
dgumbel <- function(x,a,b) 1/b*exp((a-x)/b)*exp(-exp((a-x)/b))
pgumbel <- function(q,a,b) exp(-exp((a-q)/b))
qgumbel <- function(p,a,b) a-b*log(-log(p))

mledist(log10EC50,"gumbel",start=list(a=0,b=2),optim.method="Nelder-Mead")

# (7) scaling problem
# the simulated dataset (below) has particularly small values,
# hence without scaling ( $10^0$ ),
# the optimization raises an error. The for loop shows how scaling by  $10^i$ 
# for  $i=1,\dots,6$  makes the fitting procedure work correctly.

x2 <- rnorm(100, 1e-4, 2e-4)
for(i in 6:0)
  cat(i, try(mledist(x*10^i, "cauchy")$estimate, silent=TRUE), "\n")

# (17) small example for the zero-modified geometric distribution
#

dzmgeom <- function(x, p1, p2) p1 * (x == 0) + (1-p1)*dgeom(x-1, p2) #pdf
x2 <- c(2, 4, 0, 40, 4, 21, 0, 0, 0, 2, 5, 0, 13, 2) #simulated dataset
initp1 <- function(x) list(p1=mean(x == 0)) #init as MLE
mledist(x2, "zmgeom", fix.arg=initp1, start=list(p2=1/2))

```

Description

Fit of univariate distributions by matching moments (raw or centered) for non censored data.

Usage

```
mmedist(data, distr, order, memp, start = NULL, fix.arg = NULL, optim.method = "default",
        lower = -Inf, upper = Inf, custom.optim = NULL, weights = NULL, silent = TRUE,
        gradient = NULL, checkstartfix=FALSE, calcvcov=FALSE, ...)
```

Arguments

data	A numeric vector for non censored data.
distr	A character string "name" naming a distribution (see 'details').
order	A numeric vector for the moment order(s). The length of this vector must be equal to the number of parameters to estimate.
memp	A function implementing empirical moments, raw or centered but has to be consistent with <code>distr</code> argument (and <code>weights</code> argument). See details below.
start	A named list giving the initial values of parameters of the named distribution or a function of data computing initial values and returning a named list. This argument may be omitted (default) for some distributions for which reasonable starting values are computed (see the 'details' section of <code>mledist</code>).
fix.arg	An optional named list giving the values of fixed parameters of the named distribution or a function of data computing (fixed) parameter values and returning a named list. Parameters with fixed value are thus NOT estimated.
optim.method	"default" or optimization method to pass to <code>optim</code> .
lower	Left bounds on the parameters for the "L-BFGS-B" method (see <code>optim</code>).
upper	Right bounds on the parameters for the "L-BFGS-B" method (see <code>optim</code>).
custom.optim	a function carrying the optimization .
weights	an optional vector of weights to be used in the fitting process. Should be NULL or a numeric vector with strictly positive integers (typically the number of occurrences of each observation). If non-NULL, weighted MME is used, otherwise ordinary MME.
silent	A logical to remove or show warnings when bootstrapping.
gradient	A function to return the gradient of the squared difference for the "BFGS", "CG" and "L-BFGS-B" methods. If it is NULL, a finite-difference approximation will be used, see details.
checkstartfix	A logical to test starting and fixed values. Do not change it.
calcvcov	A logical indicating if (asymptotic) covariance matrix is required.
...	further arguments passed to the <code>optim</code> , <code>constrOptim</code> or <code>custom.optim</code> function.

Details

The argument `distr` can be one of the base R distributions: "norm", "lnorm", "exp" and "pois", "gamma", "logis", "nbinom", "geom", "beta" and "unif". In that case, no other arguments than `data` and `distr` are required, because the estimate is computed by a closed-form formula. For distributions characterized by one parameter ("geom", "pois" and "exp"), this parameter is simply estimated by matching theoretical and observed means, and for distributions characterized by two parameters, these parameters are estimated by matching theoretical and observed means and variances (Vose, 2000). Note that for these closed-form formula, `fix.arg` cannot be used and `start` is ignored.

The argument `distr` can also be the distribution name as long as a corresponding `mdistr` function exists, e.g. "pareto" if "mpareto" exists. In that case arguments `arguments` order and `memp` have to be supplied in order to carry out the matching numerically, by minimization of the sum of squared differences between observed and theoretical moments. Optionnally other arguments can be supplied to control optimization (see the 'details' section of `mledist` for details about arguments for the control of optimization). In that case, `fix.arg` can be used and `start` is taken into account.

For non closed-form estimators, `memp` must be provided to compute empirical moments. When `weights=NULL`, this function must have two arguments `x`, `order`: `x` the numeric vector of the data and `order` the order of the moment. When `weights!=NULL`, this function must have three arguments `x`, `order`, `weights`: `x` the numeric vector of the data, `order` the order of the moment, `weights` the numeric vector of weights. See examples below.

Optionally, a vector of `weights` can be used in the fitting process. By default (when `weights=NULL`), ordinary MME is carried out, otherwise the specified weights are used to compute (raw or centered) weighted moments. For closed-form estimators, weighted mean and variance are computed by `wtdmean` and `wtdvar` from the `Hmisc` package. When a numerical minimization is used, weighted are expected to be computed by the `memp` function. It is not yet possible to take into account weights in functions `plotdist`, `plotdistcens`, `plot.fitdist`, `plot.fitdistcens`, `cdfcomp`, `cdfcompdens`, `denscomp`, `ppcomp`, `qqcomp`, `gofstat` and `descdist` (developments planned in the future).

This function is not intended to be called directly but is internally called in `fitdist` and `bootdist` when used with the matching moments method.

Since Version 1.2-0, `mmedist` automatically computes the asymptotic covariance matrix using I. Ibragimov and R. Has'minskii (1981), hence the theoretical moments `mdist` should be defined up to an order which equals to twice the maximal order given `order`. For instance, the normal distribution, we fit against the expectation and the variance and we need to have `mnorm` up to order $2 \times 2 = 4$.

Value

`mmedist` returns a list with following components,

<code>estimate</code>	the parameter estimates.
<code>convergence</code>	an integer code for the convergence of <code>optim</code> defined as below or defined by the user in the user-supplied optimization function. 0 indicates successful convergence. 1 indicates that the iteration limit of <code>optim</code> has been reached. 10 indicates degeneracy of the Nelder-Mead simplex. 100 indicates that <code>optim</code> encountered an internal error.

value	the minimal value reached for the criterion to minimize.
hessian	a symmetric matrix computed by <code>optim</code> as an estimate of the Hessian at the solution found or computed in the user-supplied optimization function.
optim.function	(if appropriate) the name of the optimization function used for maximum likelihood.
optim.method	(if appropriate) when <code>optim</code> is used, the name of the algorithm used, the field method of the custom. <code>optim</code> function otherwise.
fix.arg	the named list giving the values of parameters of the named distribution that must kept fixed rather than estimated by maximum likelihood or NULL if there are no such parameters.
fix.arg.fun	the function used to set the value of <code>fix.arg</code> or NULL.
weights	the vector of weights used in the estimation process or NULL.
counts	A two-element integer vector giving the number of calls to the log-likelihood function and its gradient respectively. This excludes those calls needed to compute the Hessian, if requested, and any calls to log-likelihood function to compute a finite-difference approximation to the gradient. <code>counts</code> is returned by <code>optim</code> or the user-supplied function or set to NULL.
optim.message	A character string giving any additional information returned by the optimizer, or NULL. To understand exactly the message, see the source code.
loglik	the log-likelihood value.
method	either "closed formula" or the name of the optimization method.
order	the order of the moment(s) matched.
memp	the empirical moment function.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

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See Also

See [mledist](#), [qmedist](#), [mgedist](#), [fitdist](#), [fitdistcens](#), [optim](#), [bootdistcens](#) and [bootdist](#).
Please visit the [Frequently Asked Questions](#).

Examples

```

set.seed(123) # here just to make random sampling reproducible

# (1) basic fit of a normal distribution with moment matching estimation
#

n <- 100
x1 <- rnorm(n=n)
mmedist(x1, "norm")

#weighted
w <- c(rep(1, n/2), rep(10, n/2))
mmedist(x1, "norm", weights=w)$estimate

# (2) fit a discrete distribution (Poisson)
#

x2 <- rpois(n=30,lambda = 2)
mmedist(x2, "pois")

# (3) fit a finite-support distribution (beta)
#

x3 <- rbeta(n=100,shape1=5, shape2=10)
mmedist(x3, "beta")

# (4) fit a Pareto distribution
#

require("actuar")
#simulate a sample
x4 <- rpareto(1000, 6, 2)

#empirical raw moment
memp <- function(x, order) mean(x^order)
memp2 <- function(x, order, weights) sum(x^order * weights)/sum(weights)

#fit by MME
mmedist(x4, "pareto", order=c(1, 2), memp=memp,
        start=list(shape=10, scale=10), lower=1, upper=Inf)
#fit by weighted MME
w <- rep(1, length(x4))
w[x4 < 1] <- 2
mmedist(x4, "pareto", order=c(1, 2), memp=memp2, weights=w,
        start=list(shape=10, scale=10), lower=1, upper=Inf)

```

msedist	<i>Maximum spacing estimation of univariate distributions</i>
---------	---

Description

Fit of univariate distribution by maximizing (log) spacings for non censored data.

Usage

```
msedist(data, distr, phidiv="KL", power.phidiv=NULL, start = NULL, fix.arg = NULL,
  optim.method = "default", lower = -Inf, upper = Inf, custom.optim = NULL,
  weights=NULL, silent = TRUE, gradient = NULL, checkstartfix=FALSE, calvcov=FALSE, ...)
```

Arguments

data	A numeric vector for non censored data.
distr	A character string "name" naming a distribution for which the corresponding quantile function qname and the corresponding density distribution dname must be classically defined.
phidiv	A character string coding for the name of the phi-divergence used : "KL" for Kullback-Leibler information (corresponds to classic maximum spacing estimation), "J" for Jeffreys' divergence, "R" for Renyi's divergence, "H" for Hellinger distance, "V" for Vajda's measure of information, see details.
power.phidiv	If relevant, a numeric for the power used in some phi-divergence : should be NULL when phidiv="KL" or phidiv="J" , should be positive and different from 1 when phidiv="R", should be greater or equal to 1 when phidiv="H" or phidiv="V", see details.
start	A named list giving the initial values of parameters of the named distribution or a function of data computing initial values and returning a named list. This argument may be omitted (default) for some distributions for which reasonable starting values are computed (see the 'details' section of mledist).
fix.arg	An optional named list giving the values of fixed parameters of the named distribution or a function of data computing (fixed) parameter values and returning a named list. Parameters with fixed value are thus NOT estimated.
optim.method	"default" or optimization method to pass to optim .
lower	Left bounds on the parameters for the "L-BFGS-B" method (see optim).
upper	Right bounds on the parameters for the "L-BFGS-B" method (see optim).
custom.optim	a function carrying the optimization.
weights	an optional vector of weights to be used in the fitting process. Should be NULL or a numeric vector with strictly positive integers (typically the number of occurrences of each observation). If non-NULL, weighted MSE is used, otherwise ordinary MSE.
silent	A logical to remove or show warnings when bootstrapping.

gradient	A function to return the gradient of the gof distance for the "BFGS", "CG" and "L-BFGS-B" methods. If it is NULL, a finite-difference approximation will be used.
checkstartfix	A logical to test starting and fixed values. Do not change it.
calvcov	A logical indicating if (asymptotic) covariance matrix is required. (currently ignored)
...	further arguments passed to the <code>optim</code> , <code>constrOptim</code> or <code>custom.optim</code> function.

Details

The `msedist` function numerically maximizes a phi-divergence function of spacings, where spacings are the differences of the cumulative distribution function evaluated at the sorted dataset. The classical maximum spacing estimation (MSE) was introduced by Cheng and Amin (1986) and Ranneby (1984) independently where the phi-divergence is the logarithm, see Anatolyev and Kosenok (2005) for a link between MSE and maximum likelihood estimation.

MSE was generalized by Ranneby and Ekstrom (1997) by allowing different phi-divergence function. Generalized MSE maximizes

$$S_n(\theta) = \frac{1}{n+1} \sum_{i=1}^{n+1} \phi(F(x_{(i)}; \theta) - F(x_{(i-1)}; \theta)),$$

where $F(; \theta)$ is the parametric distribution function to be fitted, ϕ is the phi-divergence function, $x_{(1)} < \dots < x_{(n)}$ is the sorted sample, $x_{(0)} = -\infty$ and $x_{(n+1)} = +\infty$. The possible phi-divergence function is

- Kullback-Leibler information (when `phidiv="KL"` and corresponds to classical MSE)

$$\phi(x) = \log(x)$$

- Jeffreys' divergence (when `phidiv="J"`)

$$\phi(x) = (1-x) \log(x)$$

- Renyi's divergence (when `phidiv="R"` and `power.phidiv=alpha`)

$$\phi(x) = x^\alpha \times \text{sign}(1-x) \text{ with } \alpha > 0, \alpha \neq 1$$

- Hellinger distance (when `phidiv="H"` and `power.phidiv=p`)

$$\phi(x) = -|1-x^{1/p}|^p \text{ with } p \geq 1$$

- Vajda's measure of information (when `phidiv="V"` and `power.phidiv=beta`)

$$\phi(x) = -|1-x|^\beta \text{ with } \beta \geq 1$$

The optimization process is the same as `mledist`, see the 'details' section of that function.

This function is not intended to be called directly but is internally called in `fitdist` and `bootdist`.

This function is intended to be used only with non-censored data.

NB: if your data values are particularly small or large, a scaling may be needed before the optimization process, see `mledist`'s examples.

Value

msedist returns a list with following components,

estimate	the parameter estimates.
convergence	an integer code for the convergence of <code>optim</code> defined as below or defined by the user in the user-supplied optimization function. 0 indicates successful convergence. 1 indicates that the iteration limit of <code>optim</code> has been reached. 10 indicates degeneracy of the Nelder-Mead simplex. 100 indicates that <code>optim</code> encountered an internal error.
value	the minimal value reached for the criterion to minimize.
hessian	a symmetric matrix computed by <code>optim</code> as an estimate of the Hessian at the solution found or computed in the user-supplied optimization function.
optim.function	the name of the optimization function used for maximum likelihood.
optim.method	when <code>optim</code> is used, the name of the algorithm used, the field <code>method</code> of the custom <code>optim</code> function otherwise.
fix.arg	the named list giving the values of parameters of the named distribution that must kept fixed rather than estimated by maximum likelihood or NULL if there are no such parameters.
fix.arg.fun	the function used to set the value of <code>fix.arg</code> or NULL.
weights	the vector of weights used in the estimation process or NULL.
counts	A two-element integer vector giving the number of calls to the log-likelihood function and its gradient respectively. This excludes those calls needed to compute the Hessian, if requested, and any calls to log-likelihood function to compute a finite-difference approximation to the gradient. <code>counts</code> is returned by <code>optim</code> or the user-supplied function or set to NULL.
optim.message	A character string giving any additional information returned by the optimizer, or NULL. To understand exactly the message, see the source code.
loglik	the log-likelihood value.
phidiv	The character string coding for the name of the phi-divergence used either "KL", "J", "R", "H" or "V".
power.phidiv	Either NULL or a numeric for the power used in the phi-divergence.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

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Ranneby, B. and Ekstroem, M. (1997). *Maximum spacing estimates based on different metrics*. Umea universitet.

See Also

See [mmedist](#), [mledist](#), [qmedist](#), [mgedist](#), [fitdist](#) for other estimation methods.

Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) Fit of a Weibull distribution to serving size data by maximum
# spacing estimation
#

data(groundbeef)
serving <- groundbeef$serving
msedist(serving, "weibull")

# (2) Fit of an exponential distribution
#

x1 <- rexp(1e3)
#the convergence is quick
msedist(x1, "exp", control=list(trace=0, REPORT=1))
```

plotdist

Plot of empirical and theoretical distributions for non-censored data

Description

Plots an empirical distribution (non-censored data) with a theoretical one if specified.

Usage

```
plotdist(data, distr, para, histo = TRUE, breaks = "default",
  demp = FALSE, discrete, ...)
```

Arguments

data	A numeric vector.
distr	A character string "name" naming a distribution for which the corresponding density function dname, the corresponding distribution function pname and the corresponding quantile function qname must be defined, or directly the density function. This argument may be omitted only if para is omitted.
para	A named list giving the parameters of the named distribution. This argument may be omitted only if distr is omitted.
histo	A logical to plot the histogram using the hist function.
breaks	If "default" the histogram is plotted with the function hist with its default breaks definition. Else breaks is passed to the function hist . This argument is not taken into account if discrete is TRUE.
demp	A logical to plot the empirical density on the first plot (alone or superimposed on the histogram depending of the value of the argument histo) using the density function.
discrete	If TRUE, the distribution is considered as discrete. If both distr and discrete are missing, discrete is set to FALSE. If discrete is missing but not distr, discrete is set to TRUE when distr belongs to "binom", "nbinom", "geom", "hyper" or "pois".
...	further graphical arguments passed to graphical functions used in plotdist.

Details

Empirical and, if specified, theoretical distributions are plotted in density and in cdf. For the plot in density, the user can use the arguments histo and demp to specify if he wants the histogram using the function [hist](#), the density plot using the function [density](#), or both (at least one of the two arguments must be put to "TRUE"). For continuous distributions, the function [hist](#) is used with its default breaks definition if breaks is "default" or passing breaks as an argument if it differs from "default". For continuous distribution and when a theoretical distribution is specified by both arguments distname and para, Q-Q plot (plot of the quantiles of the theoretical fitted distribution (x-axis) against the empirical quantiles of the data) and P-P plot (i.e. for each value of the data set, plot of the cumulative density function of the fitted distribution (x-axis) against the empirical cumulative density function (y-axis)) are also given (Cullen and Frey, 1999). The function [ppoints](#) (with default parameter for argument a) is used for the Q-Q plot, to generate the set of probabilities at which to evaluate the inverse distribution. NOTE THAT FROM VERSION 0.4-3, [ppoints](#) is also used for P-P plot and cdf plot for continuous data. To personalize the four plots proposed for continuous data, for example to change the plotting position, we recommend the use of functions [cdfcomp](#), [denscomp](#), [qqcomp](#) and [ppcomp](#).

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

Cullen AC and Frey HC (1999), *Probabilistic techniques in exposure assessment*. Plenum Press, USA, pp. 81-155.

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:10.18637/jss.v064.i04.

See Also

See [graphcomp](#), [descdist](#), [hist](#), [plot](#), [plotdistcens](#) and [ppoints](#).

Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) Plot of an empirical distribution with changing
# of default line types for CDF and colors
# and optionally adding a density line
#
x1 <- rnorm(n=30)
plotdist(x1)
plotdist(x1,demp = TRUE)
plotdist(x1,histo = FALSE, demp = TRUE)
plotdist(x1, col="blue", type="b", pch=16)
plotdist(x1, type="s")

# (2) Plot of a discrete distribution against data
#
x2 <- rpois(n=30, lambda = 2)
plotdist(x2, discrete=TRUE)
plotdist(x2, "pois", para=list(lambda = mean(x2)))
plotdist(x2, "pois", para=list(lambda = mean(x2)), lwd="2")

# (3) Plot of a continuous distribution against data
#
xn <- rnorm(n=100, mean=10, sd=5)
plotdist(xn, "norm", para=list(mean=mean(xn), sd=sd(xn)))
plotdist(xn, "norm", para=list(mean=mean(xn), sd=sd(xn)), pch=16)
plotdist(xn, "norm", para=list(mean=mean(xn), sd=sd(xn)), demp = TRUE)
plotdist(xn, "norm", para=list(mean=mean(xn), sd=sd(xn)),
histo = FALSE, demp = TRUE)

# (4) Plot of serving size data
#
data(groundbeef)
plotdist(groundbeef$serving, type="s")

# (5) Plot of numbers of parasites with a Poisson distribution
data(toxocara)
number <- toxocara$number
plotdist(number, discrete = TRUE)
plotdist(number,"pois",para=list(lambda=mean(number)))
```

plotdistcens *Plot of empirical and theoretical distributions for censored data*

Description

Plots an empirical distribution for censored data with a theoretical one if specified.

Usage

```
plotdistcens(censdata, distr, para, leftNA = -Inf, rightNA = Inf,
             NPMLE = TRUE, Turnbull.confint = FALSE,
             NPMLE.method = "Wang", ...)
```

Arguments

censdata	A dataframe of two columns respectively named left and right, describing each observed value as an interval. The left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The right column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations.
distr	A character string "name" naming a distribution, for which the corresponding density function dname and the corresponding distribution function pname must be defined, or directly the density function.
para	A named list giving the parameters of the named distribution. This argument may be omitted only if distr is omitted.
leftNA	the real value of the left bound of left censored observations : -Inf or a finite value such as 0 for positive data for example.
rightNA	the real value of the right bound of right censored observations : Inf or a finite value such as a realistic maximum value.
NPMLE	if TRUE an NPMLE (nonparametric maximum likelihood estimate) technique is used to estimate the cdf curve of the censored data and previous arguments leftNA and rightNA are not used (see details)
Turnbull.confint	if TRUE confidence intervals will be added to the Turnbull plot. In that case NPMLE.method is forced to "Turnbull.middlepoints"
NPMLE.method	Three NPMLE techniques are provided, "Wang", the default one, rewritten from the package npsurv using function constrOptim from the package stats for optimisation, "Turnbull.middlepoints", an older one which is implemented in the package survival and "Turnbull.intervals" that uses the same Turnbull algorithm from the package survival but associates an interval to each equivalence class instead of the midpoint of this interval (see details). Only "Wang" and "Turnbull.intervals" enable the derivation of a Q-Q plot and a P-P plot.
...	further graphical arguments passed to other methods. The title of the plot can be modified using the argument main only for the CDF plot.

Details

If `NPMLE` is `TRUE`, and `NPMLE.method` is `"Wang"`, empirical distributions are plotted in cdf using either the constrained Newton method (Wang, 2008) or the hierarchical constrained Newton method (Wang, 2013) to compute the overall empirical cdf curve. If `NPMLE` is `TRUE`, and `NPMLE.method` is `"Turnbull.intervals"`, empirical are plotted in cdf using the EM approach of Turnbull (Turnbull, 1974). In those two cases, grey rectangles represent areas where the empirical distribution function is not unique. In cases where a theoretical distribution is specified, two goodness-of-fit plots are also provided, a Q-Q plot (plot of the quantiles of the theoretical fitted distribution (x-axis) against the empirical quantiles of the data) and a P-P plot (i.e. for each value of the data set, plot of the cumulative density function of the fitted distribution (x-axis) against the empirical cumulative density function (y-axis)). Grey rectangles in a Q-Q plot or a P-P plot also represent areas of non uniqueness of empirical quantiles or probabilities, directly derived from non uniqueness areas of the empirical cumulative distribution.

If `NPMLE` is `TRUE`, and `NPMLE.method` is `"Turnbull.middlepoints"`, empirical and, if specified, theoretical distributions are plotted in cdf using the EM approach of Turnbull (Turnbull, 1974) to compute the overall empirical cdf curve, with confidence intervals if `Turnbull.confint` is `TRUE`, by calls to functions `survfit` and `plot.survfit` from the `survival` package.

If `NPMLE` is `FALSE` empirical and, if specified, theoretical distributions are plotted in cdf, with data directly reported as segments for interval, left and right censored data, and as points for non-censored data. Before plotting, observations are ordered and a rank r is associated to each of them. Left censored observations are ordered first, by their right bounds. Interval censored and non censored observations are then ordered by their mid-points and, at last, right censored observations are ordered by their left bounds. If `leftNA` (resp. `rightNA`) is finite, left censored (resp. right censored) observations are considered as interval censored observations and ordered by mid-points with non-censored and interval censored data. It is sometimes necessary to fix `rightNA` or `leftNA` to a realistic extreme value, even if not exactly known, to obtain a reasonable global ranking of observations. After ranking, each of the n observations is plotted as a point (one x-value) or a segment (an interval of possible x-values), with a y-value equal to r/n , r being the rank of each observation in the global ordering previously described. This second method may be interesting but is certainly less rigorous than the other methods that should be preferred.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

- Turnbull BW (1974), *Nonparametric estimation of a survivorship function with doubly censored data*. Journal of American Statistical Association, 69, 169-173, doi:10.2307/2285518.
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See Also

See [plotdist](#), [survfit.formula](#).

Please visit the [Frequently Asked Questions](#).

Examples

```
# (1) Plot of an empirical censored distribution (censored data) as a CDF
# using the default Wang method
#
data(smokedfish)
d1 <- as.data.frame(log10(smokedfish))
plotdistcens(d1)

# (2) Add the CDF of a normal distribution
#
plotdistcens(d1, "norm", para=list(mean = -1.6, sd = 1.5))

# (3) Various plots of the same empirical distribution
#
# default Wang plot with representation of equivalence classes
plotdistcens(d1, NPML = TRUE, NPML.method = "Wang")
# same plot but using the Turnbull algorithm from the package survival
plotdistcens(d1, NPML = TRUE, NPML.method = "Wang")
# Turnbull plot with middlepoints (as in the package survival)
plotdistcens(d1, NPML = TRUE, NPML.method = "Turnbull.middlepoints")
# Turnbull plot with middlepoints and confidence intervals
plotdistcens(d1, NPML = TRUE, NPML.method = "Turnbull.middlepoints", Turnbull.confint = TRUE)
# with intervals and points
plotdistcens(d1, rightNA=3, NPML = FALSE)
# with intervals and points
# defining a minimum value for left censored values
plotdistcens(d1, leftNA=-3, NPML = FALSE)
```

prefit

Pre-fitting procedure

Description

Search good starting values

Usage

```
prefit(data, distr, method = c("mle", "mme", "qme", "mge"),
       feasible.par, memp=NULL, order=NULL,
       probs=NULL, qtype=7, gof=NULL, fix.arg=NULL, lower,
       upper, weights=NULL, silent=TRUE, ...)
```

Arguments

<code>data</code>	A numeric vector.
<code>distr</code>	A character string "name" naming a distribution for which the corresponding density function <code>dname</code> , the corresponding distribution function <code>pname</code> and the corresponding quantile function <code>qname</code> must be defined, or directly the density function.
<code>method</code>	A character string coding for the fitting method: "mle" for 'maximum likelihood estimation', "mme" for 'moment matching estimation', "qme" for 'quantile matching estimation' and "mge" for 'maximum goodness-of-fit estimation'.
<code>feasible.par</code>	A named list giving the initial values of parameters of the named distribution or a function of data computing initial values and returning a named list. This argument may be omitted (default) for some distributions for which reasonable starting values are computed (see the 'details' section of mledist). It may not be into account for closed-form formulas.
<code>order</code>	A numeric vector for the moment order(s). The length of this vector must be equal to the number of parameters to estimate.
<code>memp</code>	A function implementing empirical moments, raw or centered but has to be consistent with <code>distr</code> argument (and <code>weights</code> argument).
<code>probs</code>	A numeric vector of the probabilities for which the quantile matching is done. The length of this vector must be equal to the number of parameters to estimate.
<code>qtype</code>	The quantile type used by the R quantile function to compute the empirical quantiles, (default 7 corresponds to the default quantile method in R).
<code>gof</code>	A character string coding for the name of the goodness-of-fit distance used : "CvM" for Cramer-von Mises distance, "KS" for Kolmogorov-Smirnov distance, "AD" for Anderson-Darling distance, "ADR", "ADL", "AD2R", "AD2L" and "AD2" for variants of Anderson-Darling distance described by Luceno (2006).
<code>fix.arg</code>	An optional named list giving the values of fixed parameters of the named distribution or a function of data computing (fixed) parameter values and returning a named list. Parameters with fixed value are thus NOT estimated by this maximum likelihood procedure. The use of this argument is not possible if <code>method="mme"</code> and a closed-form formula is used.
<code>weights</code>	an optional vector of weights to be used in the fitting process. Should be NULL or a numeric vector. If non-NULL, weighted MLE is used, otherwise ordinary MLE.
<code>silent</code>	A logical to remove or show warnings.
<code>lower</code>	Lower bounds on the parameters.

upper Upper bounds on the parameters.

... Further arguments to be passed to generic functions, or to one of the functions "mledist", "mmedist", "qmedist" or "mgedist" depending of the chosen method. See [mledist](#), [mmedist](#), [qmedist](#), [mgedist](#) for details on parameter estimation.

Details

Searching good starting values is achieved by transforming the parameters (from their constraint interval to the real line) of the probability distribution. Indeed,

- positive parameters in $(0, Inf)$ are transformed using the logarithm (typically the scale parameter `sd` of a normal distribution, see [Normal](#)),
- parameters in $(1, Inf)$ are transformed using the function $\log(x - 1)$,
- probability parameters in $(0, 1)$ are transformed using the logit function $\log(x/(1 - x))$ (typically the parameter `prob` of a geometric distribution, see [Geometric](#)),
- negative probability parameters in $(-1, 0)$ are transformed using the function $\log(-x/(1 + x))$,
- real parameters are of course not transformed at all, typically the mean of a normal distribution, see [Normal](#).

Once parameters are transformed, an optimization is carried out by a quasi-Newton algorithm (typically BFGS) and then we transform them back to original parameter value.

Value

A named list.

Author(s)

Christophe Dutang and Marie-Laure Delignette-Muller.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, [doi:10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [mledist](#), [mmedist](#), [qmedist](#), [mgedist](#) for details on parameter estimation. See [fitdistr](#) for the main procedure.

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) fit of a gamma distribution by maximum likelihood estimation
#
x <- rgamma(1e3, 5/2, 7/2)
```

```
prefit(x, "gamma", "mle", list(shape=3, scale=3), lower=-Inf, upper=Inf)
```

qmedist

Quantile matching fit of univariate distributions

Description

Fit of univariate distribution by matching quantiles for non censored data.

Usage

```
qmedist(data, distr, probs, start = NULL, fix.arg = NULL, qtype = 7,
        optim.method = "default", lower = -Inf, upper = Inf,
        custom.optim = NULL, weights = NULL, silent = TRUE, gradient = NULL,
        checkstartfix=FALSE, calcvcov=FALSE, ...)
```

Arguments

data	A numeric vector for non censored data.
distr	A character string "name" naming a distribution for which the corresponding quantile function qname and the corresponding density distribution dname must be classically defined.
probs	A numeric vector of the probabilities for which the quantile matching is done. The length of this vector must be equal to the number of parameters to estimate.
start	A named list giving the initial values of parameters of the named distribution or a function of data computing initial values and returning a named list. This argument may be omitted (default) for some distributions for which reasonable starting values are computed (see the 'details' section of mledist).
fix.arg	An optional named list giving the values of fixed parameters of the named distribution or a function of data computing (fixed) parameter values and returning a named list. Parameters with fixed value are thus NOT estimated.
qtype	The quantile type used by the R quantile function to compute the empirical quantiles, (default 7 corresponds to the default quantile method in R).
optim.method	"default" or optimization method to pass to optim .
lower	Left bounds on the parameters for the "L-BFGS-B" method (see optim).
upper	Right bounds on the parameters for the "L-BFGS-B" method (see optim).
custom.optim	a function carrying the optimization.
weights	an optional vector of weights to be used in the fitting process. Should be NULL or a numeric vector with strictly positive integers (typically the number of occurrences of each observation). If non-NULL, weighted QME is used, otherwise ordinary QME.

silent	A logical to remove or show warnings when bootstrapping.
gradient	A function to return the gradient of the squared difference for the "BFGS", "CG" and "L-BFGS-B" methods. If it is NULL, a finite-difference approximation will be used, see details.
checkstartfix	A logical to test starting and fixed values. Do not change it.
calvcov	A logical indicating if (asymptotic) covariance matrix is required. (currently ignored)
...	further arguments passed to the <code>optim</code> , <code>constrOptim</code> or <code>custom.optim</code> function.

Details

The `qmedist` function carries out the quantile matching numerically, by minimization of the sum of squared differences between observed and theoretical quantiles. Note that for discrete distribution, the sum of squared differences is a step function and consequently, the optimum is not unique, see the FAQ.

The optimization process is the same as `mledist`, see the 'details' section of that function.

Optionally, a vector of weights can be used in the fitting process. By default (when `weights=NULL`), ordinary QME is carried out, otherwise the specified weights are used to compute weighted quantiles used in the squared differences. Weighted quantiles are computed by `wtdquantile` from the `Hmisc` package. It is not yet possible to take into account weights in functions `plotdist`, `plotdistcens`, `plot.fitdist`, `plot.fitdistcens`, `cdfcomp`, `cdfcompdens`, `denscomp`, `ppcomp`, `qqcomp`, `gofstat` and `descdist` (developments planned in the future).

This function is not intended to be called directly but is internally called in `fitdist` and `bootdist`.

Value

`qmedist` returns a list with following components,

estimate	the parameter estimates.
convergence	an integer code for the convergence of <code>optim</code> defined as below or defined by the user in the user-supplied optimization function. 0 indicates successful convergence. 1 indicates that the iteration limit of <code>optim</code> has been reached. 10 indicates degeneracy of the Nelder-Mead simplex. 100 indicates that <code>optim</code> encountered an internal error.
value	the minimal value reached for the criterion to minimize.
hessian	a symmetric matrix computed by <code>optim</code> as an estimate of the Hessian at the solution found or computed in the user-supplied optimization function.
optim.function	the name of the optimization function used for maximum likelihood.
optim.method	when <code>optim</code> is used, the name of the algorithm used, the field <code>method</code> of the <code>custom.optim</code> function otherwise.
fix.arg	the named list giving the values of parameters of the named distribution that must kept fixed rather than estimated by maximum likelihood or NULL if there are no such parameters.
fix.arg.fun	the function used to set the value of <code>fix.arg</code> or NULL.

weights	the vector of weights used in the estimation process or NULL.
counts	A two-element integer vector giving the number of calls to the log-likelihood function and its gradient respectively. This excludes those calls needed to compute the Hessian, if requested, and any calls to log-likelihood function to compute a finite-difference approximation to the gradient. counts is returned by <code>optim</code> or the user-supplied function or set to NULL.
optim.message	A character string giving any additional information returned by the optimizer, or NULL. To understand exactly the message, see the source code.
loglik	the log-likelihood value.
probs	the probability vector on which quantiles are matched.

Author(s)

Christophe Dutang and Marie Laure Delignette-Muller.

References

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- Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:[10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See `mmedist`, `mledist`, `mgedist`, `fitdistr` for other estimation methods and `quantile` for empirical quantile estimation in R.

Please visit the [Frequently Asked Questions](#).

Examples

```
set.seed(123) # here just to make random sampling reproducible

# (1) basic fit of a normal distribution
#
x1 <- rnorm(n=100)
qmedist(x1, "norm", probs=c(1/3, 2/3))

# (2) defining your own distribution functions, here for the Gumbel
# distribution for other distributions, see the CRAN task view dedicated
# to probability distributions

dgumbel <- function(x, a, b) 1/b*exp((a-x)/b)*exp(-exp((a-x)/b))
qgumbel <- function(p, a, b) a - b*log(-log(p))
qmedist(x1, "gumbel", probs=c(1/3, 2/3), start=list(a=10,b=5))

# (3) fit a discrete distribution (Poisson)
```

```

#

x2 <- rpois(n=30,lambda = 2)
qmedist(x2, "pois", probs=1/2)

# (4) fit a finite-support distribution (beta)
#

x3 <- rbeta(n=100,shape1=5, shape2=10)
qmedist(x3, "beta", probs=c(1/3, 2/3))

# (5) fit frequency distributions on USArrests dataset.
#

x4 <- USArrests$Assault
qmedist(x4, "pois", probs=1/2)
qmedist(x4, "nbinom", probs=c(1/3, 2/3))

```

quantile

Quantile estimation from a fitted distribution

Description

Quantile estimation from a fitted distribution, optionally with confidence intervals calculated from the bootstrap result.

Usage

```

## S3 method for class 'fitdist'
quantile(x, probs = seq(0.1, 0.9, by=0.1), ...)
## S3 method for class 'fitdistcens'
quantile(x, probs = seq(0.1, 0.9, by=0.1), ...)
## S3 method for class 'bootdist'
quantile(x, probs = seq(0.1, 0.9, by=0.1), CI.type = "two.sided",
  CI.level = 0.95, ...)
## S3 method for class 'bootdistcens'
quantile(x, probs = seq(0.1, 0.9, by=0.1), CI.type = "two.sided",
  CI.level = 0.95, ...)
## S3 method for class 'quantile.fitdist'
print(x, ...)
## S3 method for class 'quantile.fitdistcens'
print(x, ...)
## S3 method for class 'quantile.bootdist'
print(x, ...)
## S3 method for class 'quantile.bootdistcens'
print(x, ...)

```

Arguments

x	An object of class "fitdist", "fitdistcens", "bootdist", "bootdistcens" or "quantile.fitdist", "quantile.fitdistcens", "quantile.bootdist", "quantile.bootdistcens" for the print generic function.
probs	A numeric vector of probabilities with values in [0, 1] at which quantiles must be calculated.
CI.type	Type of confidence intervals: either "two.sided" or one-sided intervals ("less" or "greater").
CI.level	The confidence level.
...	Further arguments to be passed to generic functions.

Details

Quantiles of the parametric distribution are calculated for each probability specified in probs, using the estimated parameters. When used with an object of class "bootdist" or "bootdistcens", percentile confidence intervals and medians estimates are also calculated from the bootstrap result. If CI.type is two.sided, the CI.level two-sided confidence intervals of quantiles are calculated. If CI.type is less or greater, the CI.level one-sided confidence intervals of quantiles are calculated. The print functions show the estimated quantiles with percentile confidence intervals and median estimates when a bootstrap resampling has been done previously, and the number of bootstrap iterations for which the estimation converges if it is inferior to the whole number of bootstrap iterations.

Value

quantile returns a list with 2 components (the first two described below) when called with an object of class "fitdist" or "fitdistcens" and 8 components (described below) when called with an object of class "bootdist" or "bootdistcens":

quantiles	a dataframe containing the estimated quantiles for each probability value specified in the argument probs (one row, and as many columns as values in probs).
probs	the numeric vector of probabilities at which quantiles are calculated.
bootquant	A data frame containing the bootstrapped values for each quantile (many rows, as specified in the call to <code>bootdist</code> in the argument niter, and as many columns as values in probs)
quantCI	If CI.type is two.sided, the two bounds of the CI.level percent two.sided confidence interval for each quantile (two rows and as many columns as values in probs). If CI.type is less, right bound of the CI.level percent one.sided confidence interval for each quantile (one row). If CI.type is greater, left bound of the CI.level percent one.sided confidence interval for each quantile (one row).
quantmedian	Median of bootstrap estimates (per probability).
CI.type	Type of confidence interval: either "two.sided" or one-sided intervals ("less" or "greater").
CI.level	The confidence level.
nbboot	The number of samples drawn by bootstrap.
nbconverg	The number of iterations for which the optimization algorithm converges.

Author(s)

Marie-Laure Delignette-Muller and Christophe Dutang.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:10.18637/jss.v064.i04.

See Also

See [fitdist](#), [bootdist](#), [fitdistcens](#), [bootdistcens](#) and [CIcdfplot](#).

Please visit the [Frequently Asked Questions](#).

Examples

```
# (1) Fit of a normal distribution on acute toxicity log-transformed values of
# endosulfan for nonarthropod invertebrates, using maximum likelihood estimation
# to estimate what is called a species sensitivity distribution
# (SSD) in ecotoxicology, followed by estimation of the 5, 10 and 20 percent quantile
# values of the fitted distribution, which are called the 5, 10, 20 percent hazardous
# concentrations (HC5, HC10, HC20) in ecotoxicology, followed with calculations of their
# confidence intervals with various definitions, from a small number of bootstrap
# iterations to satisfy CRAN running times constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.
#
data(endosulfan)
ATV <- subset(endosulfan, group == "NonArthroInvert")$ATV
log10ATV <- log10(subset(endosulfan, group == "NonArthroInvert")$ATV)
fln <- fitdist(log10ATV, "norm")
quantile(fln, probs = c(0.05, 0.1, 0.2))
bln <- bootdist(fln, bootmethod="param", niter=101)
quantile(bln, probs = c(0.05, 0.1, 0.2))
quantile(bln, probs = c(0.05, 0.1, 0.2), CI.type = "greater")
quantile(bln, probs = c(0.05, 0.1, 0.2), CI.level = 0.9)

# (2) Draw of 95 percent confidence intervals on quantiles of the
# previously fitted distribution
#
cdfcomp(fln)
q1 <- quantile(bln, probs = seq(0,1,length=101))
points(q1$quantCI[1,],q1$probs,type="l")
points(q1$quantCI[2,],q1$probs,type="l")

# (2b) Draw of 95 percent confidence intervals on quantiles of the
# previously fitted distribution
# using the NEW function CIcdfplot
#
CIcdfplot(bln, CI.output = "quantile", CI.fill = "pink")

# (3) Fit of a distribution on acute salinity log-transformed tolerance
# for riverine macro-invertebrates, using maximum likelihood estimation
```

```

# to estimate what is called a species sensitivity distribution
# (SSD) in ecotoxicology, followed by estimation of the 5, 10 and 20 percent quantile
# values of the fitted distribution, which are called the 5, 10, 20 percent hazardous
# concentrations (HC5, HC10, HC20) in ecotoxicology, followed with calculations of
# their confidence intervals with various definitions.
# from a small number of bootstrap iterations to satisfy CRAN running times constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.
#
data(salinity)
log10LC50 <- log10(salinity)
flncens <- fitdistcens(log10LC50, "norm")
quantile(flncens, probs = c(0.05, 0.1, 0.2))
blncens <- bootdistcens(flncens, niter = 101)
quantile(blncens, probs = c(0.05, 0.1, 0.2))
quantile(blncens, probs = c(0.05, 0.1, 0.2), CI.type = "greater")
quantile(blncens, probs = c(0.05, 0.1, 0.2), CI.level = 0.9)

```

salinity

Species-Sensitivity Distribution (SSD) for salinity tolerance

Description

72-hour acute salinity tolerance (LC50 values) of riverine macro-invertebrates.

Usage

```
data(salinity)
```

Format

salinity is a data frame with 2 columns named left and right, describing each observed LC50 value (in electrical conductivity, millisiemens per centimeter) as an interval. The left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The right column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for noncensored observations.

Source

Kefford, B.J., Nuggeoda, D., Metzeling, L., Fields, E. 2006. Validating species sensitivity distributions using salinity tolerance of riverine macroinvertebrates in the southern Murray-darling Basin (Vitoria, Australia). *Canadian Journal of Fisheries and Aquatic Science*, **63**, 1865-1877.

Examples

```

# (1) load of data
#
data(salinity)

```

```

# (2) plot of data using Turnbull cdf plot
#
log10LC50 <- log10(salinity)
plotdistcens(log10LC50)

# (3) fit of a normal and a logistic distribution to data in log10
# (classical distributions used for species sensitivity
# distributions, SSD, in ecotoxicology))
# and visual comparison of the fits using Turnbull cdf plot
#
f1n <- fitdistcens(log10LC50, "norm")
summary(f1n)

f1l <- fitdistcens(log10LC50, "logis")
summary(f1l)

cdfcompdens(list(f1n, f1l), legendtext = c("normal", "logistic"),
  xlab = "log10(LC50)", xlim = c(0.5, 2), lines01 = TRUE)

# (4) estimation of the 5 percent quantile value of
# the normal fitted distribution (5 percent hazardous concentration : HC5)
# with its two-sided 95 percent confidence interval calculated by
# non parametric bootstrap
# from a small number of bootstrap iterations to satisfy CRAN running times constraint.
# For practical applications, we recommend to use at least niter=501 or niter=1001.
#
# in log10(LC50)
b1n <- bootdistcens(f1n, niter = 101)
HC51n <- quantile(b1n, probs = 0.05)
# in LC50
10^(HC51n$quantiles)
10^(HC51n$quantCI)

# (5) estimation of the HC5 value
# with its one-sided 95 percent confidence interval (type "greater")
#
# in log10(LC50)
HC51nb <- quantile(b1n, probs = 0.05, CI.type = "greater")

# in LC50
10^(HC51nb$quantiles)
10^(HC51nb$quantCI)

```

smokedfish

Contamination data of Listeria monocytogenes in smoked fish

Description

Contamination data of *Listeria monocytogenes* in smoked fish on the Belgian market in the period 2005 to 2007.

Usage

```
data(smokedfish)
```

Format

smokedfish is a data frame with 2 columns named left and right, describing each observed value of *Listeria monocytogenes* concentration (in CFU/g) as an interval. The left column contains either NA for left censored observations, the left bound of the interval for interval censored observations, or the observed value for non-censored observations. The right column contains either NA for right censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations.

Source

Busschaert, P., Geereard, A.H., Uyttendaele, M., Van Impe, J.F., 2010. Estimating distributions out of qualitative and (semi) quantitative microbiological contamination data for use in risk assessment. *International Journal of Food Microbiology*. **138**, 260-269.

Examples

```
# (1) load of data
#
data(smokedfish)

# (2) plot of data in CFU/g
#
plotdistcens(smokedfish)

# (3) plot of transformed data in log10[CFU/g]
#
Clog10 <- log10(smokedfish)
plotdistcens(Clog10)

# (4) Fit of a normal distribution to data in log10[CFU/g]
#
fitlog10 <- fitdistcens(Clog10, "norm")
summary(fitlog10)
plot(fitlog10)
```

Surv2fitdistcens

Handling of data formatted as in the survival package for use in fitdistcens()

Description

Provide a function to prepare a data frame needed by fitdistcens() from data classically coded when using the Surv() function of the survival package

Usage

```
Surv2fitdistcens(time, time2, event,  
                 type = c('right', 'left', 'interval', 'interval2'))
```

Arguments

time	for right censored data, this is the follow up time. For interval data, the first argument is the starting time for the interval.
event	The status indicator, normally 0=alive, 1=dead. Other choices are TRUE/FALSE (TRUE = death) or 1/2 (2=death). For interval censored data, the status indicator is 0=right censored, 1=event at time, 2=left censored, 3=interval censored. For factor data, assume that it has only two levels with the second level coding death.
time2	ending time of the interval for interval censored. Intervals are assumed to be open on the left and closed on the right, (start, end].
type	character string specifying the type of censoring. Possible values are "right", "left", "interval", "interval2".

Details

Surv2fitdistcens makes a data.frame with two columns respectively named left and right, describing each observed value as an interval as required in fitdistcens(): the left column contains either NA for left-censored observations, the left bound of the interval for interval-censored observations, or the observed value for non-censored observations. The right column contains either NA for right-censored observations, the right bound of the interval for interval censored observations, or the observed value for non-censored observations.

Value

Surv2fitdistcens returns a data.frame with two columns respectively named left and right.

Author(s)

Christophe Dutang and Marie-Laure Delignette-Muller.

References

Delignette-Muller ML and Dutang C (2015), *fitdistrplus: An R Package for Fitting Distributions*. Journal of Statistical Software, 64(4), 1-34, doi:[10.18637/jss.v064.i04](https://doi.org/10.18637/jss.v064.i04).

See Also

See [fitdistrplus](#) for an overview of the package. See [fitdistcens](#) for fitting of univariate distributions to censored data and [fremale](#) for the full dataset used in examples below. See [Surv](#) for survival objects which use the same arguments.

Please visit the [Frequently Asked Questions](#).

Examples

```

# (1) randomized fictive survival data - right-censored
#
origdata <- data.frame(rbind(
  c( 43.01, 55.00, 0),
  c( 36.37, 47.17, 0),
  c( 33.10, 34.51, 0),
  c( 71.00, 81.15, 1),
  c( 80.89, 81.91, 1),
  c( 67.81, 78.48, 1),
  c( 73.98, 76.92, 1),
  c( 53.19, 54.80, 1)))
colnames(origdata) <- c("AgeIn", "AgeOut", "Death")

# add of follow-up time (for type = "right" in Surv())
origdata$followuptime <- origdata$AgeOut - origdata$AgeIn
origdata

### use of default survival type "right"
# in Surv()
survival::Surv(time = origdata$followuptime, event = origdata$Death, type = "right")
# for fitdistcens()
Surv2fitdistcens(origdata$followuptime, event = origdata$Death, type = "right")

# use of survival type "interval"
# in Surv()
survival::Surv(time = origdata$followuptime, time2 = origdata$followuptime,
  event = origdata$Death, type = "interval")
# for fitdistcens()
Surv2fitdistcens(time = origdata$followuptime, time2 = origdata$followuptime,
  event = origdata$Death, type = "interval")

# use of survival type "interval2"
origdata$survivalt1 <- origdata$followuptime
origdata$survivalt2 <- origdata$survivalt1
origdata$survivalt2[1:3] <- Inf
origdata
survival::Surv(time = origdata$survivalt1, time2 = origdata$survivalt2,
  type = "interval2")
Surv2fitdistcens(origdata$survivalt1, time2 = origdata$survivalt2,
  type = "interval2")

# (2) Other examples with various left, right and interval censored values
#
# with left censored data
(d1 <- data.frame(time = c(2, 5, 3, 7), ind = c(0, 1, 1, 1)))
survival::Surv(time = d1$time, event = d1$ind, type = "left")
Surv2fitdistcens(time = d1$time, event = d1$ind, type = "left")

(d1bis <- data.frame(t1 = c(2, 5, 3, 7), t2 = c(2, 5, 3, 7),
  censtype = c(2, 1, 1, 1)))

```

```

survival::Surv(time = d1bis$t1, time2 = d1bis$t2,
  event = d1bis$censtype, type = "interval")
Surv2fitdistcens(time = d1bis$t1, time2 = d1bis$t2,
  event = d1bis$censtype, type = "interval")

# with interval, left and right censored data
(d2 <- data.frame(t1 = c(-Inf, 2, 3, 4, 3, 7), t2 = c(2, 5, 3, 7, 8, Inf)))
survival::Surv(time = d2$t1, time2 = d2$t2, type = "interval2")
Surv2fitdistcens(time = d2$t1, time2 = d2$t2, type = "interval2")

(d2bis <- data.frame(t1 = c(2, 2, 3, 4, 3, 7), t2 = c(2, 5, 3, 7, 8, 7),
  censtype = c(2,3,1,3,3,0)))
survival::Surv(time = d2bis$t1, time2 = d2bis$t2,
  event = d2bis$censtype, type = "interval")
Surv2fitdistcens(time = d2bis$t1, time2 = d2bis$t2,
  event = d2bis$censtype, type = "interval")

```

toxocara

Parasite abundance in insular feral cats

Description

Toxocara cati abundance in feral cats living on Kerguelen island.

Usage

```
data(toxocara)
```

Format

toxocara is a data frame with 1 column (number: number of parasites in digestive tract)

Source

Fromont, E., Morvilliers, L., Artois, M., Pontier, D. 2001. Parasite richness and abundance in insular and mainland feral cats. *Parasitology*, **123**, 143-151.

Examples

```

# (1) load of data
#
data(toxocara)

# (2) description and plot of data
#
number <- toxocara$number
descdist(number, discrete = TRUE, boot = 11)
plotdist(number, discrete = TRUE)

# (3) fit of a Poisson distribution to data

```

```
#
fitp <- fitdist(number, "pois")
summary(fitp)
plot(fitp)

# (4) fit of a negative binomial distribution to data
#
fitnb <- fitdist(number, "nbinom")
summary(fitnb)
plot(fitnb)
```

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