

Package ‘iDINGO’

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Type Package

Title Integrative Differential Network Analysis in Genomics

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Imports mvtnorm, glasso, parallel, GGMridge, visNetwork, scales

Depends igraph

Description Fits covariate dependent partial correlation matrices for integrative models to identify differential networks between two groups. The methods are described in Class et. al., (2018) <[doi:10.1093/bioinformatics/btx750](https://doi.org/10.1093/bioinformatics/btx750)> and Ha et. al., (2015) <[doi:10.1093/bioinformatics/btv4](https://doi.org/10.1093/bioinformatics/btv4)>

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iDINGO-package	<i>iDINGO: Integrative Differential Network Analysis in Genomics</i>
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Description

This packages jointly estimates group specific partial correlations in a multi-level/platform data set.

Details

This packages jointly estimates group specific partial correlations in a multi-level/platform data set, considering the directionality of effects between platforms using a chain graph model.

Author(s)

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brca	<i>Modified TCGA Breast Cancer data</i>
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Description

Modified TCGA Breast Cancer data.

Usage

```
data(brca)
```

Format

Three data frames, with columns as standardized miRNA, gene, and protein expressions. One vector with the two classes of the samples (tumor or normal tissue). Also one iDINGO fit object, obtained by running the example in the idingo manual entry.

dingo

*Fit DINGO model***Description**

This function fit DINGO model and calculates edge-wise differential scores for all pairwise edges among p variables.

Usage

```
dingo(dat,x,rhoarray=NULL,diff.score=T,B=100,verbose=T,cores=1)
```

Arguments

dat	n x p data with colnames as gene name
x	a length n vector representing a binary covariate
rhoarray	a vector representing candidate tuning parameters of glasso for fitting global network model. If it is one value, then we use the value as the tuning parameter. It is set by NULL as default and we select 100 candidate values.
diff.score	a logical value. If TRUE, edge-wise differential scores are calculated from bootstrap standard error. Otherwise, we fit Steps 1 and 2 of DINGO model to get group specific GGMs (partial correlations)
B	the number of bootstrap samples to calculate differential scores.
verbose	if TRUE, lists the procedure
cores	the number of cores to run in parallel for bootstrapping, set to 1 as a default. If more cores are specified than the recommended maximum (the number of cores detected minus 1), this value will be replaced by the recommended value.

Value

genepair	a $p(p-1)/2 \times 2$ matrix indicating all pairs of genes
levels.x	a length 2 vector indicating levels of the binary covariate x , the first element is for group 1 and the second element is for group 2
R1	a length $p(p-1)/2$ vector indicating partial correlations for group 1 and the order is corresponding to the order of genepair
R2	a length $p(p-1)/2$ vector indicating partial correlations for group 2 and the order is corresponding to the order of genepair
boot.diff	a $p(p-1)/2 \times \text{boot.B}$ matrix indicating bootstrapped difference, Fisher's Z transformed $R1 - R2$. The rows are corresponding to the order of gene pair and the columns are corresponding to the bootstrap samples
diff.score	a $p(p-1)/2$ vector of differential score corresponding to genepair
p.val	a $p(p-1)/2$ vector of corrected p-values corresponding to genepair
rho	selected tuning parameter of glasso fit

P	p by p matrix of Global component of the DINGO model
Q	p by 2 matrix of the coefficient parameter of the local group specific component L(x) of the DINGO model.
Psi	p by p diagonal matrix of the noise covariance parameter of the local group specific component L(x) of the DINGO model.
step.times	a length 3 vector containing the elapsed time for Step 1, Step 2, and Bootstrap Scoring, respectively.

Author(s)

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Examples

```
data(gbm)
# Run DINGO (the first column, 'x', contains the group data).
# This may take 5-10 minutes.
## Not run: fit <- dingo(gbm[,-1], gbm$x, diff.score = TRUE, B = 100, cores = 2)
```

extendedBIC

Extended bayesian information criteria for gaussian graphical models

Description

Extended bayesian information criteria for gaussian graphical models

Usage

```
extendedBIC(gamma, omegahat, S, n)
```

Arguments

gamma	a tuning parameter taking a scalar in [0,1] and leading to stronger penalization of large graphs
omegahat	a p x p matrix indicating an estimates of precision (inverse covariance) matrix
S	a p x p matrix indicating sample covariance matrix
n	a scalar indicating sample size

Value

Extended BIC penalized by the size of graphs

Author(s)

Min Jin Ha <mjha@mdanderson.org>

References

Foygel, R. and Drton, M. (2010). Extended bayesian information criteria for gaussian graphical models. arXiv preprint arXiv:1011.6640 .

Examples

```
library(glasso)
data(gbm)
x = gbm[,1]
Y = gbm[,-1]

# Estimating inverse covariance matrix using GLasso #
S = cov(Y)

rhoarray = exp(seq(log(0.001),log(1),length=100))
BIC = rep(0,length(rhoarray))
for (rh in 1:length(rhoarray)) {
  fit.g11 = glasso(S,rho=rhoarray[rh])
  BIC[rh] = extendedBIC(gamma=0,omegahat=fit.g11$wi,S=S,n=nrow(Y))
}
rho = rhoarray[which.min(BIC)]
fit.g12 = glasso(S,rho=rho)
Omega = fit.g12$wi
```

gbm

Modified TCGA Glioblastoma data

Description

Modified TCGA Glioblastoma data.

Usage

```
data(gbm)
```

Format

A data frame with first column as a covariate and other columns as standardized gene expressions.

Description

This function fits the covariance regression model by Hoff and Niu (2012) using EM algorithm with the restriction of diagonal matrix for the noise variance

Usage

```
Greg.em(formula, data = NULL, R = 1, tol = 1e-10, itmax = 1000, verbose = F)
```

Arguments

formula	an object of class "formula" used in model.frame function
data	a data frame used in model.frame function
R	rank of the model
tol	a stopping criterion
itmax	maximum number of iteration
verbose	If true, estimation results for each iteration are printed

Value

A	MLE of the baseline covariance matrix
B	MLE of the regression coefficients

Author(s)

Min Jin Ha <mjha@mdanderson.org>

References

Hoff, P. D. and Niu, X. (2012) A covariance regression model. *Statistica Sinica*, 22, 729-753.

Examples

```
library(glasso)
data(gbm)
x = gbm[,1]
Y = as.matrix(gbm[,-1])
p = ncol(Y)
# Estimating inverse covariance matrix using GLasso #
S = cov(Y)
w.upper = which(upper.tri(S))

rhoarray = exp(seq(log(0.001),log(1),length=100))
BIC = rep(0,length(rhoarray))
```

```

for (rh in 1:length(rhoarray)) {
  fit.g11 = glasso(S,rho=rhoarray[rh])
  BIC[rh] = extendedBIC(gamma=0,omegahat=fit.g11$wi,S=S,n=nrow(Y))
}
rho = rhoarray[which.min(BIC)]
fit.g12 = glasso(S,rho=rho)
Omega = fit.g12$wi

# Fitting (Covariance Regression on transformed data)
diag.Omega = diag(Omega)
P = -Omega/diag.Omega
diag(P) = 0

tY = Y
mdat = apply(tY,2,mean)
sdat = apply(tY,2,sd)
std.tY = t((t(tY) - mdat)/sdat)
smat = diag(sdat)

## rank 1 covariance regression
fit.g = Greg.em(std.tY~x,R=1)

```

idingo

Fit iDINGO model

Description

This function fits the iDINGO model and calculates edge-wise differential scores for all pairwise edges among p variables between multiple platforms.

Usage

```
idingo(dat,dat2=NULL,dat3=NULL,x,plats=NULL,rhoarray=NULL,
diff.score=T,B=100,verbose=T,cores=1)
```

Arguments

dat	nxp dataframe/matrix with colnames as genename
dat2	Second nxp dataframe/matrix with colnames as genename (optional)
dat3	Third nxp dataframe/matrix with colnames as genename (optional)
x	a length n vector representing a binary covariate
plats	a length 1-3 vector (corresponding to the number of data sets submitted, with names for the platforms/levels of the data, such as "microRNA" or "RNAseq". This is optional, and default names "platN" will be used if names are not provided.
rhoarray	a vector representing candidate tuning parameters of glasso for fitting global network model. If it is one value, then we use the value as the tuning parameter. It is set by NULL as default and we select 100 candidate values.

diff.score	a logical value. If TRUE, edge-wise differential scores are calculated from bootstrap standard error. Otherwise, we fit Steps 1 and 2 of DINGO model to get group specific GGMs (partial correlations)
B	the number of bootstrap samples to calculate differential scores.
verbose	if TRUE, lists the procedure
cores	the number of cores to run in parallel for bootstrapping, set to 1 as a default. If more cores are specified than the recommended maximum (the number of cores detected minus 1), this value will be replaced by the recommended value.

Value

genepair	a $p(p-1)/2$ x 2 matrix indicating all pairs of genes
levels.x	a length 2 vector indicating levels of the binary covariate x, the first element is for group 1 and the second element is for group 2
R1	a length $p(p-1)/2$ vector indicating partial correlations for group 1 and the order is corresponding to the order of genepair
R2	a length $p(p-1)/2$ vector indicating partial correlations for group 2 and the order is corresponding to the order of genepair
diff.score	a $p(p-1)/2$ vector of differential score corresponding to genepair
p.val	a $p(p-1)/2$ vector of corrected p-values corresponding to genepair

Author(s)

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Examples

```
data(brca)

# Run iDINGO with microRNA, RNA, and protein data.
# Generally, we recommend a minimum of 100 bootstraps.
## Not run: fit <- idingo(brca$mirna, dat2 = brca$rna, dat3 = brca$prot,
  x = brca$class, plats = c("microRNA", "RNA", "Protein"),
  diff.score = TRUE, B = 20, cores = 2)
## End(Not run)
```

plotNetwork

Plot differential network

Description

This function plots the differential network from a completed DINGO or iDINGO model.

Usage

```
plotNetwork(fit, threshold=0.05, thresh.type="p.val", layout="circular",
  legend.pos="left")
```

Arguments

fit	output from running dingo() or idingo()
threshold	a numeric value containing the threshold for which edges will be included in the differential network plot. If 'thresh.type' is 'p.val', all edges with p-values below this threshold will be included in the plot. If 'thresh.type' is 'diff.score', all edges with absolute differential scores above this threshold will be included in the plot.
thresh.type	either 'p.val' or 'diff.score', defining which variable is used as threshold for edge inclusion.
layout	either 'circular' or one of igraph's supported layouts. If 'circular', dingo networks will be plotted in a circle, and idingo networks will be plotted as a cylinder (with each platform/level as a separate circle).
legend.pos	Legend position for multi-platform networks, in c("left", "right"). Legend is not included for single-platform networks.

Value

visNet	a network plot, using igraph and visNetwork
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Note

To calculate differential scores and p-values for use in network plot thresholding, diff.score must be set to TRUE in dingo() or idingo().

Author(s)

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Examples

```
data(brca)

# Plot the iDINGO result using a p-value threshold of 0.01.
plotNetwork(brca$fit, threshold = 0.01, thresh.type = "p.val")
```

scaledMat	<i>scale a square matrix</i>
-----------	------------------------------

Description

scale a square matrix to have unit diagonal elements.

Usage

```
scaledMat(x)
```

Arguments

x a square matrix with positive diagonal elements

Value

scaled matrix of x

Author(s)

Min Jin Ha mjha@mdanderson.org

scoring.boot

Calculating differential score

Description

This function calculates standard errors for edge-wise partial correlation differences obtained from DINGO model.

Usage

```
scoring.boot(stddat, z, Omega, A, B, boot.B=100, verbose=T)
```

Arguments

stddat standardized nxp data with colnames as genename

z a length n vector representing a binary covariate

Omega a p x p precision matrix for std dat which implies the global network

A p x p matrix of the MLE for the baseline covariance matrix which is obtained from A value of the Greg.em function.

B p x 2 matrix of the MLE for the regression coefficient which is obtained from B value of the Greg.em function

boot.B a scalar indicating the number of bootstraps

verbose if TRUE, lists the bootstrap replications

Value

genepair a $p(p-1)/2 \times 2$ matrix indicating all pairs of genes

levels.z a length 2 vector indicating levels of the binary covariate z, the first element is for group 1 and the second element is for group 2

R1 a length $p(p-1)/2$ vector indicating partial correlations for group 1 and the order is corresponding to the order of genepair

R2 a length $p(p-1)/2$ vector indicating partial correlations for group 2 and the order is corresponding to the order of genepair

boot.diff	a $p(p-1)/2$ x boot.B matrix indicating bootstrapped difference, Fisher's Z transformed R1 - R2. The rows are corresponding to the order of gene pair and the columns are corresponding to the bootstrap samples
diff.score	a $p(p-1)/2$ vector of differential score corresponding to genepair
p.val	a $p(p-1)/2$ vector of corrected p-values corresponding to genepair

Author(s)

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scoring.boot.parallel *Calculating differential score with parallel bootstrap scoring*

Description

This function calculates standard errors for edge-wise partial correlation differences obtained from DINGO model. Bootstrapping is done in parallel using parSapply from the "parallel" library.

Usage

```
scoring.boot.parallel(stddat, z, Omega, A, B, boot.B=100, verbose=T, cores=1)
```

Arguments

stddat	standardized nxp data with colnames as genename
z	a length n vector representing a binary covariate
Omega	a $p \times p$ precision matrix for std dat which implies the global network
A	$p \times p$ matrix of the MLE for the baseline covariance matrix which is obtained from A value of the Greg.em function.
B	$p \times 2$ matrix of the MLE for the regression coefficient which is obtained from B value of the Greg.em function
boot.B	a scalar indicating the number of bootstraps
verbose	if TRUE, lists the bootstrap replications
cores	the number of cores to run in parallel for bootstrapping, set to 1 as a default.

Value

genepair	a $p(p-1)/2 \times 2$ matrix indicating all pairs of genes
levels.z	a length 2 vector indicating levels of the binary covariate z, the first element is for group 1 and the second element is for group 2
R1	a length $p(p-1)/2$ vector indicating partial correlations for group 1 and the order is corresponding to the order of genepair
R2	a length $p(p-1)/2$ vector indicating partial correlations for group 2 and the order is corresponding to the order of genepair

<code>boot.diff</code>	a $p(p-1)/2$ x <code>boot.B</code> matrix indicating bootstrapped difference, Fisher's Z transformed $R1 - R2$. The rows are corresponding to the order of gene pair and the columns are corresponding to the bootstrap samples
<code>diff.score</code>	a $p(p-1)/2$ vector of differential score corresponding to <code>genepair</code>
<code>p.val</code>	a $p(p-1)/2$ vector of corrected p-values corresponding to <code>genepair</code>

Author(s)

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Sigmax

group specific covariance matrices

Description

From parameters of DINGO model, group specific covariance matrices are obtained

Usage

`Sigmax(P = NULL, Q, Psi, x)`

Arguments

<code>P</code>	a $p \times p$ matrix specifying global component
<code>Q</code>	the coefficient parameter matrix of covariance regression model using <code>Greg.em</code> function
<code>Psi</code>	the diagonal error variance matrix of covariance regression model using <code>Greg.em</code> function
<code>x</code>	a vector specifying group. This must be corresponding to the design matrix of <code>Greg.em</code> function

Value

group specific precision matrix

Author(s)

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Examples

```

library(glasso)
data(gbm)
x = gbm[,1]
Y = as.matrix(gbm[,-1])
p = ncol(Y)
# Estimating inverse covariance matrix using GLasso #
S = cov(Y)
w.upper = which(upper.tri(S))

rhoarray = exp(seq(log(0.001),log(1),length=100))
BIC = rep(0,length(rhoarray))
for (rh in 1:length(rhoarray)) {
  fit.g11 = glasso(S,rho=rhoarray[rh])
  BIC[rh] = extendedBIC(gamma=0,omegahat=fit.g11$wi,S=S,n=nrow(Y))
}
rho = rhoarray[which.min(BIC)]
fit.g12 = glasso(S,rho=rho)
Omega = fit.g12$wi

# Fitting (Covariance Regression on transformed data)
diag.Omega = diag(Omega)
P = -Omega/diag.Omega
diag(P) = 0

tY = Y
mdat = apply(tY,2,mean)
sdat = apply(tY,2,sd)
std.tY = t((t(tY) - mdat)/sdat)
smat = diag(sdat)

## rank 1 covariance regression
fit.g = Greg.em(std.tY~x,R=1)
## obtain covariance matrix of Y when x=1
sigmaX1 = Sigmax(Q=fit.g$B,P=P,Psi=fit.g$A,x=c(1,1))

```

single.boot

Calculating differential score for a single bootstrap

Description

This function calculates the edge-wise partial correlation difference for a single bootstrap.

Usage

```
single.boot(i, z, n, tY.org, P, levels.z, w.upper)
```

Arguments

<code>i</code>	iteration number. This is not used within this function, but necessary for <code>parSapply</code> within <code>scoring.boot.parallel</code> function.
<code>z</code>	a length <code>n</code> vector representing a binary covariate
<code>n</code>	the number of rows in data
<code>tY.org</code>	the transformed standardized data
<code>P</code>	the global correlation component
<code>levels.z</code>	the levels of the covariates
<code>w.upper</code>	the upper triangular of Omega

Value

<code>boot.diff</code>	the difference for this bootstrap
------------------------	-----------------------------------

Author(s)

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`trans.Fisher`

Fisher's Z-transformation

Description

Fisher's Z-transformation of (partial) correlation.

Arguments

<code>x</code>	a vector having entries between -1 and 1
----------------	--

Value

Fisher's Z-transformed values

Author(s)

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